

SUPPORTING INFORMATION FOR

Cobalt(III) Werner Complexes with 1,2-Diphenylethylenediamine Ligands:
Readily Available, Inexpensive, and Modular Chiral Hydrogen Bond Donor
Catalysts for Enantioselective Organic Synthesis

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5 EXPERIMENTAL SECTION

General Data. NMR spectra were recorded on a Varian NMRS 500 MHz spectrometer at ambient probe temperatures. Chemical shifts (δ in ppm) were referenced to residual solvent signals (^1H : CHCl_3 , 7.26; $\text{DMSO-}d_5$, 2.50; CHD_2OD , 3.30; CDHCl_2 , 5.32; acetone- d_5 , 2.05; ^{13}C : CDCl_3 , 77.2; $\text{DMSO-}d_6$, 39.5; CD_3OD , 49.0; CD_2Cl_2 , 54.0; acetone- d_6 , 29.8). IR spectra were recorded on a Shimadzu IRAffinity-1 spectrometer (Pike MIRacle ATR system, diamond/ZnSe crystal). UV-visible spectra were obtained with a Shimadzu UV-1800 UV spectrometer. Optical rotations were measured with a Rudolph Autopol II Automatic Polarimeter. Melting points were determined using an OptiMelt MPA 100 instrument. Microanalyses were conducted by Atlantic Microlab. HPLC analyses were carried out with a Shimadzu instrument package (pump/autosampler/detector LC-20AD/SIL-20A/SPD-M20A).

NMR solvents (Cambridge Isotopes) were treated as follows unless noted: $\text{DMSO-}d_6$, distilled under vacuum and stored over molecular sieves; CDCl_3 , CD_2Cl_2 , acetone- d_6 , and CD_3OD , stored over molecular sieves. HPLC solvents (hexanes, Fischer; isopropanol, JT Baker; both HPLC Grade) were degassed before use. Other solvents were treated as follows: DMSO (Fischer, ACS grade), distilled under vacuum and stored over molecular sieves; CH_2Cl_2 (EMD Chemicals, ACS grade), CH_3OH (EMD, anhydrous, 99.8%), hexanes (Macron, ACS grade), EtOAc (Macron, ACS grade), and acetone (BDH, ACS grade), used as received unless noted.

Reactions were conducted under air unless noted. Chemicals were treated as follows. The *trans*- β -nitrostyrene (**4a**, Alfa Aesar, 98%) was column chromatographed (silica gel, 9:1 v/v hexanes/EtOAc). The *trans*-4-methoxy- β -nitrostyrene (**4d**), 3,4-dichloro- β -nitrostyrene (**4e**), and 1-(2-furyl)-2-nitroethylene (**4k**) were used as received from Alfa Aesar (NMR showed **4e,k** to be >98% *trans*). Other previously reported nitroolefins (**4b-c**, **4f-g**, **4j**, **4l**; see Scheme 4) were prepared by Henry reactions of nitromethane and the corresponding aldehydes.^{s1} Dimethyl malonate (**5c**, Alfa Aesar, 98%), diethyl malonate (**5b**, Alfa Aesar, 99%), diisopropyl malonate (**5a**, TCI, 99%), $\text{Co}(\text{OAc})_2 \cdot 4\text{H}_2\text{O}$ (Alfa Aesar, 98%), $\text{Co}(\text{ClO}_4)_2 \cdot 6\text{H}_2\text{O}$ (Alfa Aesar, reagent grade), (*S,S*)-dpen ((*S,S*)-**2**, dpen = 1,2-diphenylethylenediamine, Oakwood or Combi-blocks), (*R,R*)-**2**,

(Oakwood), activated charcoal (Acros, Norit SX 4), HClO₄ (70%, Macron, ACS grade), Ag⁺ BF₄⁻ (Aldrich, 98%), Ag⁺ PF₆⁻ (Strem, 99%), *trans*-2-(2-nitrovinyl)phenol (Aldrich, 97%), di-*t*-butyl azodicarboxylate (**7**, Aldrich, 98%), ethyl 2-oxocyclopentanecarboxylate and 2-oxocyclohexanecarboxylate (**8a,b**, Aldrich, 95%), acetyl chloride (Aldrich, 98%), benzoyl chloride (Aldrich, 99%), Et₃N (Alfa Aesar, 98%), *N*-methyl morpholine (Alfa Aesar, 99%), DOWEX 50WX2 (Aldrich), silica gel (Silicycle SiliaFlash® F60), Celite 545 (Aldrich), and Na₂SO₄ (EMD) were used as received. The Na⁺ BAr_f⁻ was prepared by a literature procedure (BAr_f⁻ = B(3,5-C₆H₃-(CF₃)₂)₄⁻).^{s2}

Λ -[Co((*S,S*)-dp_{en})₃]³⁺ 3Cl⁻ (Λ -(*S,S*)-**3**³⁺ 3Cl⁻; Scheme 1, step A).^{s3-s5} A gas circulating flask was charged with a solution of Co(OAc)₂·4H₂O (0.296 g, 1.19 mmol) in CH₃OH (50 mL). Activated charcoal (0.1 g) and (*S,S*)-**2** (0.849 g, 4.00 mmol, 3.36 equiv) were added with stirring. Air was passed through the vigorously stirred suspension. After 17 h, the mixture was filtered through Celite and aqueous HCl (1.5 mL, 12.0 M) was added. The solvent was removed by rotary evaporation to give an orange solid. A portion was dissolved in CD₃OD. A ¹³C{¹H} NMR spectrum showed CHPh signals at δ 63.5, 66.1, and 66.7 ppm (67/10/23), which were attributed to the Λ and Δ diastereomers of (*S,S*)-**3**³⁺ 3Cl⁻, and *trans*-[Co((*S,S*)-dp_{en})₂(Cl)₂]⁺ Cl⁻,^{s6} respectively.^{s7} Then H₂O (100 mL) was added. The suspension was filtered and the orange filter cake was washed with H₂O (100 mL). The cake was dissolved in CH₃OH. The solvent was removed by rotary evaporation and the residue dried by oil pump vacuum (room temperature, 14 h) to give Λ -(*S,S*)-**3**³⁺ 3Cl⁻·3H₂O as an orange solid (0.662 g, 0.773 mmol, 65%), mp 220-230 °C dec (green liquid; open capillary). The ¹³C{¹H} spectrum showed only one CHPh signal, indicative of the Λ diastereomer, as confirmed crystallographically (below). Anal. Calcd. for C₄₂H₄₈Cl₃CoN₆·3H₂O (856.21): C 58.92, H 6.36, N 9.82; found C 58.95, H 6.46, N 9.68.

NMR (10:1 v/v DMSO-*d*₆/CD₃OD, δ in ppm):^{s8} ¹H (500 MHz) 7.65-7.42 (m, 12H, *o*-Ph), 7.41-7.15 (m, 18H, *m*-, *p*-Ph), 6.86 (br s, 6H, NHH'), 5.31 (br s, 6H, NHH'), 4.97 (s, 6H, CHNH₂), 4.06 (br s, 10H, CD₃OH/H₂O);^{s9-s10} ¹³C{¹H} (125 MHz) 137.3 (s, *i*-Ph), 128.9 and 128.8 (2 s, *o*-, *m*-Ph), 128.6 (s, *p*-Ph), 61.8 (s, CHNH₂).

Δ -[Co((*S,S*)-dpen)₃]³⁺ 3ClO₄⁻ (Δ -(*S,S*)-**3**³⁺ 3ClO₄⁻; Scheme 1, step B).^{s4a,s11} This synthesis involves the use and preparation of perchlorate salts that are potentially explosive.^{s12} Although no mishaps occurred, special care and protection are always required when working with perchlorate salts or HClO₄ solutions. A gas circulating flask was charged with a solution of Co(ClO₄)₂·6H₂O (0.261 g, 0.711 mmol) in CH₃OH (50 mL). Activated charcoal (0.1 g) and (*S,S*)-**2** (0.505 g, 2.38 mmol, 3.31 equiv) were added with stirring. Air was passed through the vigorously stirred suspension. After 17 h, the mixture was filtered through Celite and HClO₄ (2.0 mL, 35% in H₂O) was added. The solvent was removed by rotary evaporation to give an orange solid. A portion was dissolved in CD₃OD, and a ¹³C{¹H} NMR spectrum showed CHPh signals at δ 63.6 and 61.8 ppm (92/8). These were attributed to the Δ and Λ diastereomers of (*S,S*)-**3**³⁺ 3ClO₄⁻. The solid was washed with water and dissolved in CH₃OH. The solvent was removed by rotary evaporation to give crude (*S,S*)-**3**³⁺ 3ClO₄⁻ as an orange powder (0.422 g, 0.425 mmol, 60%) and an 83:17 mixture of Δ / Λ isomers,^{s11} as assayed by ¹³C{¹H} NMR (δ , DMSO-*d*₆, 63.2/61.4 ppm).^{s13} Diastereomerically pure Δ -(*S,S*)-**3**³⁺ 3ClO₄⁻ was obtained per Scheme 1, step D (below).

Data for Δ -(*S,S*)-**3**³⁺ 3ClO₄⁻. NMR (CD₃OD, δ in ppm):^{s8} ¹H (500 MHz) 7.41-7.37 (m, 12H, *o*-Ph), 7.34-7.30 (m, 18H, *m*-, *p*-Ph), 4.89 (br s, 26H, CD₃OH/H₂O),^{s9} 4.52 (s, 6H, CH-NH₂);^{s15} ¹³C{¹H} (125 MHz): 136.5 (s, *p*-Ph), 130.4 (s, *i*-Ph), 130.0 and 129.0 (2 s, *o*-, *m*-Ph),^{s16} 66.5 (s, CHNH₂).

Λ -[Co((*S,S*)-dpen)₃]³⁺ 3ClO₄⁻ (Λ -(*S,S*)-**3**³⁺ 3ClO₄⁻; Scheme 1, step C).^{s4a,s14} A beaker was charged with Λ -(*S,S*)-**3**³⁺ 3Cl⁻·3H₂O (0.502 g, 0.586 mmol) and boiling CH₃OH (200 mL). Then NaClO₄ (ca. 0.765 g, 6.25 mmol) was added with stirring, followed by warm water (400 mL). The solution was kept at 0 °C. After 7 h, the yellow precipitate was collected, washed with water, dried by oil pump vacuum at room temperature, and dissolved in a minimum of hot acetone. Ten volumes of ethanol were added. Ether was slowly added, initiating precipitation. More ether was added, and the mixture was kept at 0 °C. After 14 h, the orange solid was collected by filtration and dried by oil pump vacuum at room temperature to give Λ -(*S,S*)-**3**³⁺ 3Cl-

$\text{O}_4^- \cdot 4\text{H}_2\text{O}$ (0.401 g, 0.376 mmol, 64%).^{s4a,s14}

NMR (CD_3OD , δ in ppm):^{s8} ^1H (500 MHz) 7.41 (br s, 12H, *o*-Ph), 7.35 (br s, 18H, *m*-, *p*-Ph), 4.88 (br s, 24H, $\text{CD}_3\text{OH}/\text{H}_2\text{O}$),^{s9} 4.81 (s, 6H, CHNH_2);^{s15} $^{13}\text{C}\{^1\text{H}\}$ (125 MHz): 136.4 (s, *i*-Ph), 130.5 (s, *p*-Ph), 130.3 and 129.7 (2 s, *o*-, *m*-Ph),^{s16} 63.7 (s, CHNH_2).

$\Delta\text{-}[\text{Co}((S,S)\text{-dpen})_3]^{3+} 3\text{Cl}^-$ ($\Delta\text{-}(S,S)\text{-}\mathbf{3}^{3+} 3\text{Cl}^-$). **A (Scheme 1, step E with diastereomer mixture)**. A beaker was charged with the 83:17 mixture of Δ/Λ diastereomers of $(S,S)\text{-}\mathbf{3}^{3+} 3\text{ClO}_4^-$ obtained above (0.302 g, 0.304 mmol) and CH_3OH (50 mL). The solution was sorbed onto a Dowex cation exchange column (3.0×12 cm), giving an orange band. The column was washed with $\text{H}_2\text{O}/\text{CH}_3\text{OH}$ (1:1 v/v, 100 mL) and eluted with increasing concentrations of 12.0 M aq. HCl in CH_3OH (9:1 v/v $\text{CH}_3\text{OH}/\text{HCl}$ (100 mL), 8:2 v/v $\text{CH}_3\text{OH}/\text{HCl}$ (100 mL), 7:3 v/v $\text{CH}_3\text{OH}/\text{HCl}$ (100 mL), 6:4 v/v $\text{CH}_3\text{OH}/\text{HCl}$ (until product elution)). An orange band was collected and the solvents were removed by rotary evaporation (45 °C bath; base trap to scavenge HCl). This gave $(S,S)\text{-}\mathbf{3}^{3+} 3\text{Cl}^-$ (0.227 g, 0.283 mmol, 93%) as a bright orange solid and a 83:17 mixture of Δ/Λ isomers, as assayed by $^{13}\text{C}\{^1\text{H}\}$ NMR (δ , 63.7/62.0 ppm).^{s13} **B (Scheme 1, step E with Δ diastereomer)**. A beaker was charged with $\Delta\text{-}(S,S)\text{-}\mathbf{3}^{3+} 3\text{ClO}_4^-$ (0.200 g, 0.202 mmol; see below) and CH_3OH (50 mL). The solution was sorbed onto a Dowex cation exchange column. A workup identical to that in A gave $\Delta\text{-}(S,S)\text{-}\mathbf{3}^{3+} 3\text{Cl}^- \cdot 2\text{H}_2\text{O}$ (0.0139 g, 0.0166 mmol, 82%) as a bright orange solid, mp 193-197 °C dec (green liquid; open capillary). Anal. Calcd. for $\text{C}_{42}\text{H}_{48}\text{Cl}_3\text{CoN}_6 \cdot 2\text{H}_2\text{O}$ (838.19): C 60.18, H 6.25, N 10.03; found C 60.23, H 6.24, N 9.90.

Data for product from **A**. NMR (3:1 v/v $\text{DMSO}-d_6/\text{CD}_3\text{OD}$, δ in ppm):^{s8} ^1H (500 MHz) 7.54-7.49 (m, 0.17·12H, *o*-Ph), 7.48-7.43 (m, 0.83·12H, *o*-Ph), 7.34-7.22 (m, 0.17·18H and 0.83·18H, *m*-, *p*-Ph), 6.77 (br s, 0.17·6H, NHH'), 6.53 (d, $^3J_{\text{HH}} = 11$ Hz, 0.83·6H, NHH'), 5.43 (br s, 0.83·6H, NHH'), 5.32 (br s, 0.17·6H, NHH'), 4.96 (s, 0.17·6H, CHNH_2), 4.77-4.62 (m, 0.83·6H, CHNH_2), 4.06 (br s, 20H, $\text{CD}_3\text{OH}/\text{H}_2\text{O}$),^{s9} $^{13}\text{C}\{^1\text{H}\}$ (125 MHz) 137.6 (s, *i*-Ph), 129.2 (s, Ph),^{s17} 129.0 (s, Ph), 128.9 (s, Ph), 128.6 (s, Ph), 63.7 (s, CHNH_2 , 83%), 62.0 (s, CHNH_2 , 17%).

Data for product from **B**. NMR (CD_3OD , δ in ppm):^{s8} ^1H (500 MHz) 7.46-7.44 (m, 12H,

o), 7.30 (br s, 18H, *m*, *p*), 6.41 (d, 6H, $^3J_{\text{HH}} = 10$ Hz, NHH'), 5.94 (bs s, 6H, NHH'), 4.88 (br s, 13H, CD₃OH/H₂O),^{s9} 4.58 (d, 6H, $^3J_{\text{HH}} = 10$ Hz, CHNH₂); $^{13}\text{C}\{^1\text{H}\}$ (125 MHz): 137.3 (s, *i*-Ph), 130.2 (s, *p*-Ph), 130.0 and 129.1 (2 s, *o*-, *m*-Ph), 66.7 (s, CHNH₂).

Equilibration of diastereomers of (*S,S*)-3**³⁺ **3X**⁻. A (Scheme 1, step D with Λ diastereomer).**^{s18} A 5 mm NMR tube was charged with Λ -(*S,S*)-**3**³⁺ 3ClO₄⁻·4H₂O (0.027 g, 0.025 mmol), activated charcoal (0.005 g), CD₃OD (0.5 mL), and a stir bar, and kept at 70 °C with stirring. The sample was monitored by $^{13}\text{C}\{^1\text{H}\}$ NMR (stir bar removed prior to each spectrum), which showed the isomerization of Λ -(*S,S*)-**3**³⁺ 3ClO₄⁻ to Δ -(*S,S*)-**3**³⁺ 3ClO₄⁻ (δ 63.6 ppm to 66.6 ppm), with a minor byproduct (δ 71.9 ppm) becoming increasingly visible after 24 h. Data (time, ratios): 4 h, 25/75/-, 22 h, 14/86/-, 68 h, 10/76/14, 150 h, 3/81/16). No other signals were evident. **B. (Scheme 1, step D with Δ diastereomer dominant).** A round bottom flask was charged with the 83:17 Δ/Λ mixture of (*S,S*)-**3**³⁺ 3ClO₄⁻ (above; 0.118 g, 0.119 mmol), CH₃OH (10 mL), and activated charcoal (0.051 g), and fitted with a condenser. The mixture was heated at 70 °C. After 48 h, the mixture was filtered through Celite. The solvent was removed by rotary evaporation and dried by oil pump vacuum at room temperature to give Δ -(*S,S*)-**3**³⁺ 3ClO₄⁻ (0.114 g, 0.115 mmol, 97%; triplicate runs gave 95% and 99%; <4% Λ)^{s11b} as an orange solid, NMR data for which are given above. **C (Scheme 1, step F with diastereomer mixture).** A 5 mm NMR tube was charged with (*S,S*)-**3**³⁺ 3Cl⁻ (0.030 g, 0.0373 mmol; 17:83 Λ/Δ), activated charcoal (0.0063 g), CD₃OD (0.5 mL), and a stir bar, and kept at 70 °C with stirring. The sample was monitored by $^{13}\text{C}\{^1\text{H}\}$ NMR (stir bar removed prior to each spectrum), which showed the gradual isomerization to a 61:39 Λ/Δ mixture (δ 66.6 ppm to 63.5 ppm). Data (time, ratios): 18 h, 58/42, 37 h, 61/39). No other signals were evident. **D (Scheme 1, step F with Δ diastereomer).** A 5 mm NMR tube was charged with Δ -(*S,S*)-**3**³⁺ 3Cl⁻ (0.025 g, 0.0311 mmol), activated charcoal (0.0010 g), CD₃OD (0.5 mL), and a stir bar, and kept at 70 °C with stirring. The sample was monitored by $^{13}\text{C}\{^1\text{H}\}$ NMR (stir bar removed prior to each spectrum), which showed isomerization to a 57:43 Λ/Δ mixture (δ 66.6 ppm to 63.5 ppm). Data (time, ratios: 24 h, 54/46; 50 h, 57/43). No other signals were evident.

Λ -[Co((*S,S*)-dpen)₃]³⁺ 2Cl[−]BAr_f[−] (Λ -(*S,S*)-**3**³⁺ 2Cl[−]BAr_f[−]). A round bottom flask was charged with Λ -(*S,S*)-**3**³⁺ 3Cl[−]·3H₂O (0.320 g, 0.374 mmol), CH₂Cl₂ (15 mL) and Na⁺ BAr_f[−] (0.382 g, 0.431 mmol). The sample was sonicated for 5 min, during which time the solvent became bright orange and a white precipitate (NaCl) formed. The mixture was filtered. The filtrate was loaded onto a silica gel column (1.9 × 15 cm), giving an orange band. The column was eluted with CH₂Cl₂/CH₃OH (100:0 to 98:2 v/v). The main orange band was collected. The solvent was removed by rotary evaporation and the residue dried by oil pump vacuum (room temperature, 14 h) to give Λ -(*S,S*)-**3**³⁺ 2Cl[−]BAr_f[−]·2H₂O (0.5802 g, 0.3478 mmol, 93%) as an orange solid, mp 116-119 °C dec (open capillary). Anal. Calcd. for C₇₄H₆₀BCl₂CoF₂₄N₆·2H₂O (1665.95): C 53.35, H 3.87, N 5.04, Cl 4.26; found C 53.50, H 3.82, N 4.84, Cl 3.94.

NMR (CD₂Cl₂, δ in ppm): ¹H (500 MHz) BAr_f[−] at 7.73 (s, 8H, *o*), 7.54 (s, 4H, *p*); (*S,S*)-**2** ligand at 8.17 (br s, 6H, NHH'), 7.40-7.37 (m, 6H, *p*-Ph), 7.34-7.30 (m, 24H, *o*-, *m*-Ph), 4.48 (s, 6H, CHNH₂), 3.86 (br s, 6H, NHH'), 2.14 (br s, 7H, H₂O);^{s9} ¹³C {¹H} (125 MHz) BAr_f[−] at 162.3 (q, ¹J_{BC} = 49.4 Hz, *i*), 135.4 (s, *o*), 129.4 (q, ²J_{CF} = 31.3 Hz, *m*), 125.2 (q, ¹J_{CF} = 271.0 Hz, CF₃), 118.1 (s, *p*); (*S,S*)-**2** ligand at 134.8 (s, *i*-Ph), 131.1 (s, *p*-Ph), 130.6 (s, *o*-Ph), 128.0 (s, *m*-Ph);^{s16} 63.6 (s, CHNH₂). IR (powder film, cm^{−1}): 3039 (m, ν_{NH}), 1609 (m, δ_{NH}), 1354 (s, $\nu_{\text{Ar-CF}_3}$), 1275 (vs, ν_{CF}), 1119 (vs, δ_{CCN}); UV-visible (nm, 1.66 × 10^{−3} M in CH₂Cl₂ (ϵ , M^{−1} cm^{−1})): 455 (shoulder, 157). [α]₁₈⁵⁸⁹ = 0.0° (5.18 mg mL^{−1}, CH₂Cl₂); [α]₁₈⁵⁴⁶ = 23.1° (5.18 mg mL^{−1}, CH₂Cl₂).^{s19}

Δ -[Co((*S,S*)-dpen)₃]³⁺ 2Cl[−]BAr_f[−] (Δ -(*S,S*)-**3**³⁺ 2Cl[−]BAr_f[−]). A round bottom flask was charged with a 83:17 mixture of Δ -/ Λ -(*S,S*)-**3**³⁺ 3Cl[−] (0.188 g, 0.234 mmol), CH₂Cl₂ (10 mL), Na⁺ BAr_f[−] (0.240 g, 0.270 mmol), and CH₃OH (0.10 mL). The sample was sonicated for 5 min, during which time the solvent became bright orange as a small amount of a white solid (NaCl) precipitated. The mixture was filtered. The filtrate was loaded onto a silica gel column (1.9 × 14 cm), giving an orange band. The column was eluted with CH₂Cl₂/CH₃OH (100:0 to 97:3 v/v). Aliquots of the main orange band were collected. These were analyzed by TLC, and combined accordingly. The solvents were removed by rotary evaporation and the residues dried by oil pump

vacuum to give Λ -(*S,S*)-**3**³⁺ 2Cl[−]BAr_F[−]·2H₂O (0.0586 g, 0.0352 mmol, 15%) and Λ -(*S,S*)-**3**³⁺ 2Cl[−]BAr_F[−]·H₂O (0.2932 g, 0.1779 mmol, 76%).

Data for Λ -(*S,S*)-**3**³⁺ 2Cl[−]BAr_F[−]·H₂O. When warmed (open capillary), the complex continually darkened and liquefied at 110 °C. Anal. Calcd. for C₇₄H₆₀BCl₂CoF₂₄N₆·H₂O (1647.94): C 53.93, H 3.79, N 5.10, Cl 4.30; found C 53.96, H 4.02, N 4.98, Cl 4.20.

NMR (CD₂Cl₂/CH₃OH (0.40:0.001 mL, with the latter significantly enhancing solubility), δ in ppm): ¹H (500 MHz) BAr_F[−] at 7.73 (s, 8H, *o*), 7.55 (s, 4H, *p*); (*S,S*)-**2** ligand at 7.30-7.27 (m, 18H, *o*-, *p*-Ph), 7.22-7.19 (t, ³J_{HH} = 7.5 Hz, 12H, *m*-Ph), 6.58 (br s, 6H, NHH'), 5.35 (br s, 6H, NHH'), 4.25 (s, 6H, CHNH₂), 2.53 (br s, 14H, CH₃OH/H₂O); ¹³C {¹H} (125 MHz) BAr_F[−] at 162.3 (q, ¹J_{BC} = 49.6 Hz, *i*), 135.4 (s, *o*), 129.4 (q, ²J_{CF} = 31.4 Hz, *m*), 125.2 (q, ¹J_{CF} = 271.0 Hz, CF₃), 118.0 (s, *p*); (*S,S*)-**2** ligand at 134.3 (s, *i*-Ph), 130.6 (s, *p*-Ph), 129.9 (s, *o*-Ph), 128.0 (s, *m*-Ph), ¹⁶ 66.7 (s, CHNH₂). IR (powder film, cm^{−1}): 3068 (m, ν_{NH}), 1608 (m, δ_{NH}), 1574 (m, δ_{NH}), 1354 (s, $\nu_{\text{Ar-CF}_3}$), 1275 (vs, ν_{CF}), 1119 (vs, δ_{CCN}); UV-visible (nm, 1.66 × 10^{−3} M in CH₂Cl₂ (ϵ , M^{−1} cm^{−1})): 462 (191); [α]₁₈⁵⁴⁶ = −419.0° (5.25 mg mL^{−1}, CH₂Cl₂).

Λ -[Co((*S,S*)-dpen)₃]³⁺ 3BAr_F[−] (Λ -(*S,S*)-**3**³⁺ 3BAr_F[−]). **A.** A beaker was charged with Λ -(*S,S*)-**3**³⁺ 2Cl[−]BAr_F[−]·2H₂O (0.170 g, 0.102 mmol), CH₂Cl₂ (15 mL), Na⁺ BAr_F[−] (0.188 g, 0.204 mmol, 2.00 equiv), and H₂O (15 mL). The biphasic mixture was vigorously stirred for 10 min and then allowed to stand. The lower orange CH₂Cl₂ phase was separated and allowed to evaporate in a hood to give Λ -(*S,S*)-**3**³⁺ 3BAr_F[−]·8H₂O as an orange solid (0.336 g, 0.0979 mmol, 96%). **B.** A beaker was charged with Λ -(*S,S*)-**3**³⁺ 3Cl[−]·3H₂O (0.170 g, 0.199 mmol), CH₂Cl₂ (15 mL), Na⁺ BAr_F[−] (0.532 g, 0.60 mmol, 3.0 equiv), and H₂O (15 mL). The biphasic mixture was vigorously stirred for 10 min, and then allowed to stand. The lower orange CH₂Cl₂ phase was separated and allowed to evaporate in a hood to give Λ -(*S,S*)-**3**³⁺ 3BAr_F[−]·8H₂O as an orange solid (0.644 g, 0.188 mmol, 94%), dec pt 68-98 °C (no melting), liquefies at 106 °C (open capillary). Anal. Calcd. for C₁₃₈H₈₄B₃CoF₇₂N₆·8H₂O (3429.55): C 48.33, H 2.94, N 2.45; found C 48.16, H 2.90, N 2.40.

NMR (acetone-*d*₆, δ in ppm): ¹H (500 MHz) BAr_F[−] at 7.79 (s, 24H, *o*), 7.67 (s, 12H, *p*);

(*S,S*)-**2** ligand at 7.40-7.37 (m, 6H, *p*-Ph), 7.34-7.30 (m, 24H, *o*-, *m*-Ph), 6.38 (br s, 6H, NHH'), 6.22 (br s, 6H, NHH'); 5.30 (s, 6H, CHNH₂), 2.95 (br s, 24H, H₂O),^{s9} ¹³C{¹H} (125 MHz) BAr_f⁻ at 162.6 (q, ¹J_{BC} = 50.0 Hz, *i*), 135.5 (s, *o*), 130.0 (q, ²J_{CF} = 32.5 Hz, *m*), 125.3 (q, ¹J_{CF} = 270.0 Hz, CF₃), 118.4 (m, *p*); (*S,S*)-**2** ligand at 135.5 (s, *i*-Ph), 130.8 (s, *p*-Ph), 130.3 (s, *o*-Ph), 129.4 (s, *m*-Ph),^{s16} 64.1 (s, CHNH₂).

Δ -[Co((*S,S*)-dpen)₃]³⁺ 3BAr_f⁻ (Δ -(*S,S*)-**3**³⁺ 3BAr_f⁻). A beaker was charged with Δ -(*S,S*)-**3**³⁺ 2Cl-BAr_f⁻·H₂O (0.0824 g, 0.050 mmol), Na⁺ BAr_f⁻ (0.0923 g, 0.100 mmol, 2.0 equiv), CH₂Cl₂ (10 mL), and H₂O (10 mL). The biphasic mixture was vigorously stirred for 10 min and then allowed to stand. The lower orange CH₂Cl₂ phase was separated and allowed to evaporate in the hood to give Δ -(*S,S*)-**3**³⁺ 3BAr_f⁻·9H₂O as an orange solid (0.158 g, 0.0458 mmol, 92%), which when warmed (open capillary) continually darkened and then liquefied at 90 °C. Anal. Calcd. for C₁₃₈H₈₄B₃CoF₇₂N₆·9H₂O (3447.57): C 48.08, H 2.98, N 2.44; found C 48.04, H 2.78, N 2.46.

NMR (acetone-*d*₆, δ in ppm):^{s8} ¹H (500 MHz) BAr_f⁻ at 7.80 (s, 24H, *o*), 7.67 (s, 12H, *p*); (*S,S*)-**2** ligand at 7.51-7.45 (m, 12H, *o*-Ph), 7.43-7.31 (m, 18H, *m*-, *p*-Ph), 6.87-6.70 (m, 6H, NHH'), 5.93 (br s, 6H, NHH'); 5.31-5.21 (m, 6H, CHNH₂), 3.01 (br s, 20H, H₂O),^{s9} ¹³C{¹H} (125 MHz) BAr_f⁻ at 162.6 (q, ¹J_{BC} = 50.0 Hz, *i*), 135.5 (s, *o*), 130.0 (m, *m*), 125.3 (q, ¹J_{CF} = 270.0 Hz, CF₃), 118.4 (m, *p*); (*S,S*)-**2** ligand at 135.8 (s, *i*-Ph), 130.9 (s, *p*-Ph), 130.3 (s, *o*-Ph), 128.6 (s, *m*-Ph),^{s16} 65.9 (s, CHNH₂).

Λ -[Co((*S,S*)-dpen)₃]³⁺ 3BF₄⁻ (Λ -(*S,S*)-**3**³⁺ 3BF₄⁻). A round bottom flask was charged with a solution of Λ -(*S,S*)-**3**³⁺ 3Cl·3H₂O (0.173 g, 0.202 mmol) in CH₃OH (15 mL) and Ag⁺ BF₄⁻ (0.118 g, 0.606 mmol, 3.0 equiv). The mixture was sonicated for 10 min, and filtered through Celite. The solvent was removed from the filtrate by rotary evaporation. The residue was dried by oil pump vacuum (room temperature, 14 h) to give Λ -(*S,S*)-**3**³⁺ 3BF₄⁻·2H₂O as a bright orange solid (0.185 g, 0.187 mmol, 93%), mp 198-202 °C dec (open capillary). Anal. Calcd. for C₄₂H₄₈B₃CoF₁₂N₆·2H₂O (992.24): C 50.84, H 5.28, N 8.47; found C 51.16, H 5.42, N 8.47.

¹H NMR (500 MHz, acetone-*d*₆, δ in ppm):^{s8} 7.57-7.43 (m, 12H, *o*-Ph), 7.42-7.23 (m,

18H, *m*-, *p*-Ph), 5.83 (br s, 11H, NHH'), ^{s15} 5.18 (s, 6H, CHNH₂), 2.87 (s, 7H, H₂O); ^{s9} ¹³C {¹H} NMR (125 MHz, CD₃OD, δ in ppm): 136.1 (s, *i*-Ph), 130.5 (s, *p*-Ph), 130.2, (s, *o*-Ph), 129.6 (s, *m*-Ph), ^{s16} 63.7 (s, CHNH₂).

Λ-[Co((*S,S*)-dpen)₃]³⁺ 2BF₄⁻BAr_f⁻ (Λ-(*S,S*)-3**³⁺ 2BF₄⁻BAr_f⁻).** A round bottom flask was charged with a solution of Λ-(*S,S*)-**3**³⁺ 2Cl⁻BAr_f⁻·2H₂O (0.9985 g, 0.599 mmol) in CH₂Cl₂ (4 mL), and Ag⁺ BF₄⁻ (0.2412 g, 1.24 mmol, 2.07 equiv) was added with stirring. The heterogeneous mixture was sonicated for 10 min. The mixture was filtered through Celite. The solvent was removed from the filtrate by rotary evaporation. The residue was dried by oil pump vacuum (room temperature, 14 h) to give Λ-(*S,S*)-**3**³⁺ 2BF₄⁻BAr_f⁻·3H₂O as a bright orange solid (1.019 g, 0.570 mmol, 95%), mp 140-147 °C dec (open capillary). Anal. Calcd. for C₇₄H₆₀B₃CoF₃₂N₆·3H₂O (1786.67): C 49.75, H 3.72, N 4.70; found C 49.94, H 3.69, N 4.52.

NMR (CD₂Cl₂, δ in ppm): ¹H (500 MHz) BAr_f⁻ at 7.73 (s, 8H, *o*), 7.56 (s, 4H, *p*); (*S,S*)-**2** ligand at 7.45 (t, ¹J_{HH} = 7.0 Hz, 6H, *p*-Ph), 7.40 (t, ¹J_{HH} = 7.0 Hz, 12H, *m*-Ph), 7.32 (d, ¹J_{HH} = 7.0 Hz, 12H, *o*-Ph), 6.04 (br s, 6H, NHH'), 4.52 (s, 6H, CHNH₂), 4.17 (br s, 6H, NHH'), 2.01 (s, 8H, H₂O); ^{s9} ¹³C {¹H} (125 MHz) BAr_f⁻ at 162.2 (q, ¹J_{BC} = 48.8 Hz, *i*), 135.3 (s, *o*), 129.4 (q, ²J_{CF} = 27.5 Hz, *m*), 125.1 (q, ¹J_{CF} = 271.3 Hz, CF₃), 118.1 (s, *p*); (*S,S*)-**2** ligand at 133.6 (s, *p*-Ph), 131.4 (s, *i*-Ph), 130.7, (s, *o*-Ph), 127.8 (s, *m*-Ph), ^{s16} 64.5 (s, CHNH₂). IR (powder film, cm⁻¹): 3254 (m, ν_{NH}), 1608 (m, δ_{NH}), 1354 (s, ν_{Ar-CF₃}), 1277 (vs, ν_{CF}), 1121 (vs, δ_{CCN}); UV-visible (nm, 1.67 × 10⁻³ M in CH₂Cl₂ (ε, M⁻¹ cm⁻¹)): 451 (shoulder, 231); [α]₁₈⁵⁴⁶ = 50.6° (4.74 mg mL⁻¹, CH₂Cl₂).

Λ-[Co((*S,S*)-dpen)₃]³⁺ 2PF₆⁻BAr_f⁻ (Λ-(*S,S*)-3**³⁺ 2PF₆⁻BAr_f⁻).** A round bottom flask was charged with a solution of Λ-(*S,S*)-**3**³⁺ 2Cl⁻BAr_f⁻·2H₂O (0.1200 g, 0.072 mmol) in CH₂Cl₂ (2 mL). A solution of Ag⁺ PF₆⁻ (0.0373 g, 0.148 mmol, 2.1 equiv) in CH₂Cl₂ (2 mL) was added with stirring. A white precipitate immediately formed. The mixture was sonicated (1 min) and filtered through Celite. The orange filtrate was washed with H₂O (2 × 1 mL) and the solvent was removed by rotary evaporation. The residue was dried by oil pump vacuum (room temperature, 5 h) to give Λ-(*S,S*)-**3**³⁺ 2PF₆⁻BAr_f⁻·H₂O as a bright orange solid (0.1121 g, 0.060 mmol, 83%),

which when warmed (open capillary) continually darkened and then liquefied at 160 °C. Anal. Calcd. for $C_{74}H_{60}BCoF_{36}N_6P_2 \cdot H_2O$ (1866.96): C 47.61, H 3.35, N 4.50; found C 47.86, 3.49, 4.62.

NMR (CD_2Cl_2 , δ in ppm): 1H (500 MHz) $BArF^-$ at 7.71 (s, 8H, *o*), 7.55 (s, 4H, *p*); (*S,S*)-**2** ligand at 7.47 (t, $^3J_{HH} = 7.0$ Hz, 6H, *p*-Ph), 7.42 (t, $^3J_{HH} = 7.5$ Hz, 12H, *m*-Ph), 7.29 (d, $^3J_{HH} = 7.0$ Hz, 12H, *o*-Ph), 5.66 (br s, 6H, NHH'), 4.49 (s, 6H, $CHNH_2$), 4.37 (br s, 6H, NHH'), 1.85 (br s, 18H, H_2O),^{s9} $^{13}C\{^1H\}$ (125 MHz) $BArF^-$ at 162.2 (q, $^1J_{BC} = 48.8$ Hz, *i*), 135.3 (s, *o*), 129.4 (q, $^2J_{CF} = 33.8$ Hz, *m*), 125.1 (q, $^1J_{CF} = 271.2$ Hz, CF_3), 118.1 (s, *p*); (*S,S*)-**2** ligand at 133.0 (s, *i*-Ph), 131.9 (s, *p*-Ph), 131.0 (s, *o*-Ph), 127.8 (s, *m*-Ph),^{s16} 64.0 (s, $CHNH_2$). IR (powder film, cm^{-1}): 3287 (m, ν_{NH}), 1609 (m, δ_{NH}), 1354 (s, ν_{Ar-CF_3}), 1276 (vs, ν_{CF}), 1122 (vs, δ_{CCN}); UV-visible (nm, 1.67×10^{-3} M in CH_2Cl_2 (ϵ , $M^{-1} cm^{-1}$)): 449 (shoulder, 302); $[\alpha]_{18}^{546} = 47.6^\circ$ (4.42 mg mL^{-1} , CH_2Cl_2).

Representative 1H NMR titration. A 5 mm NMR tube was charged with Λ -(*S,S*)-**3**³⁺ $2Cl^-BArF^- \cdot 2H_2O$ (0.0141 g, 0.00848 mmol) and CD_2Cl_2 (0.5 mL). A solution of **4a** (2.1 M in CD_2Cl_2) was added in increments of 0.001 mL (0.25 equiv) and 1H NMR spectra were recorded. Selected peaks shifted, as shown by the spectra in Figure s3. Data (equiv of **4a**/ NH signals, δ in ppm): 0.25/4.010 and 8.127; 0.50/4.053 and 8.134; 0.75/4.094 and 8.141; 1.0/ 4.129 and 8.131; 1.25/4.165 and 8.155; 1.50/4.195 and 8.158; 1.75/4.204 and 8.162; 2.0/4.241 and 8.169; 2.50/4.28 and 8.179; 2.75/4.296 and 8.183; 3.0/4.31 and 8.178; 4.0/4.342 and 8.184; 5.0/4.368 and 8.196; 6.0/4.387 and 8.227; 7.0/4.405 and 8.236; 8.0/4.418 and 8.234; 9.0/4.432 and 8.249.

Syntheses of previously unreported nitroolefins. A (*trans*-2-acetoxy- β -nitrostyrene; **4h).** A Schlenk flask was charged with *trans*-2-(2-nitrovinyl)phenol (0.5871 g, 3.54 mmol),^{s1a} Et_3N (0.500 mL, 3.58 mmol, 1.01 equiv), and CH_2Cl_2 (10 mL, dried and degassed using a Glass Contour solvent purification system) under N_2 . The solution was cooled to 0 °C and acetyl chloride (0.375 mL, 5.28 mmol, 1.49 equiv) was added dropwise by syringe with stirring. The cold bath was removed. After 12 h, CH_2Cl_2 (50 mL) was added. The mixture was washed with H_2O (50 mL), 1.0 M HCl, (50 mL), and brine (50 mL). The organic layer was dried (Na_2SO_4) and the

solvent was removed by rotary evaporation. The residue was chromatographed on a silica gel column (80:20 v/v hexanes/EtOAc). The solvent was removed from the combined product fractions by rotary evaporation. The residue was dried by oil pump vacuum at room temperature to give **4h** as a pale yellow solid (0.473 g, 2.28 mmol, 65%), which became a wax at 69-71 °C and liquefied at 74-76 °C (open capillary). Anal. Calcd. for C₁₀H₉NO₄ (207.18): C 57.97, H 4.38, N 6.76; found C 58.13, H 4.40, N 6.73. **B. (*trans*-2-benzoyloxy- β -nitrostyrene; **4i**).** *trans*-2-(2-nitrovinyl)phenol (0.5943 g, 3.60 mmol), ^{s1a} Et₃N (0.630 mL, 4.52 mmol, 1.26 equiv), CH₂Cl₂ (10 mL, dried and degassed using a Glass Contour solvent purification system), and benzoyl chloride (0.790 mL, 6.81 mmol, 1.89 equiv) were combined in a procedure analogous to A. An identical workup gave **4i** as a pale yellow solid (0.6629 g, 2.46 mmol, 69%), which became a wax at 78-82 °C and liquefied at 87-89 °C (open capillary). Anal. Calcd. for C₁₅H₁₁NO₄ (269.25): C 66.91, H 4.12, N 5.20; found C 67.04, H 4.20, N 5.01.

Data for **4h**. NMR (CDCl₃, δ in ppm): ¹H (500 MHz) 8.06 (d, ³J_{HH} = 13.5 Hz, 1H, CH-NO₂), 7.61-7.58 (m, 2H, CH=CHNO₂ and 1H of C₆H₄), 7.52 (t, ³J_{HH} = 8.5 Hz, 1H of C₆H₄), 7.32 (t, ³J_{HH} = 7.5 Hz, 1H of C₆H₄), 7.23 (d, ³J_{HH} = 8.5 Hz, 1H of C₆H₄), 2.43 (s, 3H, CH₃); ¹³C{¹H} (125 MHz): 168.8 (s, C(O)Me), 150.2, 138.7, 133.2, 133.1, 129.1, 126.8, 123.8, 123.0 (8 \times s, C₆H₄, CH=CHNO₂), 21.2 (s, CH₃). IR (powder film, cm⁻¹): 3115 (w), 1753 (s), 1638 (m), 1510 (s), 1339 (vs), 1194 (vs), 963 (s), 758 (s).

Data for **4i**. NMR (CDCl₃, δ in ppm): ¹H (500 MHz) 8.24 (m, 2H, CHNO₂ and 1 H of C₆H₅/C₆H₄), 8.13 (d, ³J_{HH} = 13.5 Hz, 1H, CH=CHNO₂), 7.70 (t, ³J_{HH} = 7.5 Hz, 1H of C₆H₅/C₆H₄), 7.66-7.56 (m, 5H of C₆H₅/C₆H₄), 7.38-7.33 (m, 2H of C₆H₅/C₆H₄); ¹³C{¹H} (125 MHz) 164.7 (s, C(O)Ph), 150.6 138.7, 134.5, 133.2, 133.2, 130.5, 129.2, 129.1, 128.5, 126.9, 124.0, 123.4 (12 \times s, CH=CHNO₂, C₆H₄, C₆H₅). IR (powder film, cm⁻¹): 3105 (w), 1742 (s), 1340 (s), 1209 (vs), 1057 (vs), 700 (vs).

Rate profiles, Figure 2. A 5 mm NMR tube was charged with a solution of **4d** (0.0180 g, 0.101 mmol), dimethyl malonate (**5c**, 0.0127 mL, 0.111 mmol, 1.10 equiv), catalyst (0.0020 mmol, 0.020 equiv), and Ph₂SiMe₂ (0.0020 mL, internal standard) in CD₂Cl₂ (1.0 mL). A ¹H

NMR spectrum was recorded to measure the initial ratio of **4d** to the standard. Then Et₃N (0.0050 mL, 0.036 mmol, 0.36 equiv) was added and the conversion to **6ac** was periodically measured by ¹H NMR. After 2 h, the mixture was loaded directly onto a plug of silica gel. The **6ac** was eluted with hexanes/EtOAc (1:1 v/v, 50 mL). The solvent was removed by rotary evaporation. The ee was assayed by chiral HPLC.^{s20}

Reactions, Scheme 3. A (entries 1-6). A 5 mm NMR tube was charged with a solution of **4a** (0.0054 g, 0.036 mmol, 1.0 equiv), dialkyl malonate (**5**, 0.039 mmol, 1.1 equiv), catalyst (0.0036 mmol, 0.10 equiv), and Ph₂SiMe₂ (0.0020 mL, internal standard) in CD₂Cl₂ (0.40 mL). A ¹H NMR spectrum was recorded to measure the initial ratio of the nitroolefin to the standard. Then Et₃N (0.0050 mL, 0.0036 g, 0.036 mmol, 1.0 equiv) was added. The conversion was monitored by ¹H NMR. After the specified time (Scheme 3), the mixture was loaded onto a plug of silica gel. The organic product (**6aa-ac**) was eluted with hexanes/EtOAc (1:1 v/v, 50 mL). The solvent was removed by rotary evaporation. The product ee was assayed by chiral HPLC.^{s20}

B (entries 7-11). A 5 mm NMR tube was charged with a solution of **4a** (0.0149 g, 0.010 mmol, 1.0 equiv), **5c**, (0.0138 mL, 0.0158 g, 0.120 mmol, 1.2 equiv), catalyst (0.010 mmol, 0.10 equiv), and Ph₂SiMe₂ (0.0020 mL, internal standard) in acetone-*d*₆ (0.7 mL). A ¹H NMR spectrum was recorded to measure the initial ratio of the nitroolefin to the standard. The sample was cooled to 0 °C, and Et₃N (0.0140 mL, 0.0101 g, 0.10 mmol, 1.0 equiv) was added. The conversion to **6ac** was monitored by ¹H NMR. After the specified time (Scheme 3), the mixture was loaded directly onto a plug of silica gel. The **6ac** was eluted with hexanes/EtOAc (7:3 v/v, 50 mL). The solvent was removed by rotary evaporation. The ee was assayed by chiral HPLC.^{s20}

Reactions, Scheme 4. A (with Λ-(*S,S*)-3**³⁺ 2Cl⁻BArf⁻).** A 5 mm NMR tube was charged with a solution of nitroolefin (0.036 mmol, 1.0 equiv), Λ-(*S,S*)-**3**³⁺ 2Cl⁻BArf⁻·2H₂O (0.0059 g, 0.0036 mmol, 0.10 equiv), **5c** (0.0045 mL, 0.0052 g, 0.039 mmol, 1.1 equiv), and Ph₂SiMe₂ (0.0020 mL, internal standard) in acetone-*d*₆ (0.40 mL). A ¹H NMR spectrum was recorded to measure the initial ratio of the nitroolefin to the standard. The sample was cooled to 0 °C, and Et₃N (0.0050 mL, 0.0036 g, 0.036 mmol, 1.0 equiv) was added. The conversion was monitored

by ^1H NMR. After the specified time (Scheme 4), the mixture loaded onto a plug of silica gel. The organic product (**6a**–**jc**) was eluted with hexanes/EtOAc (1:1 v/v). The solvent was removed by rotary evaporation. The product was dried by oil pump vacuum at room temperature and the ee assayed by chiral HPLC (Figures s4-s17).^{s20} **B (with Λ -(*S,S*)-**3**³⁺ **2BF**₄[–]**BAr**_f[–])**. A round bottom flask was charged with a solution of nitroolefin (0.355 mmol, 1.0 equiv), Λ -(*S,S*)-**3**³⁺ **2BF**₄[–]**BAr**_f[–]·3H₂O (0.0634 g, 0.0355 mmol, 0.10 equiv), and **5c** (0.0458 mL, 0.0529 g, 1.13 equiv) in acetone (3.3 mL), and cooled to 0 °C. Then Et₃N (0.0495 mL, 0.0359 g, 0.355 mmol, 1.0 equiv) was added. The reaction was monitored by TLC. After the specified time (Scheme 4), the solvent was removed by rotary evaporation and the residue was dissolved in CH₂Cl₂ (2 mL). The orange solution was chromatographed on a silica gel column (1.9 × 14 cm, 9:1 v/v hexanes/EtOAc). The solvent was removed from the product containing fractions by rotary evaporation. The product was dried by oil pump vacuum at room temperature (yields: Scheme 4) and the ee assayed by chiral HPLC (Figures s4-s17).^{s20} **C (with Λ -(*S,S*)-**3**³⁺ **3BF**₄[–])**. A 5 mL vial was charged with a solution of nitroolefin (0.10 mmol, 1.0 equiv), Λ -(*S,S*)-**3**³⁺ **3BF**₄[–]·2H₂O (0.0099 g, 0.010 mmol, 0.1 equiv), **5c** (0.0138 mL, 0.0158 g, 0.120 mmol, 1.2 equiv), and Ph₂SiMe₂ (0.0020 mL, internal standard) in acetone-*d*₆ (0.70 mL). A ^1H NMR spectrum was recorded to measure the initial ratio of the nitroolefin to the standard. The sample was cooled to 0 °C, and Et₃N (0.0140 mL, 0.0101 g, 0.10 mmol, 1.0 equiv) was added. The reaction was monitored by ^1H NMR and TLC. After the specified time (Scheme 4), the mixture was worked up as described in B.

Reactions, Scheme 5. A 1.5 mL vial was charged with a solution of azodicarboxylate **7** (0.023 g, 0.10 mmol), catalyst (0.0050 mmol, 0.05 equiv), and β -keto ester **8** (0.11 mmol, 1.10 equiv) in CH₂Cl₂ (1.0 mL). Then N-methylmorpholine (0.011 mL, 0.10 mmol, 1.0 equiv) was added. The reaction was monitored by TLC. After the specified time (Scheme 5), the mixture loaded onto a plug of silica gel. The organic product (**9**) was eluted with hexanes/EtOAc (9:1 v/v). The solvent was removed by rotary evaporation. The product was dried by oil pump vacuum at room temperature and the ee assayed by chiral HPLC (Figures s18-s19).^{s26}

Catalyst recycling, Λ -(*S,S*)- $\mathbf{3}^{3+}$ $2\text{Cl}^-\text{BAr}_\text{F}^-$. A round bottom flask was charged with a solution of **4a** (0.0529 g, 0.355 mmol, 1.0 equiv), Λ -(*S,S*)- $\mathbf{3}^{3+}$ $2\text{Cl}^-\text{BAr}_\text{F}^-\cdot 2\text{H}_2\text{O}$ (0.0543 g, 0.0326 mmol, 0.0918 equiv), and **5c** (0.0460 mL, 0.401 mmol, 1.13 equiv) in acetone (3.3 mL), and cooled to 0 °C. Then Et₃N (0.0460 mL, 0.0333 g, 0.330 mmol, 0.93 equiv) was added. The reaction was monitored by TLC. After 15 h, the solvent was removed by rotary evaporation and the residue was dissolved in CH₂Cl₂ (2 mL). The orange solution was washed with 3.0 M HCl (1 mL) and H₂O (2 × 1 mL) and chromatographed on a silica gel column (1.9 × 14 cm, 9:1 v/v hexanes/EtOAc). The solvent was removed from the organic product containing fractions by rotary evaporation and oil pump vacuum to give **6ac** as a white solid (0.0940 g, 0.334 mmol, 94%), which was analyzed by HPLC and ¹H and ¹³C{¹H} NMR as described below. The column was then washed with CH₂Cl₂ and the orange catalyst band was eluted with CH₂Cl₂/CH₃OH (98:2 v/v). The solvent was removed by rotary evaporation and oil pump vacuum at room temperature to recover Λ -(*S,S*)- $\mathbf{3}^{3+}$ $2\text{Cl}^-\text{BAr}_\text{F}^-\cdot \text{H}_2\text{O}$ (0.0526 g, 0.0319 mmol, 98%). Anal. Calcd. for C₇₄H₆₀-BCl₂CoF₂₄N₆·H₂O (1647.94): C 53.91, H 3.79, N 5.10; found C 53.84, H 4.08, N 4.96.

Diisopropyl 2-(2-nitro-1-phenylethyl)malonate (6aa**).**^{s21} This known compound was obtained as a spectroscopically pure colorless oil per the general procedures for Scheme 3. NMR (CDCl₃, δ in ppm):^{s22} ¹H (500 MHz) 7.32-7.22 (m, 5H, C₆H₅), 5.08 (sept, ³J_{HH} = 6.0 Hz, 1H, CH(CH₃)₂), 4.92 (dd, ²J_{HH} = 13.0 Hz, ³J_{HH} = 4.5 Hz, 1H, CHH'NO₂), 4.86-4.60 (m, 2H, C'H-(C'H₃)₂ and CHH'NO₂), 4.20 (td, ³J_{HH} = 8.5 Hz, ³J_{HH} = 5.0 Hz, 1H, CH=CH₂NO₂), 3.75 (d, ³J_{HH} = 9.5 Hz, 1H, CH(CO₂iPr)₂), 1.24 (d, ³J_{HH} = 6.5 Hz, 6H, CH(CH₃)₂), 1.07 (d, ³J_{HH} = 6.0 Hz, 3H, C'H((CH₃)(C'H₃))), 1.01 (d, ³J_{HH} = 6.5 Hz, 3H, C'H((CH₃)(C'H₃))); ¹³C{¹H} (125 MHz): 167.2 (s, CO₂iPr), 166.5 (s, C'O₂iPr), 136.4 (s, *i*), 129.0 (s, *o*), 128.4 (s, *p*), 128.3 (s, *m*),^{s16} 78.1 (s, CH₂NO₂), 70.1 (s, CH(CH₃)₂), 69.7 (s, C'H(C'H₃)₂), 55.3 (s, CH(CO₂iPr)₂), 43.1 (s, CH=CH₂NO₂), 21.7, 21.6, 21.41, 21.39 (4 × s, 4 × CH₃).

Enantiomeric excesses were determined by HPLC (see Figure s4) with a Chiralcel OD column (95:05 v/v hexane/isopropanol, 0.75 mL/min, λ = 220 nm); for Λ -(*S,S*)- $\mathbf{3}^{3+}$ $2\text{Cl}^-\text{BAr}_\text{F}^-$, t_R = 10.8 min (major), 13.3 min (minor); for Δ -(*S,S*)- $\mathbf{3}^{3+}$ $2\text{Cl}^-\text{BAr}_\text{F}^-$, similar but with major/min-

or reversed.^{s20}

Diethyl 2-(2-nitro-1-phenylethyl)malonate (6ab).^{s21} This known compound was obtained as a spectroscopically pure colorless oil per the general procedures for Scheme 3. NMR (CDCl₃, δ in ppm):^{s22} ¹H (500 MHz) 7.33-7.22 (m, 5H, C₆H₅), 4.92 (dd, ²J_{HH} = 13.5 Hz, ³J_{HH} = 5.0 Hz, 1H, CHH'NO₂), 4.86 (dd, ²J_{HH} = 13.0 Hz, ³J_{HH} = 9.0 Hz, 1H, CHH'NO₂), 4.26-4.19 (m, 3H, CH₂CH₃ and CH=CH₂NO₂), 4.00 (q, ³J_{HH} = 7.5 Hz, 2H, C'H₂C'H₃), 3.81 (d, ³J_{HH} = 9.5 Hz, 1H, CH(CO₂Et)₂), 1.26 (t, ³J_{HH} = 7.0 Hz, 3H, CH₂CH₃), 1.04 (t, ³J_{HH} = 7.0 Hz, 3H, C'H₂C'H₃); ¹³C{¹H} (125 MHz): 167.6 (s, CO₂Et), 166.9 (s, C'O₂Et), 136.3 (s, *i*), 129.1 (s, *o*), 128.5 (s, *p*), 128.1 (s, *m*),^{s16} 77.8 (s, CH₂NO₂), 62.3 (s, CH₂CH₃), 62.0 (s, C'H₂C'H₃), 55.1 (s, CH(CO₂Et)₂), 43.1 (s, CH=CH₂NO₂), 14.1 (s, CH₃), 13.9 (s, C'H₃).

Enantiomeric excesses were determined by HPLC (see Figure s5) with a Chiralpak AS-H column (90:10 v/v hexane/isopropanol, 1.0 mL/min, λ = 220 nm); for Λ -(*S,S*)-**3**³⁺ 2Cl⁻BAr_F⁻, *t*_R = 12.7 min (minor), 14.3 min (major); for Δ -(*S,S*)-**3**³⁺ 2Cl⁻BAr_F⁻, similar but with major/minor reversed.^{s20}

Dimethyl 2-(2-nitro-1-phenylethyl)malonate (6ac).^{s21} This known compound was isolated as a white solid per the general procedures for Scheme 4. NMR (CDCl₃, δ in ppm):^{s22} ¹H (500 MHz) 7.33-7.21 (m, 5H, C₆H₅), 4.93 (dd, ²J_{HH} = 13.0 Hz, ³J_{HH} = 5.0 Hz, 1H, CHH'NO₂), 4.87 (dd, ²J_{HH} = 13.0 Hz, ³J_{HH} = 9.0 Hz, 1H, CHH'NO₂), 4.24 (td, ³J_{HH} = 9.0 Hz, ³J_{HH} = 5.0 Hz, 1H, CH=CH₂NO₂), 3.86 (d, ³J_{HH} = 9.0 Hz, 1H, CH(CO₂Me)₂), 3.75 (s, 3H, CH₃), 3.54 (s, 3H, C'H₃); ¹³C{¹H} (125 MHz) 167.9 (s, CO₂Me), 167.3 (s, C'O₂Me), 136.2 (s, *i*), 129.1 (s, *o*), 128.4 (s, *p*), 128.0 (s, *m*), 77.5 (s, CH₂NO₂), 54.8 (s, CH(CO₂Me)₂), 53.1 (s, CH₃), 52.9 (s, C'H₃), 43.0 (s, CH=CH₂NO₂).

Enantiomeric excesses were determined by HPLC (see Figure s6) with a Chiralpak AD column (98:02 v/v hexane/isopropanol, 1.0 mL/min, λ = 220 nm); for Λ -(*S,S*)-**3**³⁺ 2BF₄⁻BAr_F⁻ (Λ -(*S,S*)-**3**³⁺ 2Cl⁻BAr_F⁻ similar), *t*_R = 32.4 min (major), 42.3 min (minor); for Λ -(*S,S*)-**3**³⁺ 3BF₄⁻, *t*_R = 33.3 min (major), 43.4 min (minor); for Δ -(*S,S*)-**3**³⁺ 2Cl⁻BAr_F⁻, similar but with major/minor reversed.^{s20}

Dimethyl 2-(2-nitro-1- α -naphthylethyl)malonate (6bc).^{s23a} This known compound was isolated as a colorless oil per the general procedures for Scheme 4. NMR (CDCl₃, δ in ppm):^{s22} ¹H (500 MHz) 8.18 (d, ³J_{HH} = 8.5 Hz, 1H of C₁₀H₇), 7.87 (d, ³J_{HH} = 8.0 Hz, 1H of C₁₀H₇), 7.80 (d, ³J_{HH} = 8.0 Hz, 1H of C₁₀H₇), 7.62 (t, ³J_{HH} = 8.5 Hz, 1H of C₁₀H₇), 7.53 (t, ³J_{HH} = 8.0 Hz, 1H of C₁₀H₇), 7.44-7.38 (m, 2H of C₁₀H₇), 5.25-5.16 (m, 2H, CHH'NO₂ and CH=CH₂NO₂), 5.07 (dd, ²J_{HH} = 13.5 Hz, ³J_{HH} = 4.5 Hz, 1H, CHH'NO₂), 4.12 (d, ³J_{HH} = 7.5 Hz, 1H, CH(CO₂Me)₂), 3.72 (s, CH₃), 3.54 (s, C'H₃); ¹³C{¹H} (125 MHz) 168.1 (s, CO₂Me), 167.6 (s, C'O₂Me), 134.3, 132.3, 131.1, 129.4, 129.2, 127.2, 126.2, 125.2, 124.1, 122.3 (10 \times s, C₁₀H₇), 76.7 (s, CH₂NO₂), 54.6 (s, CH(CO₂Me)₂), 53.1 (s, CH₃), 53.0 (s, C'H₃), 36.9 (s, CH=CH₂NO₂).

Enantiomeric excesses were determined by HPLC (see Figure s7) with a Chiralpak AD column (90:10 v/v hexane/isopropanol, 1.0 mL/min, λ = 254 nm); for all catalysts t_R = 13.5-14.1 min (major), 18.9-19.6 min (minor).^{s20}

Dimethyl 2-(2-nitro-1- β -naphthylethyl)malonate (6cc).^{s23b} This known compound was isolated as a yellow solid per the general procedures for Scheme 4. NMR (CDCl₃, δ in ppm):^{s22} ¹H (500 MHz) 7.83-7.79 (m, 3H of C₁₀H₇), 7.70 (s, 1H of C₁₀H₇), 7.50-7.48 (m, 2H of C₁₀H₇), 7.34 (d, ³J_{HH} = 8.5 Hz, 1H of C₁₀H₇), 5.01 (d, ³J_{HH} = 8.0 Hz, 2H, CH₂NO₂), 4.43 (q, ³J_{HH} = 7.0 Hz, 1H, CH=CH₂NO₂), 3.98 (d, ³J_{HH} = 9.0 Hz, 1H, CH(CO₂Me)₂), 3.77 (s, 3H, CH₃), 3.54 (s, 3H, C'H₃); ¹³C{¹H} (125 MHz) 166.0 (s, CO₂Me), 167.4 (s, C'O₂Me), 133.7, 133.4, 133.2, 129.1, 128.1, 127.8, 127.5, 126.7, 126.6, 125.3 (10 \times s, C₁₀H₇), 77.5 (s, CH₂NO₂), 54.9 (s, CH(CO₂Me)₂), 53.2 (s, CH₃), 53.0 (s, C'H₃), 43.2 (CH=CH₂NO₂).

Enantiomeric excesses were determined by HPLC (see Figure s8) with a Chiralcel OD column (70:30 v/v hexane/isopropanol, 1.0 mL/min, λ = 254 nm); for Λ -(*S,S*)-**3**³⁺ 2BF₄⁻BARf⁻, t_R = 14.4 min (major), 39.2 min (minor); for Λ -(*S,S*)-**3**³⁺ 3BF₄⁻ (older column), t_R = 12.5 min (major), 37.0 min (minor).^{s20}

Dimethyl 2-(2-nitro-1-(4-methoxyphenyl)ethyl)malonate (6dc).^{s23a,b} This known compound was isolated as a colorless oil per the general procedures for Scheme 4. NMR (CDCl₃, δ

in ppm):^{s22} ^1H (500 MHz) 7.14 (d, $^3J_{\text{HH}} = 8.5$ Hz, 2H, *m* to CH), 6.83 (d, $^3J_{\text{HH}} = 8.5$ Hz, 2H, *o* to CH), 4.89 (dd, $^2J_{\text{HH}} = 13.0$ Hz, $^3J_{\text{HH}} = 5.0$ Hz, 1H, CHH'NO₂), 4.82 (dd, $^2J_{\text{HH}} = 13.5$ Hz, $^3J_{\text{HH}} = 9.5$ Hz, 1H, CHH'NO₂), 4.19 (td, $^3J_{\text{HH}} = 9.0$ Hz, $^3J_{\text{HH}} = 5.0$ Hz, 1H, CH=CH₂NO₂), 3.82 (d, $^3J_{\text{HH}} = 9.5$ Hz, 1H, CH(CO₂Me)₂), 3.77 (s, 3H, CO₂CH₃), 3.76 (s, 3H, C'O₂C'H₃), 3.57 (s, 3H, ArOCH₃); $^{13}\text{C}\{^1\text{H}\}$ ^{s24} (125 MHz) 168.0 (s, CO₂Me), 167.4 (s, C'O₂Me), 159.6 (s, *p* to CH), 129.1 (s, *o* to CH), 128.0 (s, *i* to CH), 114.5 (s, *m* to CH), 77.8 (s, CH₂NO₂), 55.3 (s, ArO-CH₃), 55.0 (CH(CO₂Me)₂), 53.1 (s, CO₂CH₃), 53.0 (s, C'O₂C'H₃), 42.4 (s, CH=CH₂NO₂).

Enantiomeric excesses were determined by HPLC (see Figure s9) with a Chiralpak AD column (80:20 v/v hexane/isopropanol, 1.0 mL/min, $\lambda = 220$ nm); for all catalysts $t_{\text{R}} = 11.5$ -11.7 min (major), 17.5-18.2 min (minor).^{s20}

Dimethyl 2-(2-nitro-1-(3,4-dichlorophenyl)ethyl)malonate (6e_c). This new compound was isolated as a white solid per the general procedures for Scheme 4. NMR (CDCl₃, δ in ppm):^{s22} ^1H (500 MHz) 7.40 (d, $^3J_{\text{HH}} = 9.0$ Hz, 1H of C₆H₃), 7.35 (s, 1H of C₆H₃), 7.10 (d, $^3J_{\text{HH}} = 8.5$ Hz, 1H of C₆H₃), 4.90 (dd, $^2J_{\text{HH}} = 13.5$ Hz, $^3J_{\text{HH}} = 5.0$ Hz, 1H, CHH'NO₂), 4.85 (dd, $^2J_{\text{HH}} = 13.5$ Hz, $^3J_{\text{HH}} = 9.5$ Hz, 1H, CHH'NO₂), 4.20 (td, $^3J_{\text{HH}} = 9.0$ Hz, $^3J_{\text{HH}} = 5.0$ Hz, 1H, CH=CH₂NO₂), 3.82 (d, $^3J_{\text{HH}} = 9.5$ Hz, 1H, CH(CO₂Me)₂), 3.77 (s, 3H, CH₃), 3.62 (s, 3H, C'H₃); $^{13}\text{C}\{^1\text{H}\}$ (125 MHz) 167.5 (s, CO₂Me), 167.0 (s, C'O₂Me), 136.5, 133.3, 132.9, 131.1, 130.2, 127.4 (6 \times s, C₆H₃), 76.9 (s, CH₂NO₂), 54.4 (s, CH(CO₂Me)₂), 53.3 (CH₃), 53.2 (C'H₃), 42.1 (CH=CH₂NO₂). IR (powder film, cm⁻¹): 1744 (s), 1715 (s), 1549 (vs), 1265 (s), 1611 (s). mp 78-81 °C (open capillary). Anal. Calcd. for C₁₃H₁₃Cl₂NO₆ (350.15): C 44.59, H 3.74, N 4.00; found C 44.82, H 3.82, N 4.02.

Enantiomeric excesses were determined by HPLC (see Figure s10) with a Chiralpak AS-H column (90:10 v/v hexane/isopropanol, 1.0 mL/min, $\lambda = 220$ nm); for all catalysts $t_{\text{R}} = 20.4$ -21.3 min (minor), 22.7-23.5 min (major).^{s20}

Dimethyl 2-(2-nitro-1-(pyridine-3-yl)ethyl)malonate (6f_c).^{s23a} This known compound was isolated as an off white solid per the general procedures for Scheme 4. NMR (CDCl₃, δ in ppm):^{s22} ^1H (500 MHz) 8.55-8.52 (m, 2H of C₅H₄N), 7.59 (dt, $^3J_{\text{HH}} = 8.0$ Hz, $J_{\text{HH}} = 2.0$ Hz, 1H

of C₅H₄N), 7.27-7.25 (m, 1H of C₅H₄N), 4.94 (dd, ²J_{HH} = 13.5 Hz, ³J_{HH} = 8.5 Hz, CHH'NO₂), 4.89 (dd, ²J_{HH} = 13.5 Hz, ³J_{HH} = 9.5 Hz, 1H, CHH'NO₂), 4.26 (td, ³J_{HH} = 8.5 Hz, ³J_{HH} = 5.0 Hz, CH=CH₂NO₂), 3.86 (d, ³J_{HH} = 8.5 Hz, 1H, CH(CO₂Me)₂), 3.76 (s, CH₃), 3.58 (s, C'H₃); ¹³C{¹H} (125 MHz) 167.6 (s, CO₂Me), 167.0 (s, C'O₂Me), 149.9, 149.7, 135.5, 132.1, 123.8 (5 × s, C₅H₄N), 76.9 (s, CH₂NO₂), 54.3 (s, CH(CO₂Me)₂), 53.3 (s, CH₃), 53.2 (s, C'H₃), 40.7 (s, CH=CH₂NO₂).

Enantiomeric excesses were determined by HPLC (see Figure s11) with a Chiralpak AD column (60:40 v/v hexane/isopropanol, 1.0 mL/min, λ = 220 nm); for Λ-(*S,S*)-**3**³⁺ 2BF₄⁻BAr_F⁻ (Λ-(*S,S*)-**3**³⁺ 2Cl⁻BAr_F⁻ similar), t_R = 6.8 min (major), 9.2 min (minor).^{s20}

Dimethyl 2-(2-nitro-1-(2-trifluoromethylphenyl)ethyl)malonate (6gc). This new compound was isolated as a white solid per the general procedures for Scheme 4. NMR (CDCl₃, δ in ppm):^{s22} ¹H (500 MHz) 7.72 (d, ³J_{HH} = 7.5 Hz, 1H of C₆H₄), 7.53 (t, ³J_{HH} = 7.5 Hz, 1H of C₆H₄), 7.43 (t, ³J_{HH} = 8.0 Hz, 1H of C₆H₄), 7.37 (d, ³J_{HH} = 8.0 Hz, 1H of C₆H₄), 5.16 (dd, ²J_{HH} = 13.5 Hz, ³J_{HH} = 7.5 Hz, 1H, CHH'NO₂), 4.94 (dd, ²J_{HH} = 13.5 Hz, ³J_{HH} = 4.5 Hz, 1H, CHH'NO₂), 4.65 (td, ³J_{HH} = 7.5 Hz, ³J_{HH} = 4.5 Hz, 1H, CH=CH₂NO₂), 4.11 (d, ³J_{HH} = 7.5 Hz, 1H, CH(CO₂Me)₂), 3.75 (s, 3H, CH₃), 3.64 (s, 3H, C'H₃); ¹³C (125 MHz) 168.0 (s, CO₂Me), 167.3 (s, C'O₂Me), 135.1, 132.5 (2 × s, C₆H₄), 129.0 (q, ²J_{CF} = 29.8 Hz, CCF₃), 128.6, 128.0 (2 × s, C₆H₄), 127.3 (q, ³J_{CF} = 5.9 Hz, CHCCF₃), 124.2 (q, ¹J_{CF} = 272.6 Hz, CF₃), 76.6 (s, CH₂NO₂), 54.0 (s, CH(CO₂Me)₂), 53.2 (CH₃), 53.1 (C'H₃), 38.1 (s, CH=CH₂NO₂). IR (powder film, cm⁻¹): 2957 (w), 1751 (s), 1734 (s), 1549 (s), 1126 (vs), 777 (s). mp 76-79 °C (open capillary). Anal. Calcd. for C₁₄H₁₄F₃NO₆ (349.26): C 48.14, H 4.04, N 4.01; found C 47.91, H 3.93, N 3.91.

Enantiomeric excesses were determined by HPLC (see Figure s12) with a Chiralcel OD column (95:05 v/v hexane/isopropanol, 1.0 mL/min, λ = 220 nm); for all catalysts t_R = 12.7 min (minor), 26.6-28.6 min (major).^{s20}

Dimethyl 2-(2-nitro-1-(2-acetoxyphenyl)ethyl)malonate (6hc). This new compound was isolated as a colorless oil per the general procedures for Scheme 4. NMR (CDCl₃, δ in

ppm):^{s22} ^1H (500 MHz) 7.32 (t, $^3J_{\text{HH}} = 7.0$ Hz, 1H of C_6H_4), 7.27-7.25 (m, 1H of C_6H_4), 7.20 (t, $^3J_{\text{HH}} = 7.5$ Hz, 1H of C_6H_4), 7.15 (d, $^3J_{\text{HH}} = 8.0$ Hz, 1H of C_6H_4), 4.94-4.86 (m, 2H, $\text{CH}_2\text{-NO}_2$), 4.50 (q, $^3J_{\text{HH}} = 7.0$ Hz, 1H, $\text{CH=CH}_2\text{NO}_2$), 3.92 (d, $^3J_{\text{HH}} = 8.5$ Hz, 1H, $\text{CH}(\text{CO}_2\text{Me})_2$), 3.74 (s, 3H, OCH_3), 3.59 (s, 3H, $\text{OC}'\text{H}_3$), 2.39 (s, 3H, CH_3); $^{13}\text{C}\{^1\text{H}\}$ (125 MHz) 169.2, 168.0, 167.4 (3 \times s, CO_2Me , $\text{C}'\text{O}_2\text{Me}$, $\text{C}(\text{O})\text{Me}$), 148.7, 129.4, 128.5, 128.2, 126.5, 123.5 (6 \times s, C_6H_4), 76.7 (s, CH_2NO_2), 53.8 (s, $\text{CH}(\text{CO}_2\text{Me})_2$), 53.1 (s, OCH_3 , $\text{OC}'\text{H}_3$), 36.7 (s, $\text{CH=CH}_2\text{NO}_2$), 21.2 (s, CH_3), IR (neat oil, cm^{-1}): 2957 (w), 1734 (vs), 1555 (s), 1192 (s), 1171 (s). Anal. Calcd. for $\text{C}_{15}\text{H}_{17}\text{NO}_8$ (339.30): C 53.10, H 5.05, N 4.13; found C 53.39, H 5.11, N 4.11.

Enantiomeric excesses were determined by HPLC (see Figure s13) with a Chiralcel OD column (90:10 v/v hexane/isopropanol, 1.0 mL/min, $\lambda = 220$ nm); for $\Lambda\text{-(S,S)-}\mathbf{3}^{3+} 2\text{BF}_4^-\text{BARf}^-$ ($\Lambda\text{-(S,S)-}\mathbf{3}^{3+} 2\text{Cl}^-\text{BARf}^-$ similar), $t_{\text{R}} = 18.9$ min (minor), 28.5 min (major)); for $\Lambda\text{-(S,S)-}\mathbf{3}^{3+} 3\text{BF}_4^-$ (older column), $t_{\text{R}} = 19.7$ min (minor), 31.3 min (major).^{s20}

Dimethyl 2-(2-nitro-1-(2-benzoyloxyphenyl)ethyl)malonate (6ic). This new compound was isolated as a pale yellow oil per the general procedures for Scheme 4. NMR (CDCl_3 , δ in ppm):^{s22} ^1H (500 MHz) 8.27 (d, $^3J_{\text{HH}} = 7.0$ Hz, 2H of $\text{C}_6\text{H}_5/\text{C}_6\text{H}_4$), 7.69 (t, $^3J_{\text{HH}} = 7.0$ Hz, 1H of $\text{C}_6\text{H}_5/\text{C}_6\text{H}_4$), 7.57 (t, $^3J_{\text{HH}} = 8.0$ Hz, 2H of $\text{C}_6\text{H}_5/\text{C}_6\text{H}_4$), 7.39-7.33 (m, 2H of $\text{C}_6\text{H}_5/\text{C}_6\text{H}_4$), 7.28-7.25 (m, 2H of $\text{C}_6\text{H}_5/\text{C}_6\text{H}_4$), 4.98 (dd, $^2J_{\text{HH}} = 13.5$ Hz, $^3J_{\text{HH}} = 8.5$ Hz, 1H, $\text{CHH}'\text{NO}_2$), 4.91 (dd, $^2J_{\text{HH}} = 13.5$ Hz, $^3J_{\text{HH}} = 5.0$ Hz, 1H, $\text{CHH}'\text{NO}_2$), 4.60 (td, $^3J_{\text{HH}} = 8.5$ Hz, $^3J_{\text{HH}} = 5.0$ Hz, 1H, $\text{CH=CH}_2\text{NO}_2$), 3.96 (d, $^2J_{\text{HH}} = 8.5$ Hz, 1H, $\text{CH}(\text{CO}_2\text{Me})_2$), 3.72 (s, 3H, CH_3), 3.53 (s, 3H, $\text{C}'\text{H}_3$); $^{13}\text{C}\{^1\text{H}\}$ (125 MHz) 167.9 (s, CO_2Me), 167.4 (s, $\text{C}'\text{O}_2\text{Me}$), 164.9 (s, $\text{C}(\text{O})\text{Ph}$), 149.0, 134.1, 130.5, 129.6, 129.1, 129.0, 128.7, 128.3, 126.6, 123.6 (10 \times s, $\text{C}_6\text{H}_5/\text{C}_6\text{H}_4$), 76.6 (s, CH_2NO_2), 53.8 (s, $\text{CH}(\text{CO}_2\text{Me})_2$), 53.1 (s, CO_2Me), 53.1 (s, $\text{C}'\text{O}_2\text{Me}$), 36.7 (s, $\text{CH=CH}_2\text{-NO}_2$). IR (neat oil, cm^{-1}): 2955 (w), 1732 (vs), 1555 (s), 1213 (s), 708 (vs). Anal. Calcd. for $\text{C}_{20}\text{H}_{19}\text{NO}_8$ (401.37): C 59.85, H 4.77, N 3.49; found C 59.59, H 4.80, N 3.40.

Enantiomeric excesses were determined by HPLC (see Figure s14) with a Chiralpak AD column (90:10 v/v hexane/isopropanol, 1.0 mL/min, $\lambda = 220$ nm); for all catalysts $t_{\text{R}} = 15.5\text{-}15.7$ min (major), 26.0-26.3 min (minor).^{s20}

Dimethyl 2-(2-nitro-1-(2-benzyloxyphenyl)ethyl)malonate (6jc). This new compound was isolated as a colorless oil per the general procedures for Scheme 4. NMR (CDCl₃, δ in ppm):^{s22} ¹H (500 MHz) 7.47 (d, J = 7.5 Hz, 2H of C₆H₅/C₆H₄), 7.42 (t, 7.5 Hz, 2H of C₆H₅/C₆H₄), 7.36 (t, J = 7.5 Hz, 1H of C₆H₅/C₆H₄), 7.24 (t, J = 7.5 Hz, 1H of C₆H₅/C₆H₄), 7.17 (d, J = 7.5 Hz, 1H of C₆H₅/C₆H₄), 6.93 (d, J = 8.5 Hz, 1H of C₆H₅/C₆H₄), 6.89 (t, J = 7.5 Hz, 1H of C₆H₅/C₆H₄); 5.13 (d, ³J_{HH} = 2.5 Hz, 2H, CH₂Ph), 5.05 (dd, J = 13.0 Hz, J = 4.5 Hz, 1H, CHH'-NO₂), 4.84 (dd, J = 13.0 Hz, J = 4.5 Hz, 1H, CHH'NO₂), 4.44 (td, ³J_{HH} = 9.5 Hz, ³J_{HH} = 4.5 Hz, 1H, CH=CH₂NO₂), 4.17 (d, ³J_{HH} = 9.5 Hz, 1H, CH(CO₂Me)₂), 3.72 (s, 3H, CH₃), 3.50 (s, 3H, C'H₃); ¹³C (125 MHz) 168.3, (s, CO₂Me), 167.7 (s, C'O₂Me), 156.7, 136.5, 130.9, 129.8, 128.9, 128.3, 127.7, 123.9, 121.2, 112.4 (10 × s, C₆H₄/C₆H₅), 76.0 (s, CH₂NO₂), 70.6 (CH₂Ph), 53.0 (s, CH(CO₂Me)₂), 52.7 (s, CH₃), 52.6 (s, C'H₃), 40.5 (s, CH=CH₂NO₂). IR (neat oil, cm⁻¹): 3034 (w), 2955 (w), 2886 (w), 1732 (vs), 1553 (vs), 1238 (vs), 752 (s), 696 (m). Anal. Calcd. for C₂₀H₂₁NO₇ (387.38): C 62.01, H 5.46, N 3.62; found C 62.25, H 5.46, N 3.57.

Enantiomeric excesses were determined by HPLC (see Figure s15) with a Chiralcel OD column (90:10 v/v hexane/isopropanol, 1.0 mL/min, λ = 220 nm); for all catalysts t_R = 12.2-12.4 min (minor), 23.7-24.0 min (major).^{s20}

Dimethyl 2-(2-nitro-1-furylethyl)malonate (6kc).^{s23a,b} This known compound was isolated as a yellowish oil per the general procedures for Scheme 4. NMR (CDCl₃, δ in ppm):^{s22} ¹H (500 MHz) 7.33 (m, 1H of C₄H₃O), 6.28 (m, 1H of C₄H₃O), 6.21 (d, ³J_{HH} = 3.5 Hz, 1H of C₄H₃O), 4.89 (m, 2H, CH₂NO₂), 4.39 (td, ³J_{HH} = 17.5 Hz, ³J_{HH} = 12.5 Hz, 1H, CH=CH₂NO₂), 3.93 (d, ³J_{HH} = 8.0 Hz, 1H, CH(CO₂Me)₂), 3.75 (s, 3H, CH₃), 3.68 (s, 3H, C'H₃); ¹³C{¹H} (125 MHz) 167.6 (s, CO₂Me), 167.3 (s, C'O₂Me), 149.5, 143.0, 110.7, 106.5 (4 × s, C₄H₃O), 75.4 (s, CH₂NO₂), 53.2 (s, CH₃), 53.1 (s, C'H₃), 52.8 (s, CH(CO₂Me)₂), 36.9 (s, CH=CH₂NO₂).

Enantiomeric excesses were determined by HPLC (see Figure s16) with a Chiralcel OD column (90:10 v/v hexane/isopropanol, 1.0 mL/min, λ = 220 nm); for all catalysts t_R = 11.8-12.0 min (minor), 28.7-29.0 min (major).^{s20}

Dimethyl 2-(2-nitro-1-propylethyl)malonate (6lc).^{s25} This known compound was iso-

lated as a colorless oil per the general procedures for Scheme 4. NMR (CDCl₃, δ in ppm):^{s22} ¹H (500 MHz) 4.69 (dd, ²J_{HH} = 13.5 Hz, ³J_{HH} = 8.5 Hz, 1H, CHH'NO₂), 4.52 (dd, ²J_{HH} = 13.5 Hz, ³J_{HH} = 6.5 Hz, 1H, CHH'NO₂), 3.762 (s, 3H, OCH₃), 3.756 (s, 3H, OC'H₃), 3.66 (d, ³J_{HH} = 6.0 Hz, 1H, CH(CO₂Me)₂), 2.90 (sext, ³J_{HH} = 6.5 Hz, 1H, CH=CH₂NO₂), 1.46-1.34 (m, 4H, CH₂-CH₂), 0.92 (t, ³J_{HH} = 7.5 Hz, 3H, CH₃); ¹³C{¹H} (125 MHz) 168.5 (s, CO₂Me), 168.3 (s, C'O₂Me), 76.7 (s, CH₂NO₂), 53.0 (s, OCH₃), 52.9 (s, OC'H₃), 52.4 (s, CH(CO₂Me)₂), 36.9 (s, CH=CH₂NO₂), 32.3 (s, CH₂CH₂CH₃), 20.0 (s, CH₂CH₂CH₃), 13.9 (s, CH₃).

Enantiomeric excesses were determined by HPLC (see Figure s17) with a Chiralcel OD column (97.5:2.5 v/v hexane/isopropanol, 1.0 mL/min, λ = 220 nm); for (*S,S*)-**3**³⁺ 2BF₄⁻BARf⁻ (newer column), *t*_R = 12.4 min (minor), 20.3 min (major); for Λ -(*S,S*)-**3**³⁺ 3BF₄⁻ (older column), *t*_R = 9.7 min (minor), 18.8 min (major).^{s20}

***N,N'*-Bis(*t*-butoxycarbonyl)-1-hydrazino-2-oxocyclopentanecarboxylic acid ethyl ester (9a).**^{s26a} This known compound was obtained as a colorless oil per the general procedure for Scheme 5. NMR (CDCl₃, δ in ppm): ¹H (500 MHz) 6.51 (br s, 1H, NH), 4.32-4.16 (m, 2H, CH₂CH₃), 2.95-1.86 (m, 6H, 3 \times ring CH₂), 1.57-1.33 (m, 18H, C(CH₃)₃ and C'(C'H₃)₃), 1.25 (t, ³J_{HH} = 7.0 Hz, 3H, CH₂CH₃); ¹³C{¹H} (125 MHz) 205.6 (s, (C=O)CH₂), 167.3 (s, (C=O)OEt), 155.1 and 154.3 (2s, (C=O)O-*t*-Bu and (C'=O')O'-*t*-Bu'), 82.7 and 81.4 (2s, C(CH₃)₃ and C'(C'H₃)₃), 62.1 (s, CH₂CH₃), 35.9 (s), 32.0 (s), 28.1 and 28.0 (2s, C(CH₃)₃ and C'(C'H₃)₃), 27.7 (s), 18.4 (s), 14.0 (s, CH₂CH₃).

Enantiomeric excesses were determined by HPLC (see Figure s18) with a Chiralpak AD column (96:4 v/v hexane/isopropanol, 1.0 mL/min, λ = 210 nm); for Λ -(*S,S*)-**3**³⁺ 2Cl⁻BARf⁻, *t*_R = 11.2 min (minor), 14.5 min (major); for Δ -(*S,S*)-**3**³⁺ 2Cl⁻BARf⁻, *t*_R = 11.4 min (major), 15.8 min (minor).^{s26a,s27}

***N,N'*-Bis(*t*-butoxycarbonyl)-1-hydrazino-2-oxocyclohexanecarboxylic acid ethyl ester (9b).**^{s26} This known compound was obtained as a colorless oil per the general procedure for Scheme 5. NMR (CDCl₃, δ in ppm): ¹H (500 MHz) 6.51 (br s, 1H, NH), 4.33-4.12 (m, 2H, CH₂CH₃), 2.92-1.81 (m, 8H, 4 \times ring CH₂), 1.52-1.37 (m, 18H, C(CH₃)₃ and C'(C'H₃)₃), 1.28-

1.22 (m, 3H, CH₂CH₃); ¹³C{¹H} (125 MHz) 205.4 (s, (C=O)CH₂), 169.4 (s, (C=O)OEt), 155.1 and 154.3 (2s, (C=O)O-t-Bu and (C'=O')O'-t-Bu'), 82.7 and 81.4 (2s, C(CH₃)₃ and C'(C'H₃)₃), 62.1 (s, CH₂CH₃), 38.0 (s), 35.8 (s), 34.7 (s), 28.1 and 28.0 (2s, C(CH₃)₃ and C'(C'H₃)₃), 20.9 (s), 18.4 (s), 14.0 (s, CH₂CH₃).

Enantiomeric excesses were determined by HPLC (see Figure s19) with a Chiralpak AS-H column (99:1 v/v hexane/isopropanol, 1.0 mL/min, λ = 210 nm); for Λ-(*S,S*)-**3**³⁺ 2Cl[−]BAr_F[−], t_R = 9.7 min (major), 12.6 min (minor); for Δ-(*S,S*)-**3**³⁺ 2Cl[−]BAr_F[−], t_R = 9.6 min (minor), 11.6 min (major).^{s26b,s27}

Crystallography. A (Λ-(*S,S*)-3**³⁺ 3Cl[−]·2H₂O·2CH₃OH).** A solution of Λ-(*S,S*)-**3**³⁺ 3Cl[−] in CH₃OH/H₂O (2:1 v/v) was allowed to slowly concentrate at room temperature. After 3 d, orange rods were collected. Data were obtained as noted in Table s1. Cell parameters were determined from 60 data frames taken at 0.5° widths. Integrated intensity information for each reflection was obtained by reduction of the data frames with the program APEX2.^{s28} Data were corrected for Lorentz, polarization, and crystal decay effects. The program SADABS^{s29} was used to correct for absorption effects. The structure was solved by direct methods using SHELXTL (XS).^{s30} For each cobalt atom, two molecules of both H₂O and CH₃OH were located. Hydrogen atoms were placed in idealized positions and refined using a riding model. Non-hydrogen atoms were refined with anisotropic thermal parameters. The thermal ellipsoids of the carbon atoms of two phenyl rings showed significant elongation, indicating disorder. This was successfully modeled (C2B/C, C3B/C, C4B/C, C5B/C, C6B/C, C7B/C, 61:39 occupancy ratio; C9A/E, C10A/E, C11A/E, C12A/E, C13A/E, and C14A/E, 53:47 occupancy ratio). The parameters were refined by weighted least squares refinement on *F*² to convergence.^{s30,s31} The absolute stereochemistry was confirmed (absolute structure parameter = 0.000(12); Table s1).^{s32} **B (Δ-(*R,R*)-**3**³⁺ 3Cl[−]·H₂O·3CH₃OH).** A solution of Δ-(*R,R*)-**3**³⁺ 3Cl[−] in CH₃OH was allowed to slowly concentrate at room temperature. After 5 d, orange rods were collected. Data were obtained as noted in Table s1. Cell parameters were determined from 180 data frames taken at 0.5° widths. Integrated intensity information for each reflection was obtained by reduction of the data frames with the prog-

ram APEX2.^{s28} Data were corrected for Lorentz, polarization, and crystal decay effects. The program SADABS^{s29} was used to correct for absorption effects. The structure was solved by direct methods using SHELXTL (XS).^{s30} For each cobalt atom, one molecule of water and ca. three molecules of CH₃OH were located. The thermal ellipsoids associated with one CH₃OH molecule indicated partial occupancy, which refined to 0.88; this value was fixed for the final refinement. Hydrogen atoms were placed in idealized positions and refined using a riding model. Non-hydrogen atoms were refined with anisotropic thermal parameters. The thermal ellipsoids of the carbon atoms of one phenyl ring indicated disorder. This refined to a 76:24 occupancy ratio (C30/30A, C31/31A, C32/32A, C33/33A, C34/34A, C35/35A). The ellipsoids of the disordered atoms were constrained. The parameters were refined by weighted least squares refinement on F^2 to convergence.^{s30,s31} The absolute stereochemistry was confirmed (absolute structure parameter = 0.012 (4); Table s1).^{s32} **C (Δ -(*S,S*)-**3**³⁺ 3Cl⁻·12.5H₂O).** A solution of crude Δ -(*S,S*)-**3**³⁺ 3Cl⁻ (obtained from the Dowex column and containing some Λ diastereomer) in hot H₂O was allowed to slowly concentrate at room temperature. After 7 d, orange blocks were collected. Data were obtained as noted in Table s1. Cell parameters were determined from 180 data frames taken at 0.5° widths. Integrated intensity information for each reflection was obtained by reduction of the data frames with the program APEX2.^{s28} Data were corrected for Lorentz, polarization, and crystal decay effects. The program SADABS^{s29} was used to correct for absorption effects, and the structure was solved using SHELXTL (XS).^{s30} The asymmetric unit contained two unique Δ -(*S,S*)-**3**³⁺ moieties ($Z = 4$; $Z' = 2$) and 25 molecules of water. Hydrogen atoms were placed in idealized positions and refined using a riding model. All non-hydrogen atoms were refined with anisotropic thermal parameters. Several atoms yielded non-positive definite matrices and some exhibited elongated thermal ellipsoids. Strong constraints (listed in the Validation Response Form of the CHECKCIF file) were placed to keep the thermal ellipsoids meaningful. The parameters were refined by weighted least squares refinement on F^2 to convergence.^{s30,s31} Platon^{s33} was used to verify the absence of additional symmetry and voids. The absolute stereochemistry was confirmed (absolute structure parameter = 0.069(4); Table s1).^{s32}

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(s2) (a) Brookhart, M.; Grant, B.; Volpe, Jr., A. F. *Organometallics* **1992**, *11*, 3920-3922. (b) For hazardous aspects of this synthesis, consult Yakelis, N. A.; Bergman, R. G. *Organometallics* **2005**, *24*, 3579-3581.

(s3) This compound has been reported earlier in references [s4a,b](#). In the former paper, no data were given for this salt, but the tris(perchlorate) salt Λ -(*S,S*)-**3**³⁺ 3ClO₄⁻ was characterized by CD spectroscopy and microanalysis. The cobalt configuration was correctly assigned, but the carbon configurations were misassigned. In the latter paper, the cobalt and carbon configurations were established crystallographically following anion metathesis.

(s4) (a) Bosnich, B.; Harrowfield, J. MacB. *J. Am. Chem. Soc.* **1972**, *94*, 3425-3437. (b) Kuroda, R.; Mason, S. F. *J. Chem. Soc., Dalton Trans.* **1977**, *10*, 1016-1020.

(s5) Additional papers of historical relevance are as follows: (a) Gillard, R. D. *Tetrahedron* **1965**, *21*, 503-506. In this work, the carbon configurations were correctly assigned, but using salts with unspecified anions. There were no assignments of cobalt configurations. (b) Fereday, P. L.; Mason, S. F. *Chem. Commun.* **1971**, 1314-1315. In this work, the cobalt configurations were correctly assigned, but the carbon configurations misassigned, using CD spectra of salts with unspecified anions.

(s6) This complex has been previously reported, but no characterization was supplied (for the analogous complex of racemic **2** (a monohydrate), a microanalysis was given). See (a) Williams, O. F.; Bailar, Jr. J. C. *J. Am. Chem. Soc.* **1959**, *81*, 4464-4469 and especially the third paragraph on page 4467. (b) For the analogous perchlorate salt, see reference [s4a](#). (c) An authentic sample was prepared by the procedure in reference [s6a](#) using (*S,S*)-**2**. Selected data: green powder. NMR (CD₃OD, δ in ppm): ¹H (500 MHz) 7.43-7.22 (m, 20H, Ph), 6.61 (d, ³J_{HH} = 10 Hz, 4H, NHH'), 5.16 (br, 4H, NHH'), 4.87 (br s, CD₃OH/H₂O, 5H), 4.67 (s, 4H, CHNH₂); ¹³C{¹H}

(125 MHz) 138.8 (s, *i*), 130.2 (s, *o*), ^{s14} 129.9 (s, *p*), 128.7 (s, *m*), ^{s16} 66.8 (s, CHNH₂).

(s7) (a) Another CHPh signal was observed at δ 58.6 ppm, derived from the diprotonated ligand (*S,S*)-**2**. The intensity of this signal was disproportionate to its relative concentration (ca. \times 2), as verified by spectra of independently prepared mixtures. (b) It is presumed that the intensities of signals due to Δ and Λ diastereomers are proportional to concentration.

(s8) These NMR solvents were not dried.

(s9) The degree of hydration was generally assigned by fitting the microanalytical data. Although the ¹H NMR spectra also showed OH signals, several factors can enhance their integrals. For example, in some cases the NMR solvents were not dried (so footnoted). In other cases, the NHH' protons undergo (partial) exchange with the deuterated solvent (so footnoted; CD₃OD will then give rise to a hydroxyl peak although acetone-*d*₆ should only give acetone-*d*₅).

(s10) Previous reports of this compound did not include microanalyses or any assignment of the degree of hydration. ^{s3-s5}

(s11) (a) This procedure was reported to give exclusively Δ -(*S,S*)-**3**³⁺ 3ClO₄[−] in reference ^{s4a}, but a yield was not quoted. (b) The Δ diastereomer was also isolated as a non-hydrated crystalline byproduct from the synthesis of Λ -(*S,S*)-**3**³⁺ 3ClO₄[−], and characterized by microanalysis and CD spectroscopy. (c) In our hands, we repeatedly obtained 15-18% of the Λ diastereomer (four independent syntheses).

(s12) Robinson, W. R. *J. Chem. Ed.* **1985**, 62, 1001.

(s13) No attempts were made to characterize the degree of hydration of salts that were mixtures of diastereomers (and thus of variable composition).

(s14) This compound was similarly prepared in reference ^{s4a} and characterized by microanalysis and CD spectroscopy. The degree of hydration was presumed to be identical.

(s15) The NH and/or NH' signal was not observed or was of reduced intensity due to H/D exchange.

(s16) The ¹³C{¹H} signal with the chemical shift closest to benzene was assigned to the *meta* carbon atom: Mann, B. E. *J. Chem. Soc. Perkin Trans. 2* **1972**, 30-34.

(s17) With the exception of the least intense 129.2 ppm signal, the phenyl resonances of the minor Λ diastereomer appeared to overlap with those of the major Δ diastereomer.

(s18) A similar experiment has been mentioned in reference [s4a](#), but without any quantitative details. The conversion is represented as 100% in the experimental section, and "at least 90%" in the text.

(s19) Dewey, M. A.; Gladysz, J. A. *Organometallics* **1993**, *12*, 2390-2392.

(s20) The absolute configurations of **6aa-ac** were assigned by HPLC using conditions similar to those in the literature (see references [s23c,d](#)). Catalysts with Λ -(*S,S*) configurations gave predominantly *R* enantiomers. The dominant configurations of **6bc-6lc** were assigned assuming analogous relative retention times.

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(s27) The chiral HPLC column employed with **9a** featured the same stationary phase but a different particle size from that used earlier in the literature.^{[s26](#)} It is presumed that the well separated enantiomers exhibit identical orders of elution, and configurations were assigned accordingly. The dominant configurations of **9b** were assigned assuming analogous relative retention

times.

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(s29) Sheldrick, G. M. “SADABS (version 2008/1): Program for Absorption Correction of Area Detector Frames”, BRUKER AXS Inc., 5465 East Cheryl Parkway, Madison, WI 53711-5373 USA.

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Table s1. Summary of crystallographic data for solvates of stereoisomers of $\mathbf{3}^{3+} \mathbf{3Cl}^-$.

| Complex | Δ -(<i>S,S</i>)- $\mathbf{3}^{3+} \mathbf{3Cl}^- \cdot 2\text{H}_2\text{O} \cdot 2\text{CH}_3\text{OH}$ | Δ -(<i>R,R</i>)- $\mathbf{3}^{3+} \mathbf{3Cl}^- \cdot \text{H}_2\text{O} \cdot 3\text{CH}_3\text{OH}$ ^{<i>a</i>} | Δ -(<i>S,S</i>)- $\mathbf{3}^{3+} \mathbf{3Cl}^- \cdot 12.5\text{H}_2\text{O}$ |
|--|--|---|--|
| empirical formula | C ₄₄ H ₆₀ Cl ₃ CoN ₆ O ₄ | C _{44.88} H _{61.54} Cl ₃ CoN ₆ O _{3.88} ^{<i>a</i>} | C ₈₄ H ₁₄₆ Cl ₆ Co ₂ N ₁₂ O ₂₅ ^{<i>b</i>} |
| formula weight | 902.26 | 912.60 | 2054.69 ^{<i>b</i>} |
| temperature of collection [K] | 110(2) | 110(2) | 110(2) |
| diffractometer | Bruker APEX 2 | Bruker GADDS | Bruker GADDS |
| wavelength [Å] | 0.71073 | 1.54178 | 1.54178 |
| crystal system | tetragonal | tetragonal | monoclinic |
| space group | P4(3) | P4(1) | P2(1) |
| unit cell dimensions: | | | |
| <i>a</i> [Å] | 13.839(4) | 13.8725(7) | 12.0877(6) |
| <i>b</i> [Å] | 13.839(4) | 13.8725(7) | 25.0985(14) |
| <i>c</i> [Å] | 24.216(9) | 24.3173(13) | 16.8235(8) |
| α [deg] | 90 | 90 | 90 |
| β [deg] | 90 | 90 | 90.887(3) |
| γ [deg] | 90 | 90 | 90 |
| V [Å ³] | 4638(2) | 4679.8(4) | 5103.4(5) |
| Z | 4 | 4 | 2 ^{<i>b</i>} |
| ρ _{calc} [Mg/m ³] | 1.292 | 1.295 | 1.337 |
| Absorption coefficient [mm ⁻¹] | 0.590 | 4.822 | 4.607 |
| F(000) | 1904 | 1928 | 2180 |
| crystal size [mm ³] | 0.60 × 0.10 × 0.10 | 0.12 × 0.08 × 0.04 | 0.16 × 0.15 × 0.07 |
| Θ [deg] | 2.23 to 27.49 | 3.19 to 60.00 | 2.63 to 60.00 |
| range / indices (<i>h</i> , <i>k</i> , <i>l</i>) | −17,17; −17,17; −31,31 | −15,15; −15,15; −25,26 | −13,13; −28,27; −18,18 |
| reflections collected | 70126 | 65183 | 112447 |
| independent reflections | 10535 [R(int) = 0.0425] | 6282 [R(int) = 0.0647] | 14790 [R(int) = 0.0916] |
| completeness to Θ = 27.49° | 99.7% | 98.5% | 99.9% |

| | | | |
|--|------------------------------------|------------------------------------|------------------------------------|
| absorption correction | semiempirical from equivalents | semiempirical from equivalents | semiempirical from equivalents |
| max. and min. transmission | 0.9434 and 0.7187 | 0.8305 and 0.5953 | 0.7386 and 0.5260 |
| refinement method | full-matrix least-squares on F^2 | full-matrix least-squares on F^2 | full-matrix least-squares on F^2 |
| data / restraints / parameters | 10535 / 7 / 525 | 6282 / 21 / 546 | 14790 / 265 / 1105 |
| goodness-of-fit on F^2 | 1.059 | 1.071 | 1.149 |
| final R indices [$I > 2\sigma(I)$] | $R1 = 0.0442$, $wR2 = 0.1171$ | $R1 = 0.0407$, $wR2 = 0.0841$ | $R1 = 0.0746$, $wR2 = 0.1791$ |
| R indices (all data) | $R1 = 0.0501$, $wR2 = 0.1220$ | $R1 = 0.0510$, $wR2 = 0.0871$ | $R1 = 0.0807$, $wR2 = 0.1830$ |
| absolute structure parameter | 0.000(12) | 0.012(4) | 0.069(5) |
| largest diff. peak and hole [$e \text{ \AA}^{-3}$] | 0.945 / -0.255 | 0.362 / -0.622 | 0.972 / -0.698 |

^aThis salt is presented, for simplicity, as a tris(CH₃OH) solvate; however, as detailed in the experimental section, one CH₃OH molecule refined to an occupancy of 0.88. ^bFor the asymmetric unit; see experimental section.

Table s2. Key interatomic distances (Å) and angles (°) in crystallographically characterized complexes.^a

| atoms | Δ -(<i>S,S</i>)- 3 ³⁺ 3Cl [−] ·2H ₂ O·2CH ₃ OH | Δ -(<i>S,S</i>)- 3 ³⁺ 3Cl [−] ·12.5H ₂ O ^b | Δ -(<i>S,S</i>)- 3 ³⁺ 3Cl [−] ·12.5H ₂ O ^b | Δ -(<i>R,R</i>)- 3 ³⁺ 3Cl [−] ·H ₂ O·3CH ₃ OH |
|-----------|--|--|--|---|
| Co1-N1 | 1.910(2) | 1.967(6) | 1.976(5) | 1.962(3) |
| Co1-N2 | 1.975(3) | 1.961(6) | 1.959(6) | 1.989(3) |
| Co1-N3 | 1.985(2) | 1.954(5) | 1.955(6) | 1.957(3) |
| Co1-N4 | 1.963(2) | 1.958(6) | 1.950(6) | 1.967(3) |
| Co1-N5 | 1.963(3) | 1.963(6) | 1.973(7) | 1.989(3) |
| Co1-N6 | 1.949(3) | 1.949(5) | 1.963(6) | 1.970(3) |
| N1-C1 | 1.492(4) | 1.488(9) | 1.501(9) | 1.484(5) |
| N2-C8 | 1.511(4) | 1.489(9) | 1.505(9) | 1.483(5) |
| N3-C15 | 1.509(3) | 1.475(9) | 1.486(9) | 1.506(5) |
| N4-C22 | 1.502(4) | 1.502(9) | 1.495(9) | 1.511(5) |
| N5-C29 | 1.498(4) | 1.494(9) | 1.491(10) | 1.504(5) |
| N6-C36 | 1.506(4) | 1.508(9) | 1.489(10) | 1.504(5) |
| N1⋯Cl1 | 3.194(3) | 3.3604(1) | - | 3.279(3) |
| N3⋯Cl1 | 3.428(3) | 3.2343(2) | - | 3.194(3) |
| N5⋯Cl1 | 3.269(3) | 3.2762(1) | - | 3.445(3) |
| N2⋯Cl3 | 3.259(3) | 3.2403(1) | 3.3656(1) | 3.208(3) |
| N4⋯Cl3 | 3.198(3) | 3.2363(1) | 3.2062(2) | 3.265(3) |
| N6⋯Cl3 | 3.195(3) | 3.3277(2) | 3.2268(1) | 3.220(3) |
| N5⋯Cl2 | 3.231(3) | 4.2407(2) | - | 3.327(4) |
| N4⋯Cl2 | 3.266(3) | 3.2458(2) | - | 3.202(3) |
| N1H⋯Cl1 | 2.313 | 2.6254 | | 2.437 |
| N3H⋯Cl1 | 2.606 | 2.5448 | | 2.346 |
| N5H⋯Cl1 | 2.436 | 2.5689 | | 2.634 |
| N2H⋯Cl3 | 2.379 | 2.5258 | 2.6708 | 2.324 |
| N4H⋯Cl3 | 2.376 | 2.4211 | 2.5065 | 2.379 |
| N6H⋯Cl3 | 2.364 | 2.6166 | 2.4632 | 2.398 |
| N5H⋯Cl2 | 2.386 | 3.8632 | - | 2.498 |
| N4H⋯Cl2 | 2.387 | 2.4074 | - | 2.356 |
| N3-Co1-N6 | 172.7(1) | 175.2(2) | 174.7(3) | 172.66(14) |
| N1-Co1-N4 | 172.7(1) | 175.5(2) | 175.1(3) | 171.66(13) |
| N2-Co1-N5 | 171.9(1) | 175.3(2) | 175.4(3) | 171.62(14) |

| | | | | |
|------------|---------|---------|---------|-----------|
| N3-Co1-N1 | 90.9(1) | 92.3(2) | 91.8(2) | 90.95(14) |
| N3-Co1-N2 | 96.3(1) | 91.4(2) | 90.8(3) | 94.98(14) |
| N3-Co1-N4 | 84.4(1) | 84.4(2) | 84.4(2) | 84.56(13) |
| N3-Co1-N5 | 90.5(1) | 92.7(2) | 92.2(2) | 91.07(13) |
| N6-Co1-N1 | 94.7(1) | 92.1(2) | 92.3(2) | 94.39(14) |
| N6-Co1-N2 | 89.0(1) | 91.0(2) | 92.8(2) | 90.60(14) |
| N6-Co1-N4 | 90.5(1) | 91.3(2) | 91.7(2) | 90.74(13) |
| N6-Co1-N5 | 84.6(1) | 85.1(2) | 84.5(3) | 83.87(12) |
| N1-Co1-N2 | 84.3(1) | 84.7(2) | 84.8(2) | 83.76(14) |
| N2-Co1-N4 | 90.7(1) | 92.3(2) | 92.2(2) | 89.62(14) |
| N4-Co1-N5 | 94.3(1) | 90.5(2) | 91.5(2) | 96.72(14) |
| N5-Co1-N1 | 91.3(1) | 92.7(2) | 91.6(2) | 90.37(14) |
| N1-H...Cl1 | 165.5 | 137.35 | | 155.7 |
| N3-H...Cl1 | 152.4 | 132.09 | | 156.9 |
| N5-H...Cl1 | 154.0 | 134.07 | | 150.5 |
| N2-H...Cl3 | 165.8 | 134.78 | 132.9 | 161.2 |
| N4-H...Cl3 | 152.2 | 147.71 | 133.1 | 168.2 |
| N6-H...Cl3 | 153.6 | 134.60 | 140.5 | 151.9 |
| N5-H...Cl2 | 156.3 | 108.14 | - | 153.2 |
| N4-H...Cl2 | 165.5 | 151.51 | - | 156.6 |

^a The atoms are numbered such that for each salt, N1, N3, and N5 define one of the C₃ NH faces to which a chloride anion is hydrogen bonded, and N2, N4, and N6 the other.

^b These values are for one of the two independent trications in the unit cell. These are distinguished by a different H₂O hydrogen bonding pattern.

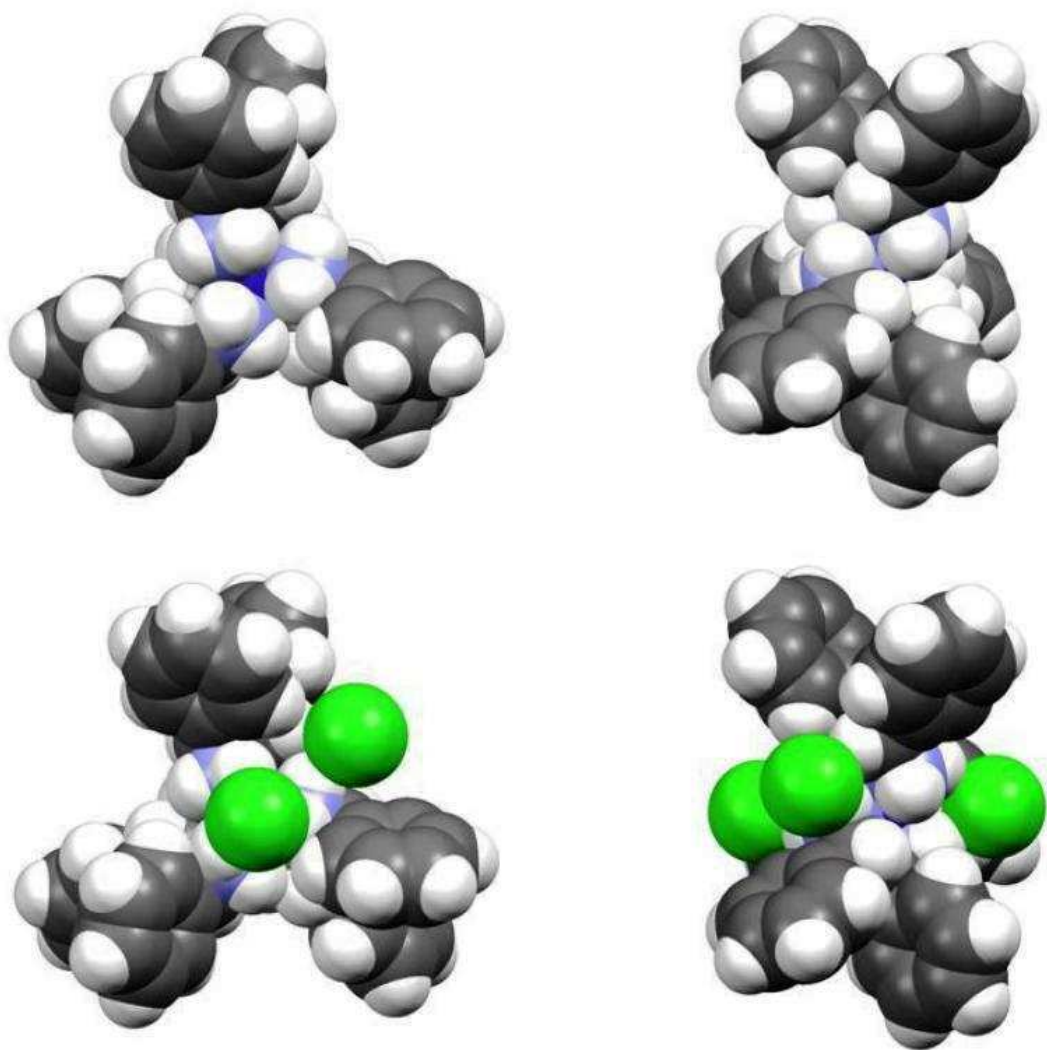


Figure s1. Thermal ellipsoid diagram (50% probability level) of $\Delta\text{-(S,S)-3}^{3+} \cdot 3\text{Cl}^{-} \cdot 12.5\text{H}_2\text{O}$ with solvent molecules removed for clarity. Upper left and right, views down the idealized C_3 and C_2 axes with chloride ions omitted; lower left and right, analogous views with chloride ions.

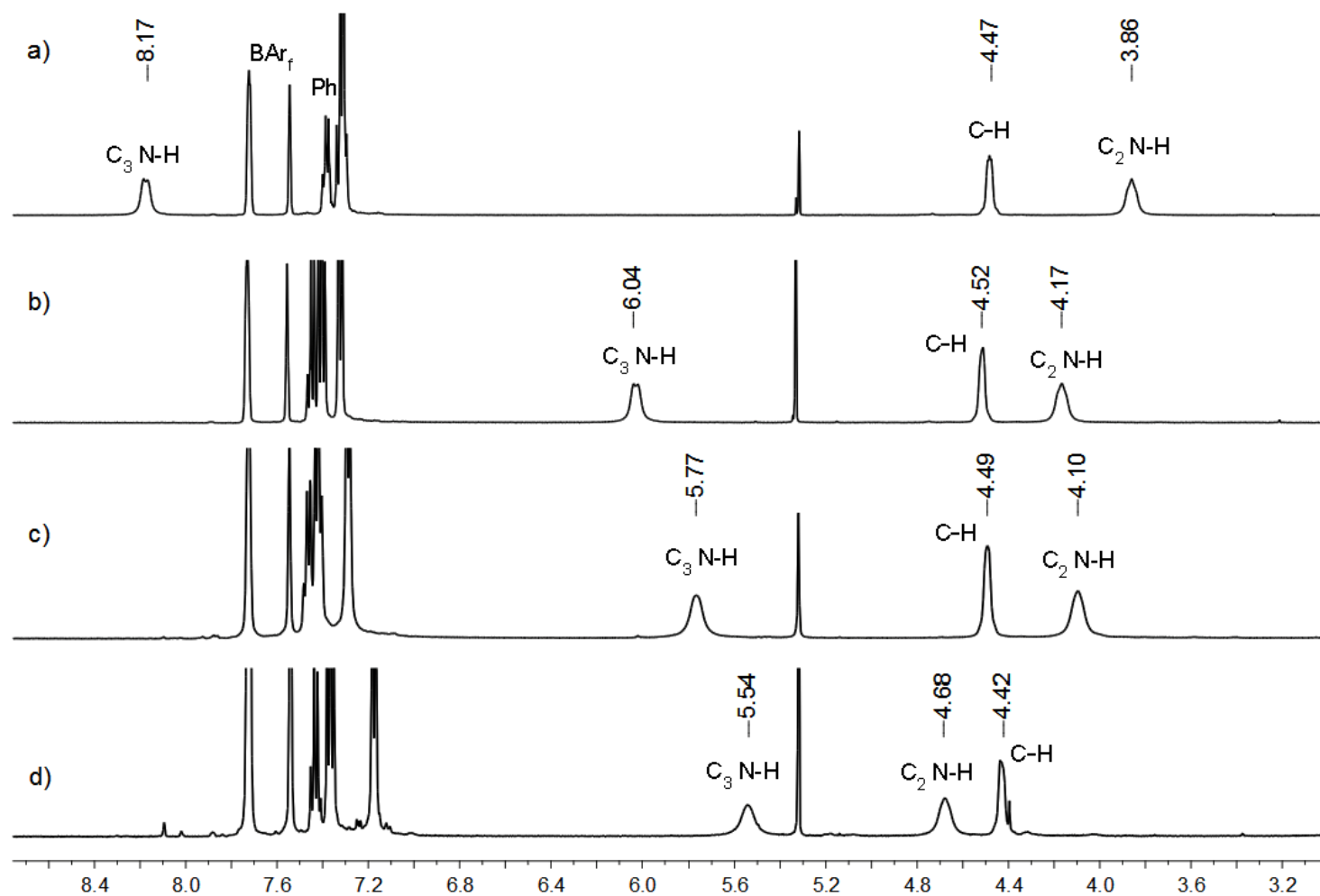


Figure s2. ^1H NMR spectra (CD_2Cl_2) of (a) Λ -(*S,S*)- $\mathbf{3}^{3+} \cdot 2\text{Cl}^- \cdot \text{BArf}^-$, (b) Λ -(*S,S*)- $\mathbf{3}^{3+} \cdot 2\text{BF}_4^- \cdot \text{BArf}^-$, (c) Λ -(*S,S*)- $\mathbf{3}^{3+} \cdot 2\text{PF}_6^- \cdot \text{BArf}^-$, and (d) Λ -(*S,S*)- $\mathbf{3}^{3+} \cdot 3\text{BArf}^-$.

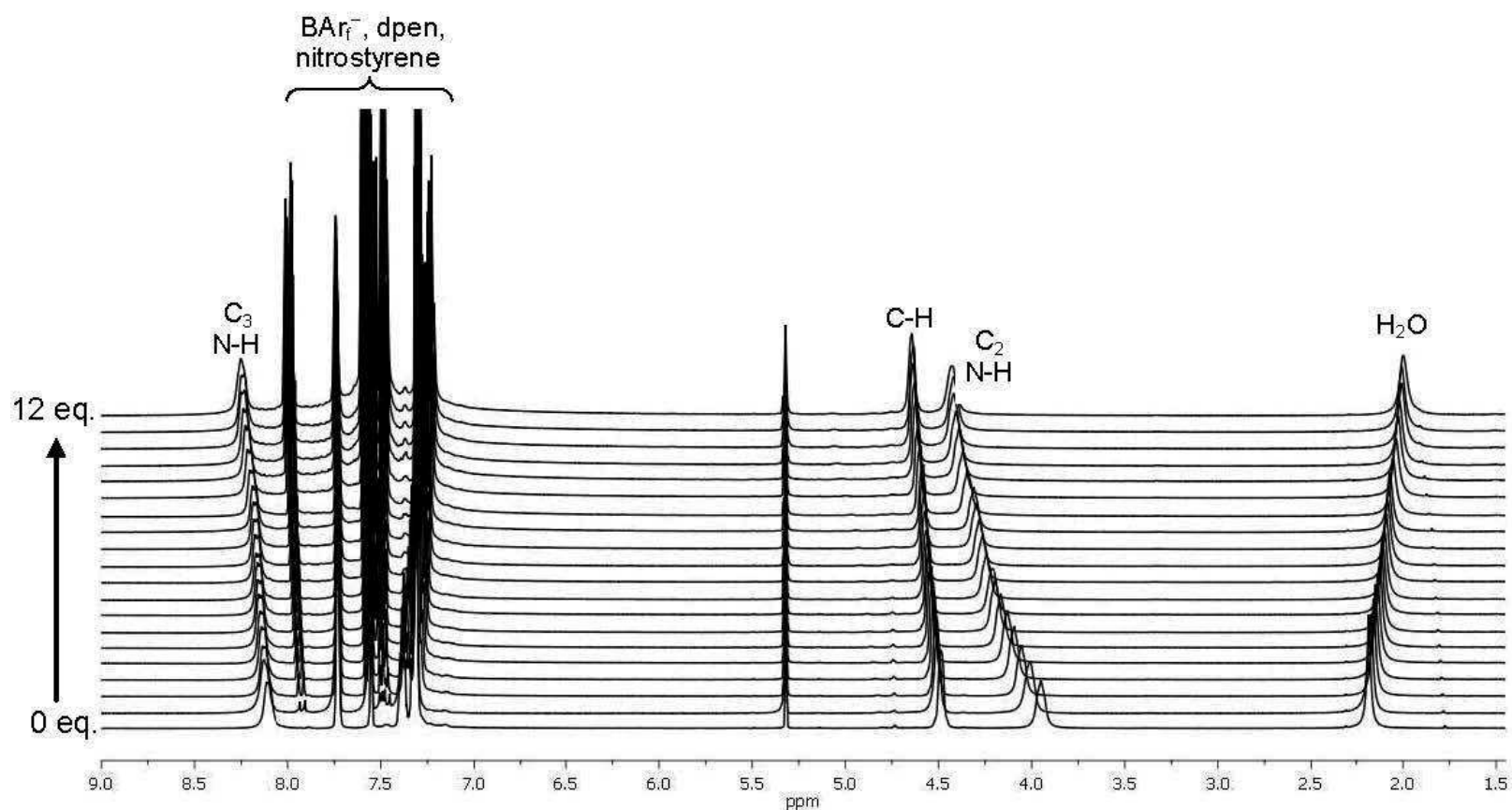
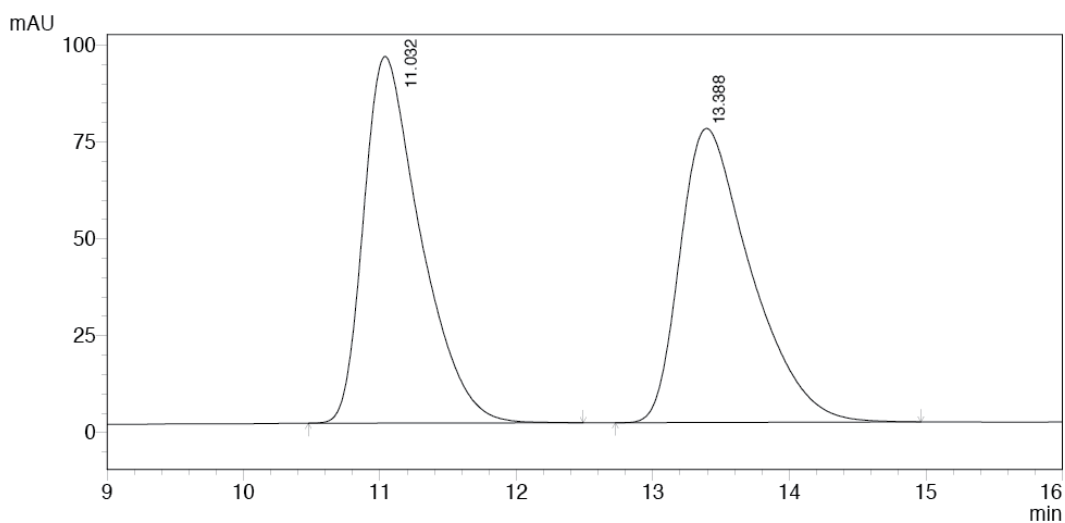


Figure s3. ^1H NMR spectra: titration of a CD_2Cl_2 solution of $\Lambda\text{-(S,S)-}\mathbf{3}^{3+} 2\text{Cl}^- \text{BArF}^-$ with *trans*- β -nitrostyrene (**4a**). Data (equiv of **4a**/NH signals, δ in ppm): 0.25/4.010 and 8.127; 0.50/4.053 and 8.134; 0.75/4.094 and 8.141; 1.0/4.129 and 8.131; 1.25/4.165 and 8.155; 1.50/4.195 and 8.158; 1.75/4.204 and 8.162; 2.0/4.241 and 8.169; 2.50/4.28 and 8.179; 2.75/4.296 and 8.183; 3.0/4.31 and 8.178; 4.0/4.342 and 8.184; 5.0/4.368 and 8.196; 6.0/4.387 and 8.227; 7.0/4.405 and 8.236; 8.0/4.418 and 8.234; 9.0/4.432 and 8.249.

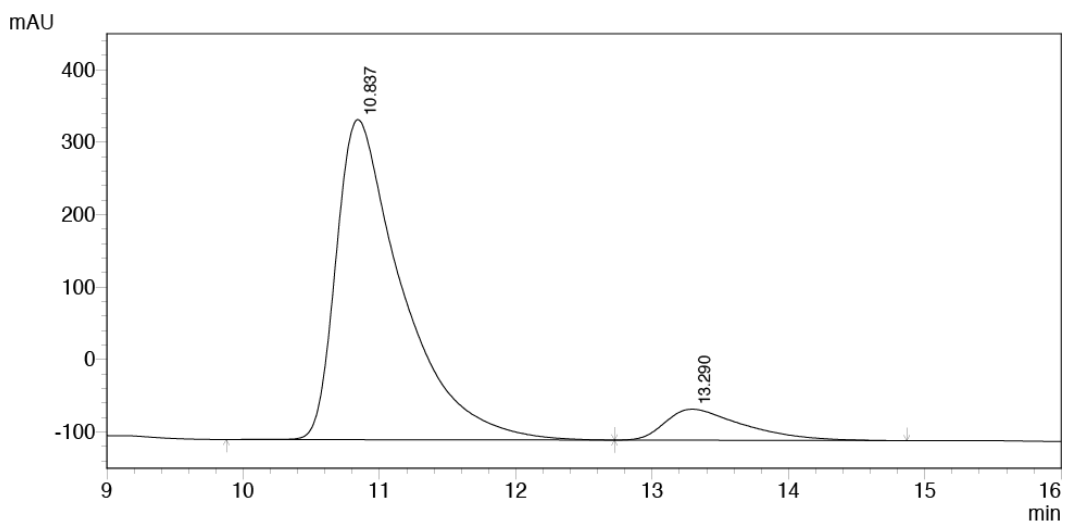
Diisopropyl 2-(2-nitro-1-phenylethyl)malonate (6aa**).**

(a)



| PDA 220nm | | | | | |
|-----------|-----------|---------|--------|---------|----------|
| Peak# | Ret. Time | Area | Height | Area % | Height % |
| 1 | 11.032 | 2745001 | 94604 | 49.931 | 55.488 |
| 2 | 13.388 | 2752610 | 75890 | 50.069 | 44.512 |
| Total | | 5497611 | 170494 | 100.000 | 100.000 |

(b)

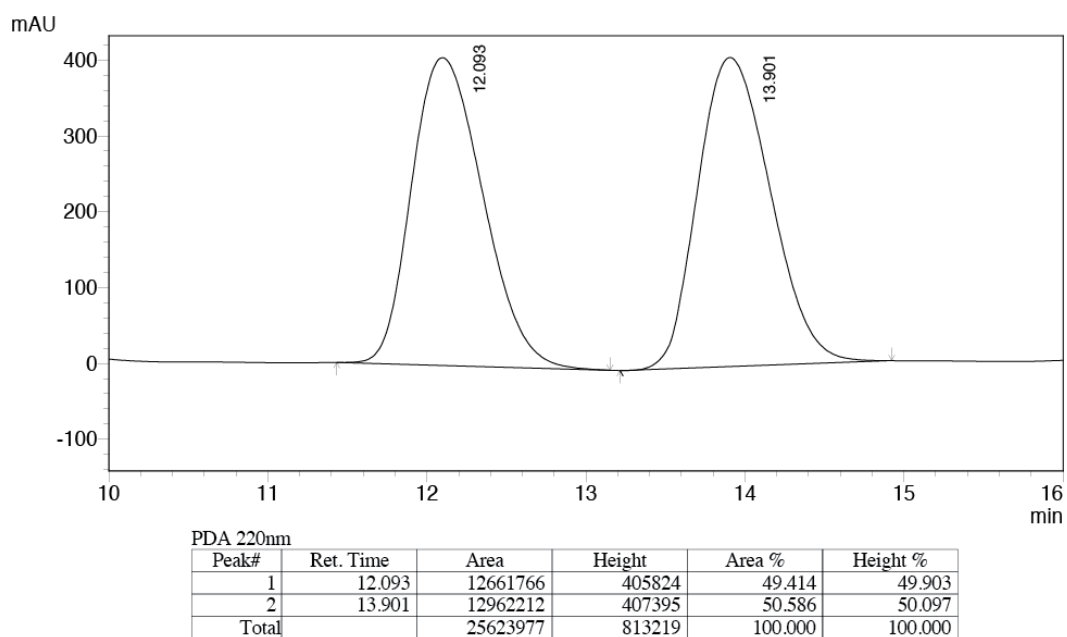


| PDA 220nm | | | | | |
|-----------|-----------|----------|--------|---------|----------|
| Peak# | Ret. Time | Area | Height | Area % | Height % |
| 1 | 10.837 | 14897274 | 441745 | 89.745 | 91.162 |
| 2 | 13.290 | 1702201 | 42826 | 10.255 | 8.838 |
| Total | | 16599476 | 484571 | 100.000 | 100.000 |

Figure s4. HPLC traces of **6aa** (Scheme 3, entry 1): (a) racemic sample; (b) catalyzed by Λ -(*S,S*)-**3**³⁺ 2Cl⁻BAr_f⁻.

Diethyl 2-(2-nitro-1-phenylethyl)malonate (6ab**).**

(a)



(b)

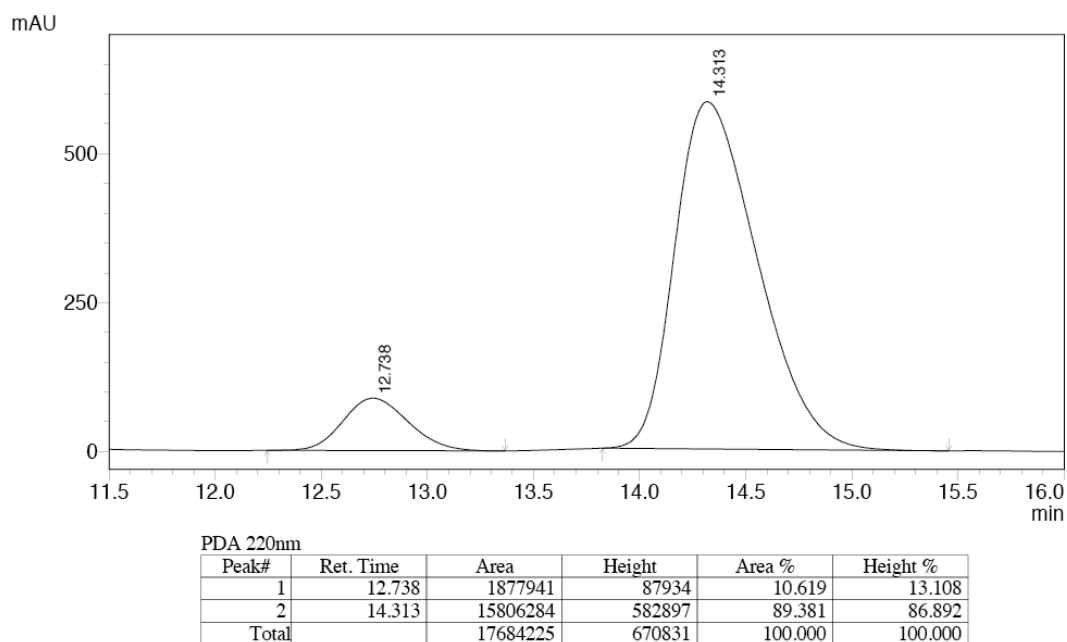
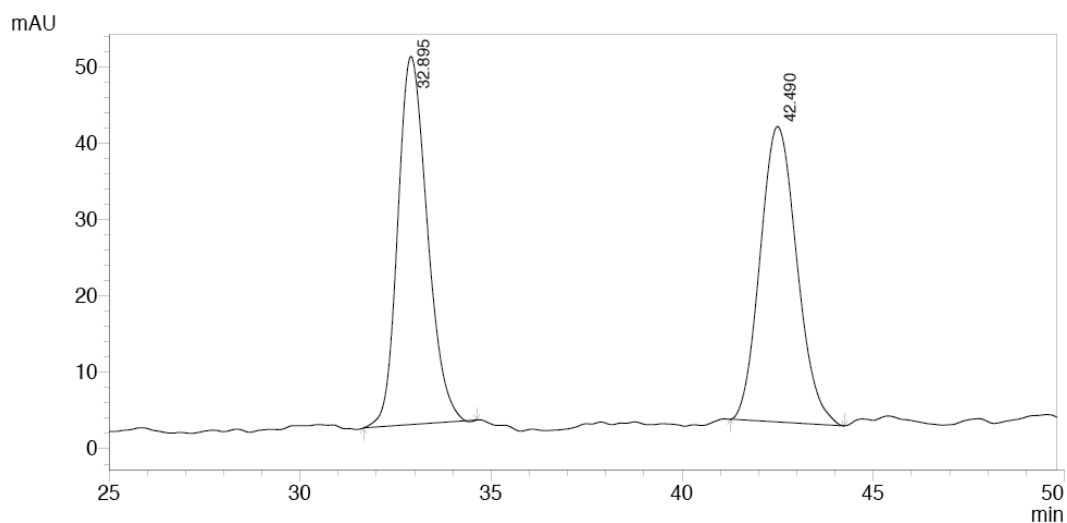


Figure s5. HPLC traces of **6ab** (Scheme 3, entry 3): (a) racemic sample; (b) catalyzed by Λ -(*S,S*)-**3**³⁺ 2Cl⁻BARf⁻.

Dimethyl 2-(2-nitro-1-phenylethyl)malonate (6ac).

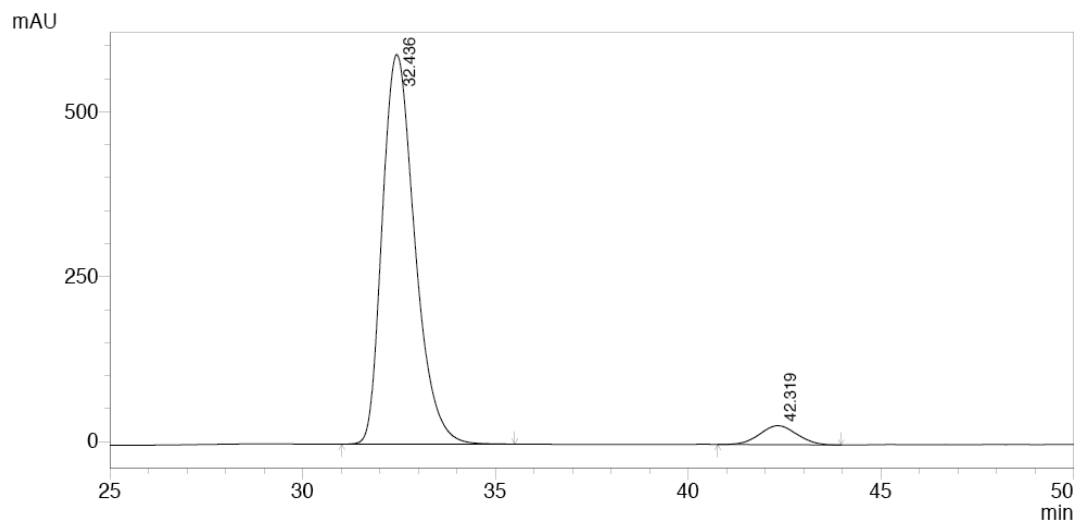
(a)



PDA 220nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|---------|--------|---------|----------|
| 1 | 32.895 | 2592054 | 48187 | 49.983 | 55.472 |
| 2 | 42.490 | 2593793 | 38681 | 50.017 | 44.528 |
| Total | | 5185847 | 86867 | 100.000 | 100.000 |

(b)



PDA 220nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 32.436 | 34674163 | 590859 | 94.706 | 95.343 |
| 2 | 42.319 | 1938180 | 28862 | 5.294 | 4.657 |
| Total | | 36612343 | 619722 | 100.000 | 100.000 |

(c)

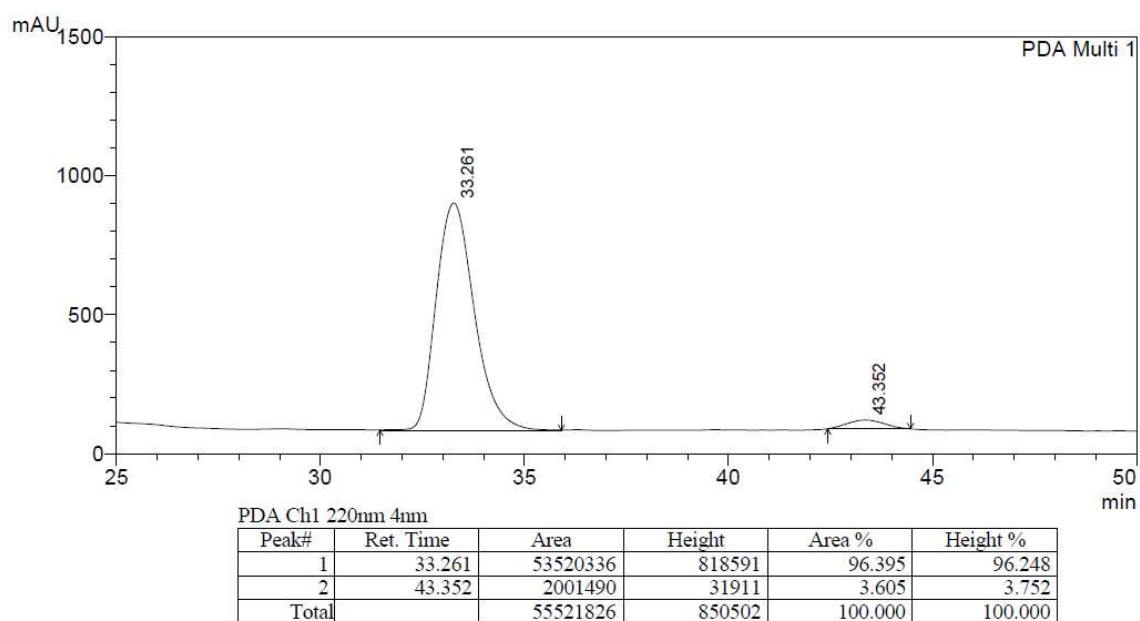
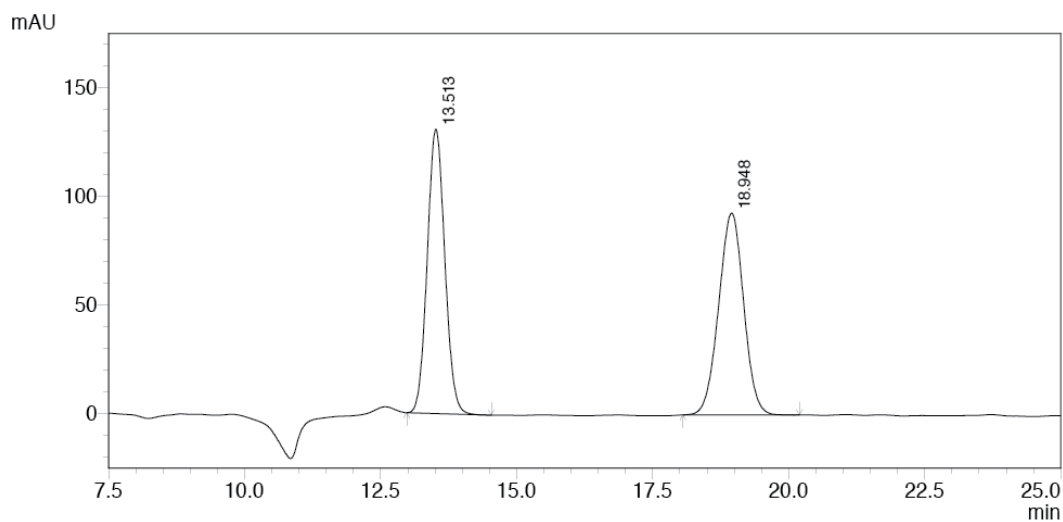


Figure s6. HPLC traces of **6ac** (Scheme 4, entry 1): (a) racemic sample; (b) catalyzed by Λ -(*S,S*)- $3^{3+} 2BF_4^-BAR_f^-$; (c) catalyzed by Λ -(*S,S*)- $3^{3+} 3BF_4^-$.

Dimethyl 2-(2-nitro-1- α -naphthylethyl)malonate (6b_c).

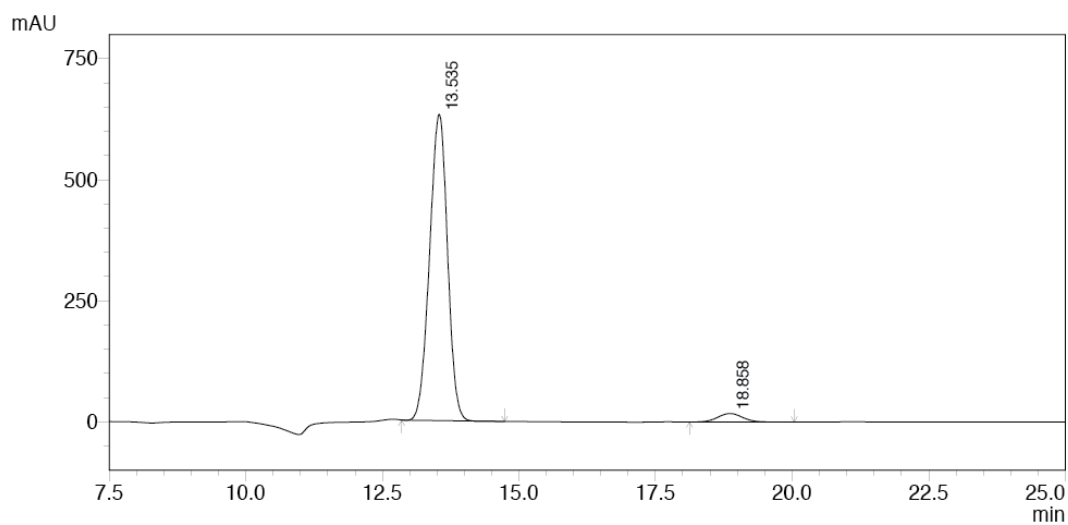
(a)



PDA 254nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|---------|--------|---------|----------|
| 1 | 13.513 | 2941994 | 130912 | 49.780 | 58.498 |
| 2 | 18.948 | 2967956 | 92876 | 50.220 | 41.502 |
| Total | | 5909950 | 223789 | 100.000 | 100.000 |

(b)



PDA 254nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 13.535 | 14427103 | 632667 | 96.261 | 97.288 |
| 2 | 18.858 | 560393 | 17639 | 3.739 | 2.712 |
| Total | | 14987496 | 650305 | 100.000 | 100.000 |

(c)

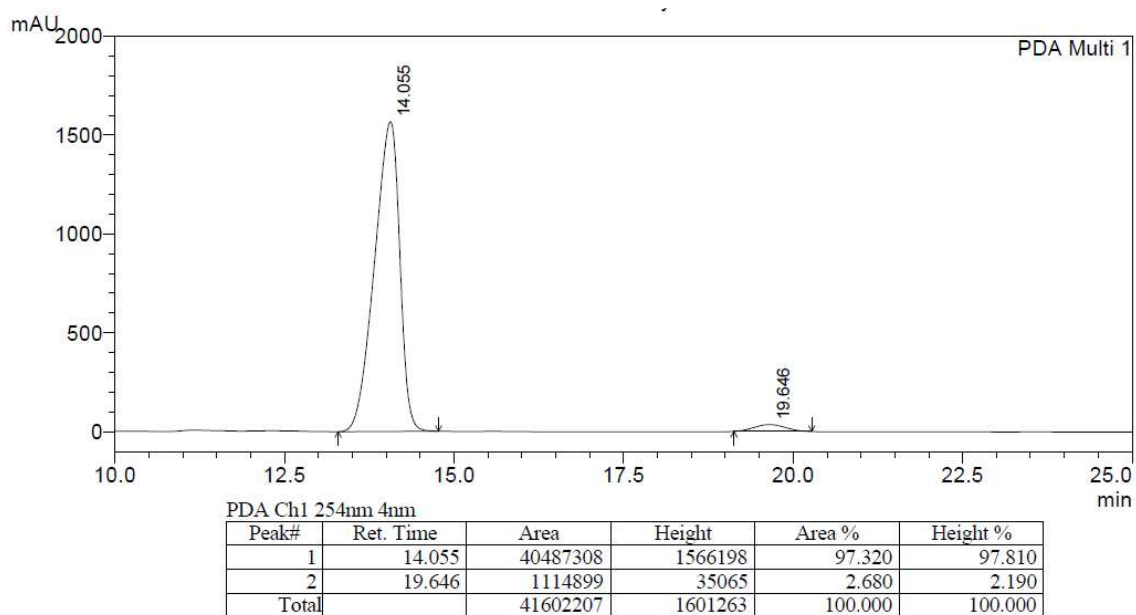
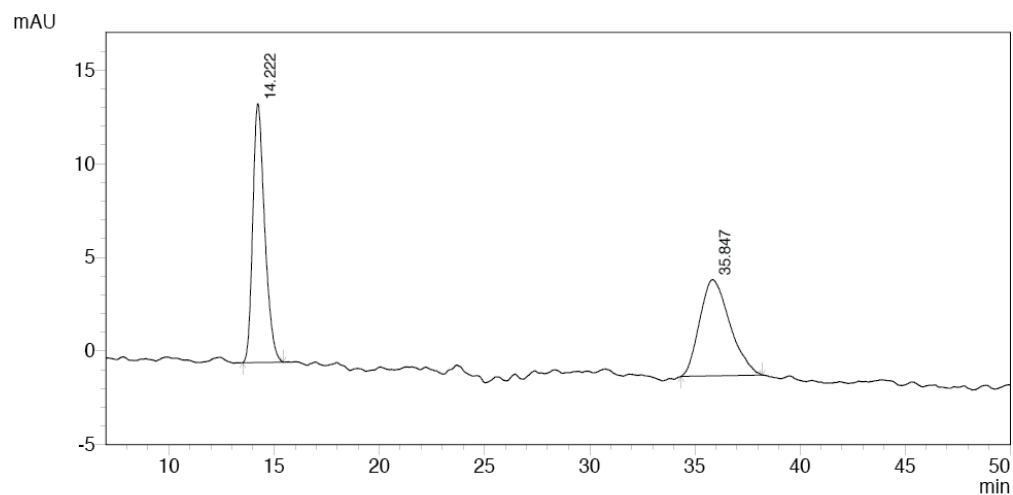


Figure s7. HPLC traces of **6bc** (Scheme 4, entry 2): (a) racemic sample; (b) catalyzed by Λ -(*S,S*)- $\mathbf{3}^{3+} 2\text{BF}_4^- \text{BAr}_f^-$; (c) catalyzed by Λ -(*S,S*)- $\mathbf{3}^{3+} 3\text{BF}_4^-$.

Dimethyl 2-(2-nitro-1- β -naphthylethyl)malonate (6cc).

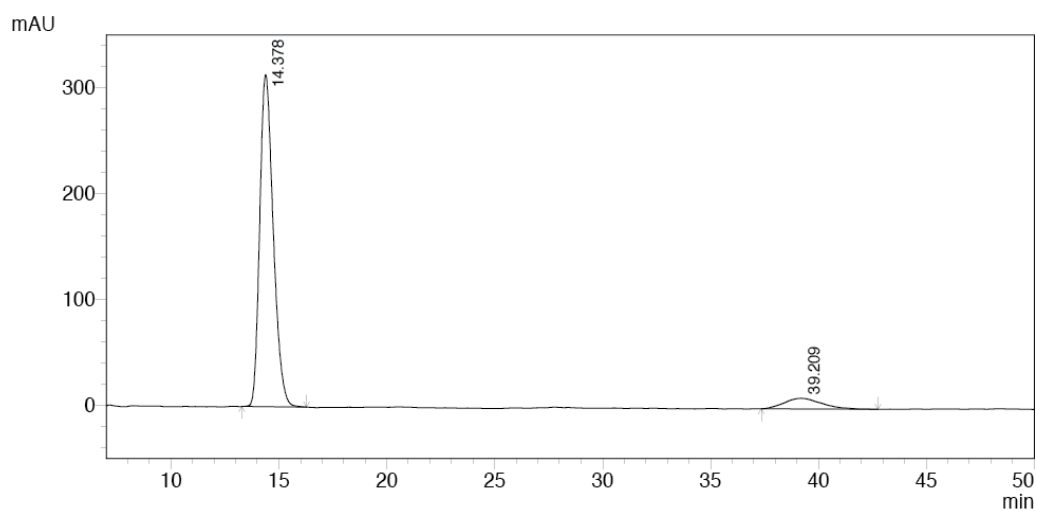
(a)



PDA 254nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|---------|--------|---------|----------|
| 1 | 14.222 | 545650 | 13812 | 51.472 | 72.833 |
| 2 | 35.847 | 514433 | 5152 | 48.528 | 27.167 |
| Total | | 1060083 | 18965 | 100.000 | 100.000 |

(b)



PDA 254nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 14.378 | 13650190 | 313478 | 91.924 | 96.896 |
| 2 | 39.209 | 1199200 | 10041 | 8.076 | 3.104 |
| Total | | 14849390 | 323518 | 100.000 | 100.000 |

(c)

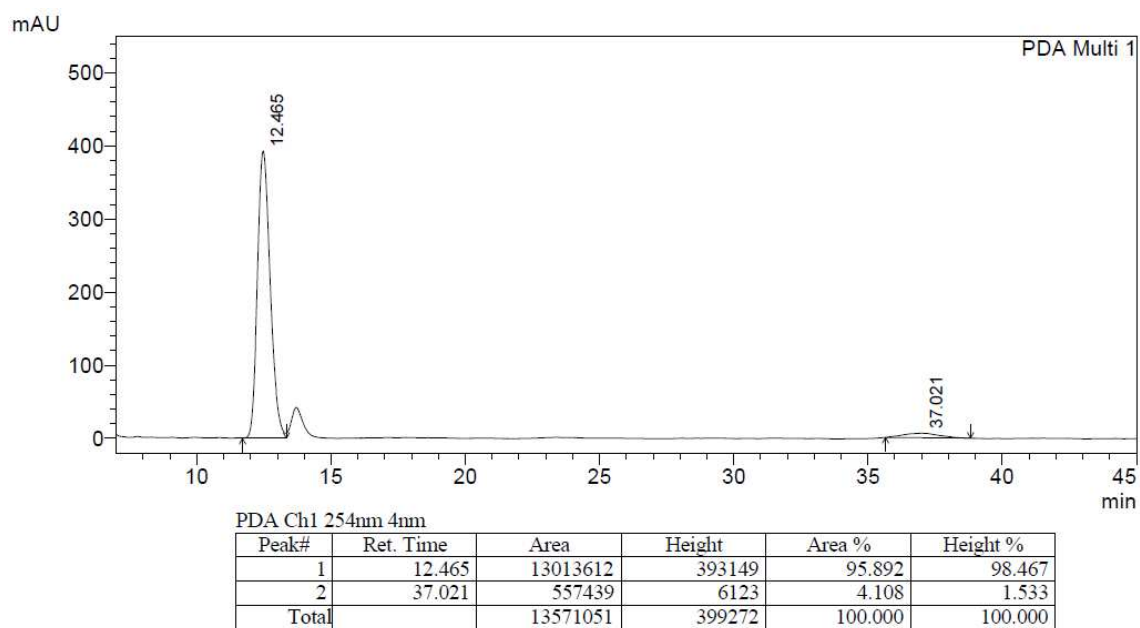
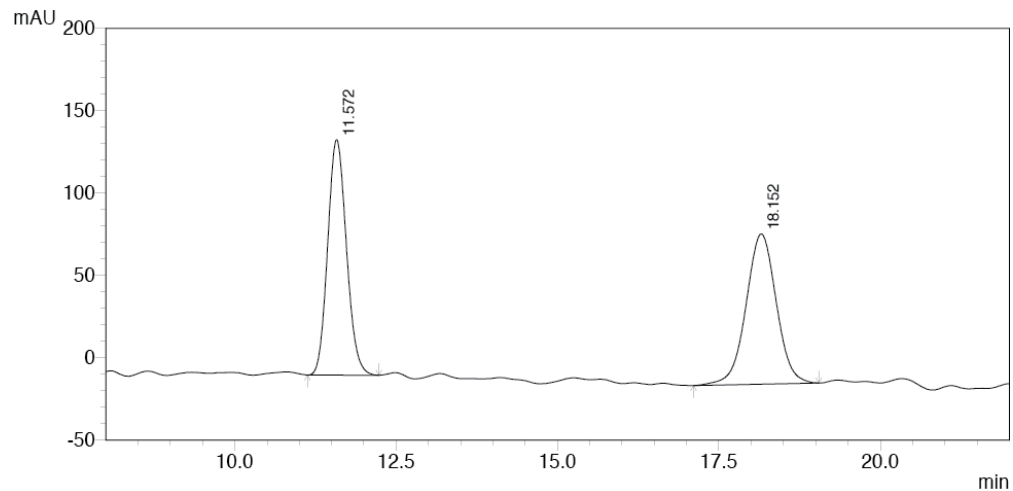


Figure s8. HPLC traces of **6cc** (Scheme 4, entry 3): (a) racemic sample; (b) catalyzed by Λ -(*S,S*)-**3**³⁺ 2BF₄⁻BARf⁻; (c) catalyzed by Λ -(*S,S*)-**3**³⁺ 3BF₄⁻ (older column).

Dimethyl 2-(2-nitro-1-(4-methoxyphenyl)ethyl)malonate (6dc)

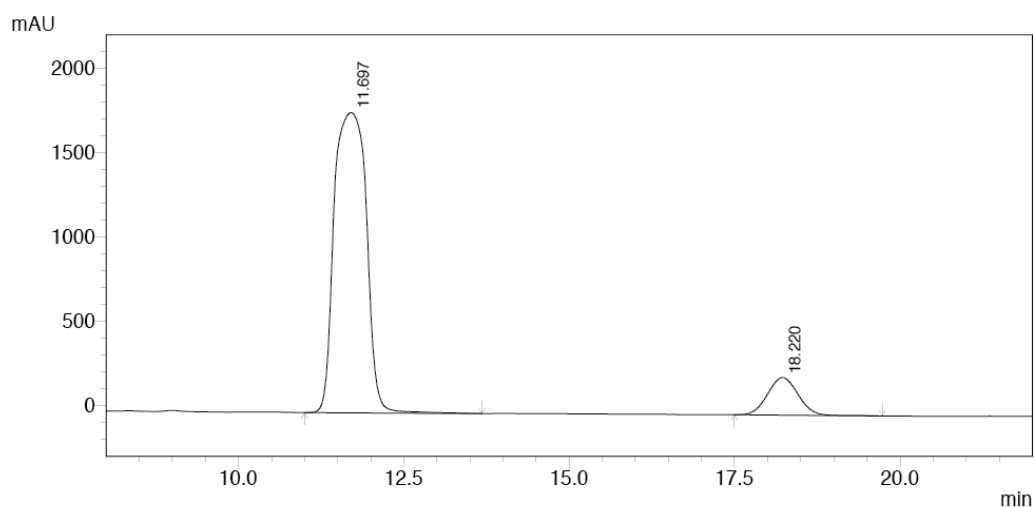
(a)



PDA 220nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|---------|--------|---------|----------|
| 1 | 11.572 | 2892734 | 142917 | 49.286 | 61.046 |
| 2 | 18.152 | 2976564 | 91196 | 50.714 | 38.954 |
| Total | | 5869298 | 234114 | 100.000 | 100.000 |

(b)



PDA 220nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|---------|---------|----------|
| 1 | 11.697 | 60314078 | 1781389 | 89.351 | 88.845 |
| 2 | 18.220 | 7188616 | 223672 | 10.649 | 11.155 |
| Total | | 67502694 | 2005061 | 100.000 | 100.000 |

(c)

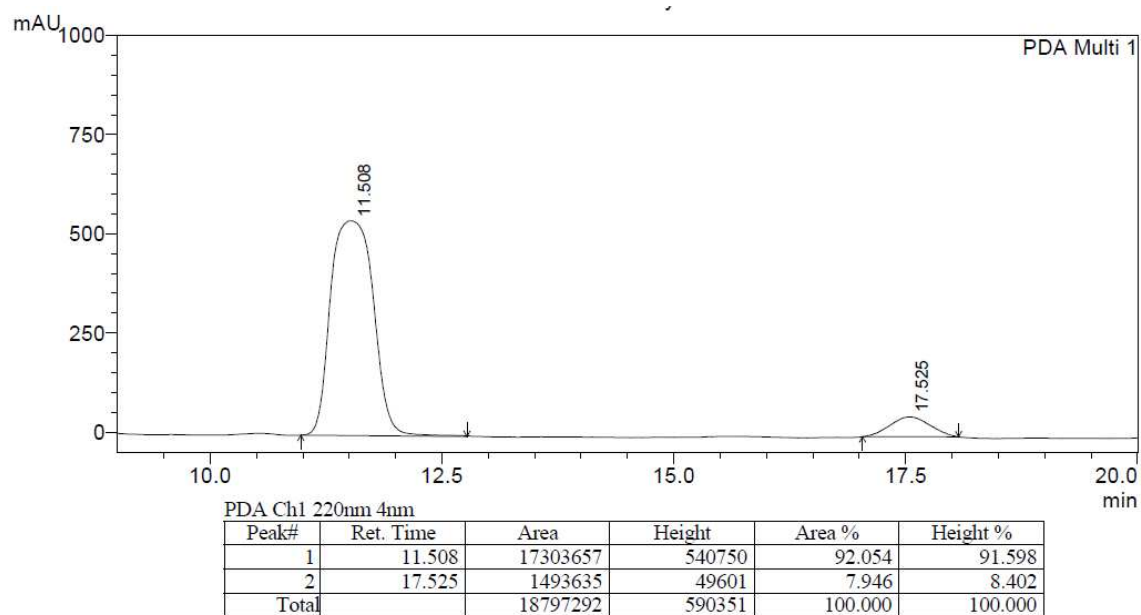
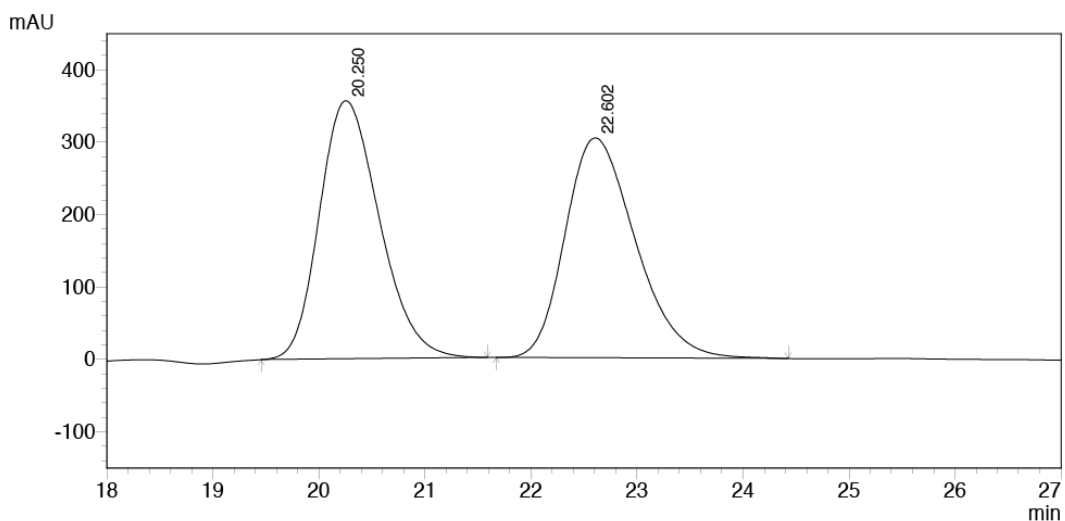


Figure s9. HPLC traces of **6dc** (Scheme 4, entry 4): (a) racemic sample; (b) catalyzed by Λ -(*S,S*)-**3**³⁺ 2BF₄⁻BARf⁻; (c) catalyzed by Λ -(*S,S*)-**3**³⁺ 3BF₄⁻.

Dimethyl 2-(2-nitro-1-(3,4-dichlorophenyl)ethyl)malonate (6ec).

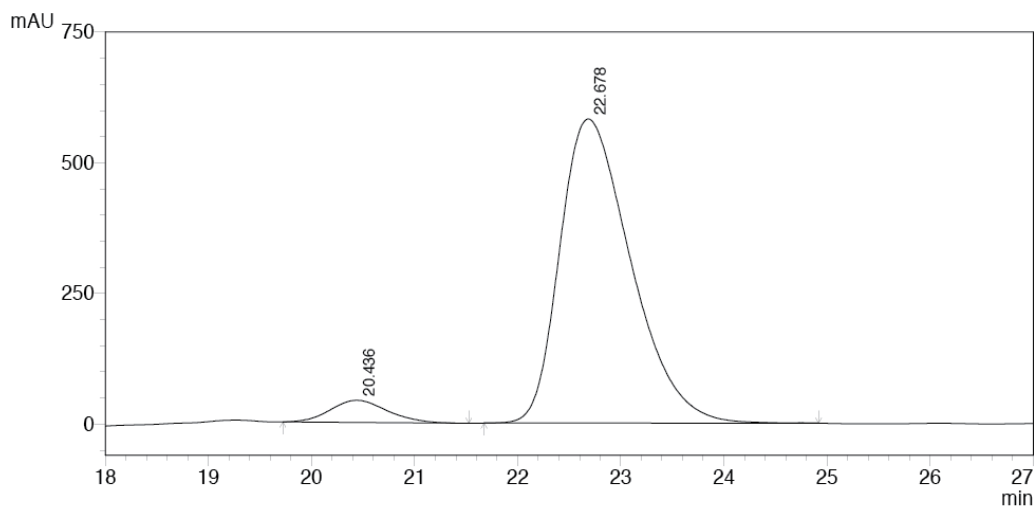
(a)



PDA 220nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 20.250 | 14034598 | 356495 | 50.085 | 53.998 |
| 2 | 22.602 | 13986920 | 303708 | 49.915 | 46.002 |
| Total | | 28021519 | 660203 | 100.000 | 100.000 |

(b)



PDA 220nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 20.436 | 1637581 | 42354 | 5.544 | 6.785 |
| 2 | 22.678 | 27899733 | 581886 | 94.456 | 93.215 |
| Total | | 29537314 | 624240 | 100.000 | 100.000 |

(c)

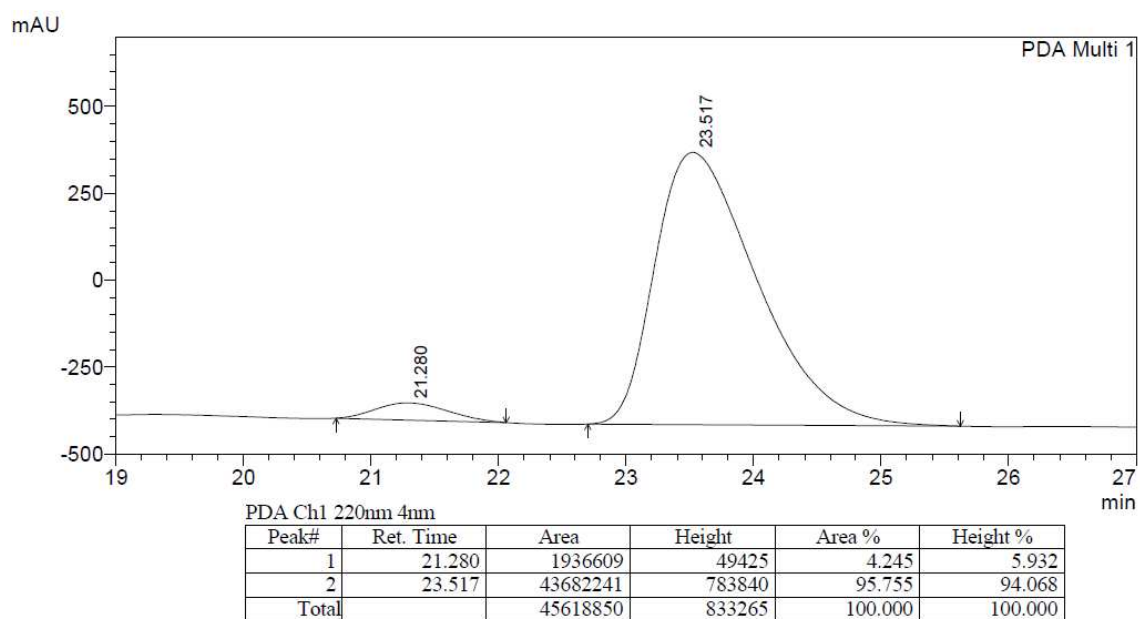
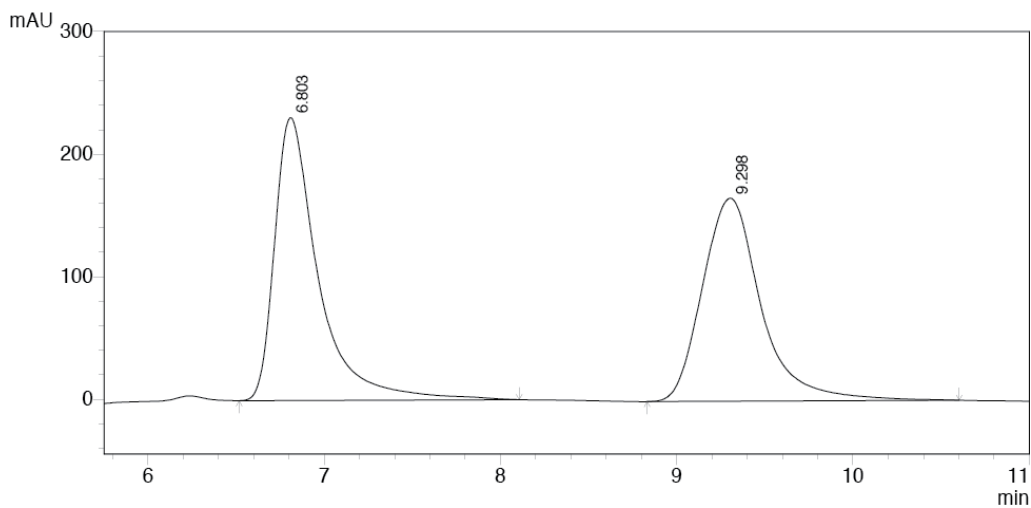


Figure s10. HPLC traces of **6ec** (Scheme 4, entry 5): (a) racemic sample; (b) catalyzed by Λ -(*S,S*)-**3**³⁺ 2BF₄⁻BAr_f⁻; (c) catalyzed by Λ -(*S,S*)-**3**³⁺ 3BF₄⁻.

Dimethyl 2-(2-nitro-1-(pyridine-3-yl)ethyl)malonate (6fc**).**

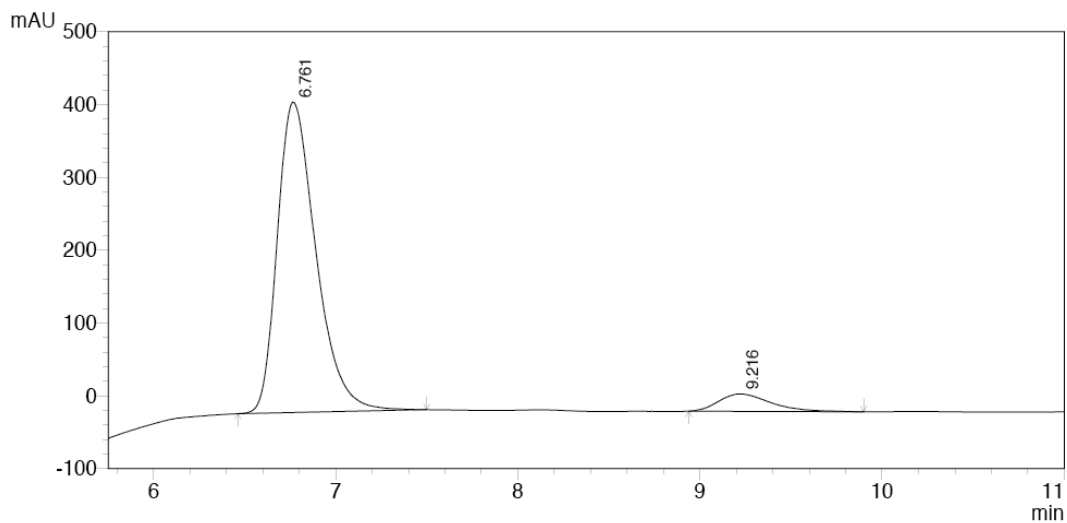
(a)



PDA 220nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|---------|--------|---------|----------|
| 1 | 6.803 | 3983767 | 230889 | 50.865 | 58.208 |
| 2 | 9.298 | 3848303 | 165770 | 49.135 | 41.792 |
| Total | | 7832070 | 396659 | 100.000 | 100.000 |

(b)



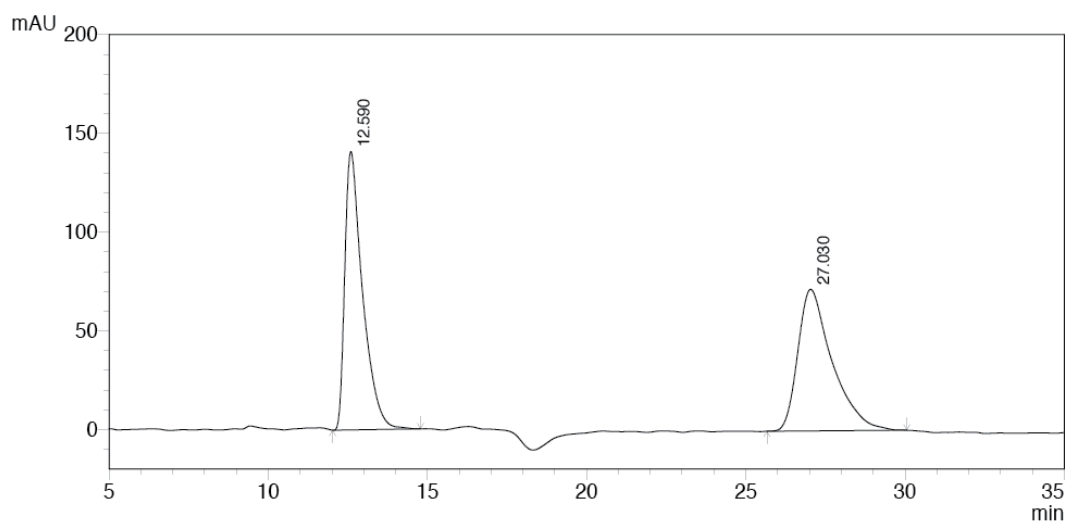
PDA 220nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|---------|--------|---------|----------|
| 1 | 6.761 | 6296885 | 426260 | 93.049 | 94.722 |
| 2 | 9.216 | 470397 | 23751 | 6.951 | 5.278 |
| Total | | 6767282 | 450011 | 100.000 | 100.000 |

Figure s11. HPLC traces of **6fc** (Scheme 4, entry 6): (a) racemic sample; (b) catalyzed by Λ -(*S,S*)- $3^{3+} 2\text{BF}_4^- \text{BAr}_f^-$.

Dimethyl 2-(2-nitro-1-(2-trifluoromethylphenyl)ethyl)malonate (6gc).

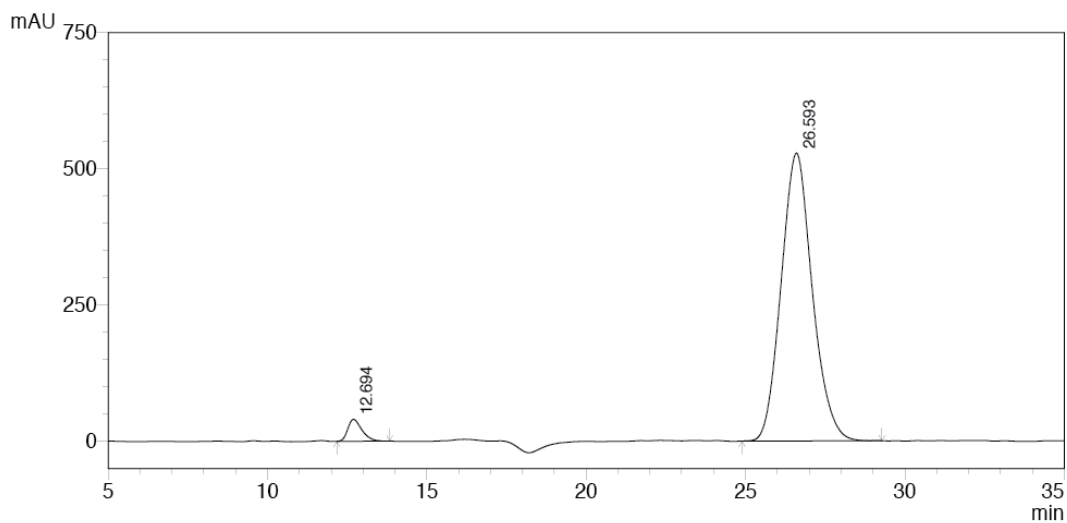
(a)



PDA 220nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 12.590 | 5460013 | 141005 | 50.419 | 66.304 |
| 2 | 27.030 | 5369188 | 71660 | 49.581 | 33.696 |
| Total | | 10829201 | 212664 | 100.000 | 100.000 |

(b)



PDA 220nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 12.694 | 1238157 | 40259 | 3.339 | 7.083 |
| 2 | 26.593 | 35848703 | 528088 | 96.661 | 92.917 |
| Total | | 37086861 | 568347 | 100.000 | 100.000 |

(c)

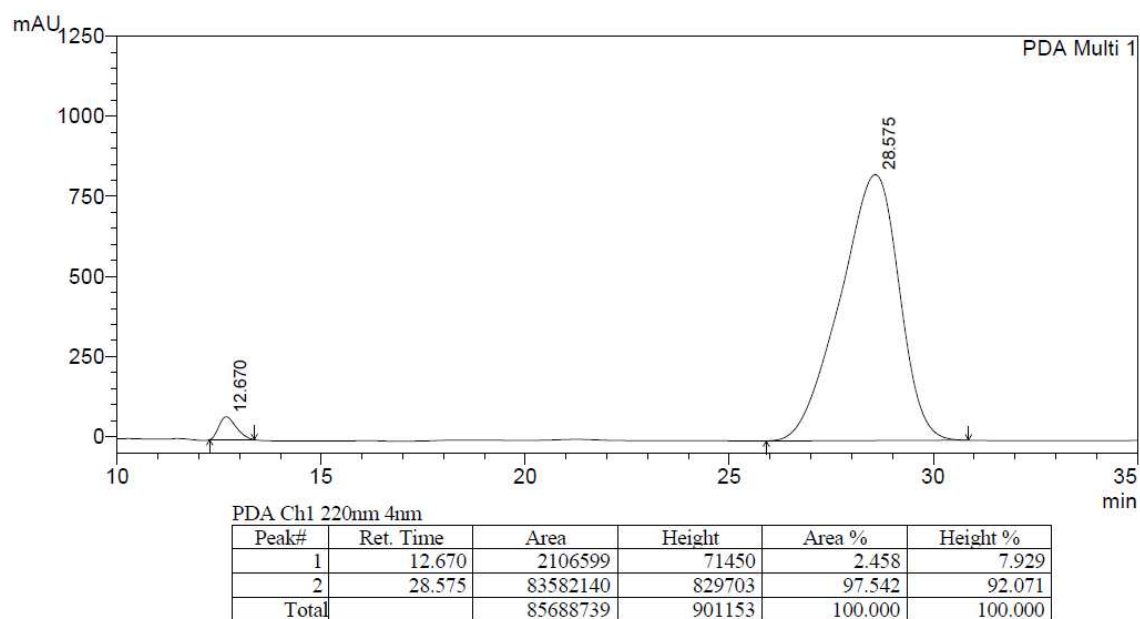
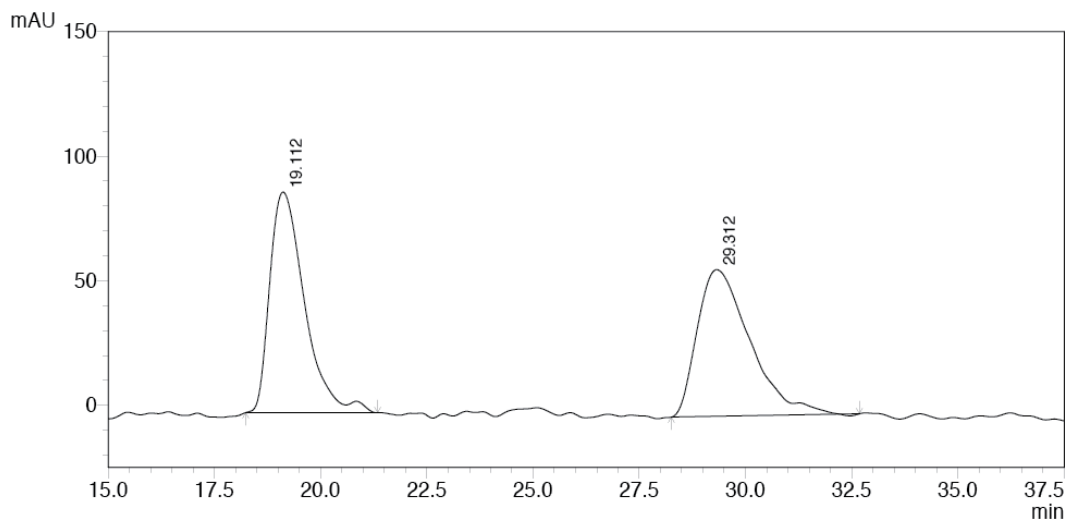


Figure s12. HPLC traces of **6gc** (Scheme 4, entry 7): (a) racemic sample; (b) catalyzed by Λ -(*S,S*)- $\mathbf{3}^{3+}$ $2\text{BF}_4^- \text{BAr}_f^-$; (c) catalyzed by Λ -(*S,S*)- $\mathbf{3}^{3+}$ 3BF_4^- .

Dimethyl 2-(2-nitro-1-(2-acetoxyphenyl)ethyl)malonate (6hc).

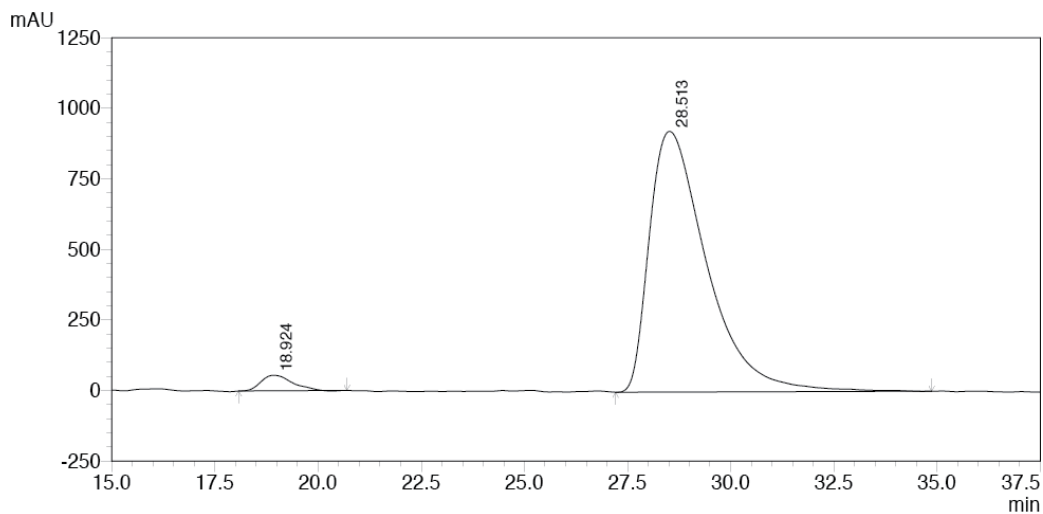
(a)



PDA 220nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 19.112 | 5040868 | 88545 | 49.885 | 60.058 |
| 2 | 29.312 | 5064198 | 58886 | 50.115 | 39.942 |
| Total | | 10105066 | 147431 | 100.000 | 100.000 |

(b)



PDA 220nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 18.924 | 2878634 | 55143 | 3.121 | 5.635 |
| 2 | 28.513 | 89360381 | 923530 | 96.879 | 94.365 |
| Total | | 92239015 | 978673 | 100.000 | 100.000 |

(c)

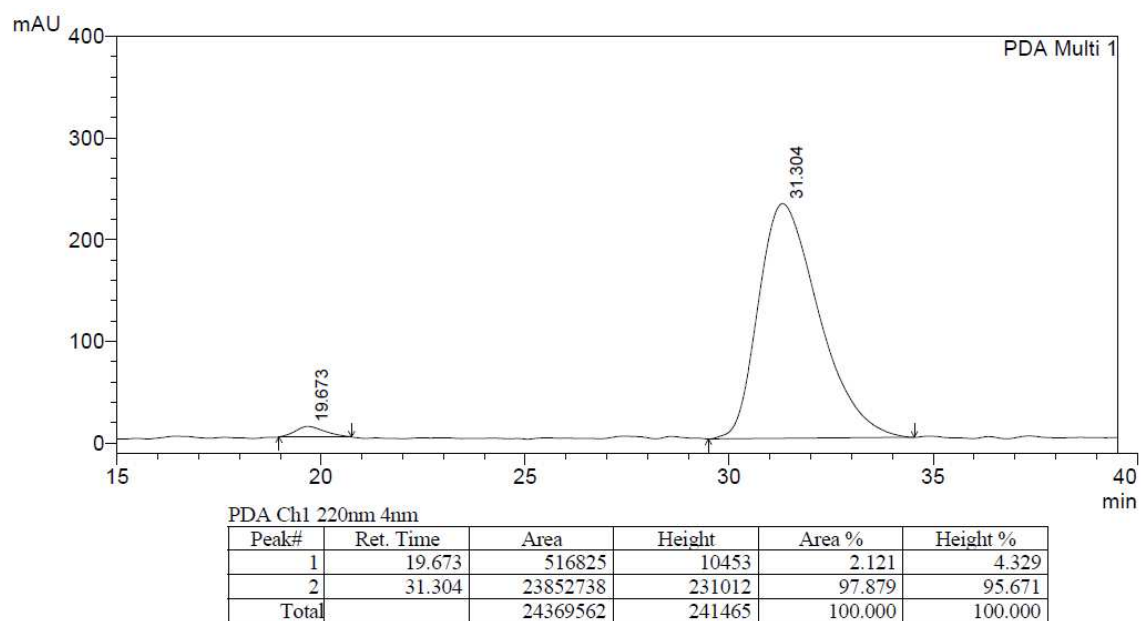
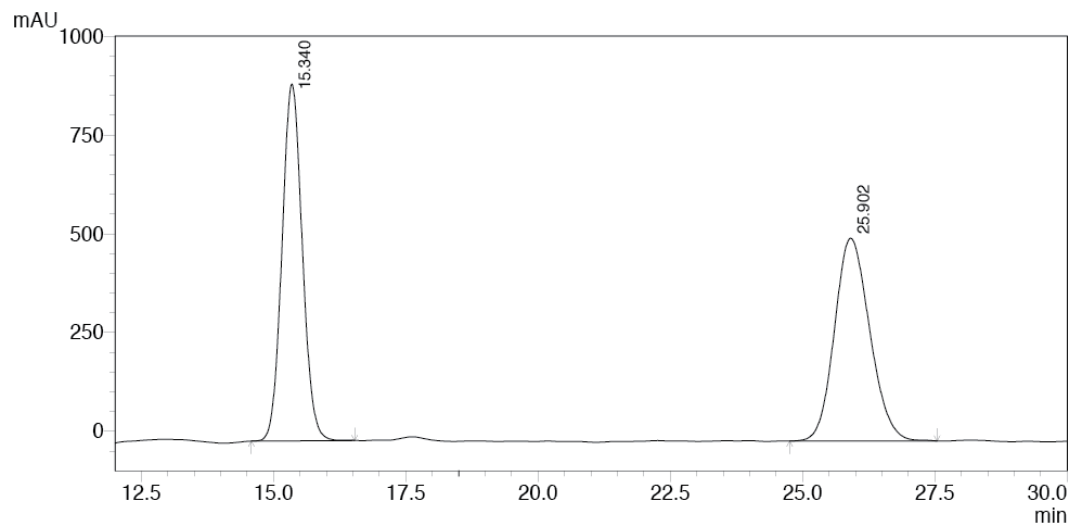


Figure s13. HPLC traces of **6hc** (Scheme 4, entry 8): (a) racemic sample; (b) catalyzed by Λ -(*S,S*)-**3**³⁺ 2BF₄⁻BAr_f⁻; (c) catalyzed by Λ -(*S,S*)-**3**³⁺ 3BF₄⁻.

Dimethyl 2-(2-nitro-1-(2-benzoyloxyphenyl)ethyl)malonate (6ic).

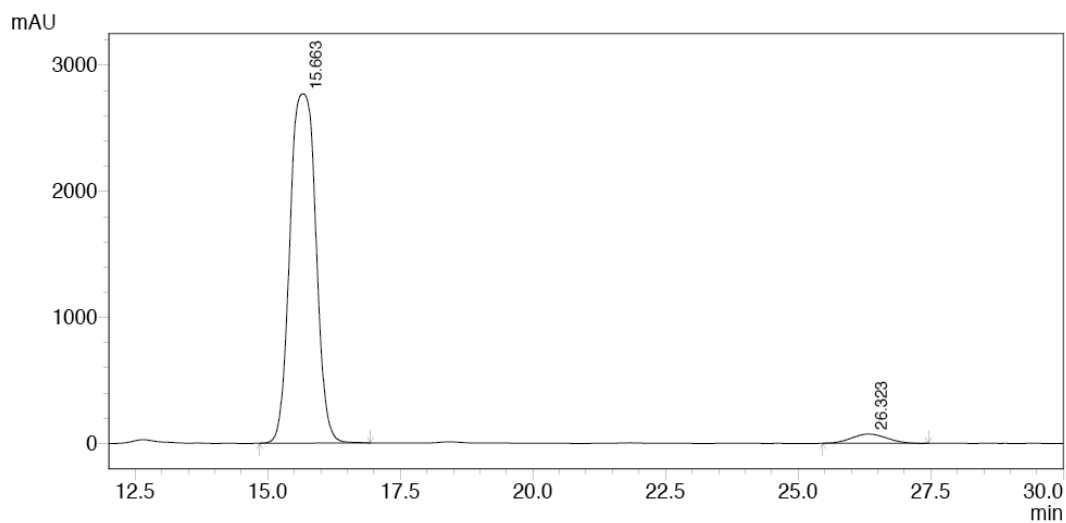
(a)



PDA220nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|---------|---------|----------|
| 1 | 15.340 | 24105156 | 903431 | 50.293 | 63.762 |
| 2 | 25.902 | 23824081 | 513442 | 49.707 | 36.238 |
| Total | | 47929237 | 1416873 | 100.000 | 100.000 |

(b)



PDA 220nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|---------|---------|----------|
| 1 | 15.663 | 94240443 | 2770460 | 96.556 | 97.428 |
| 2 | 26.323 | 3360915 | 73149 | 3.444 | 2.572 |
| Total | | 97601359 | 2843609 | 100.000 | 100.000 |

(c)

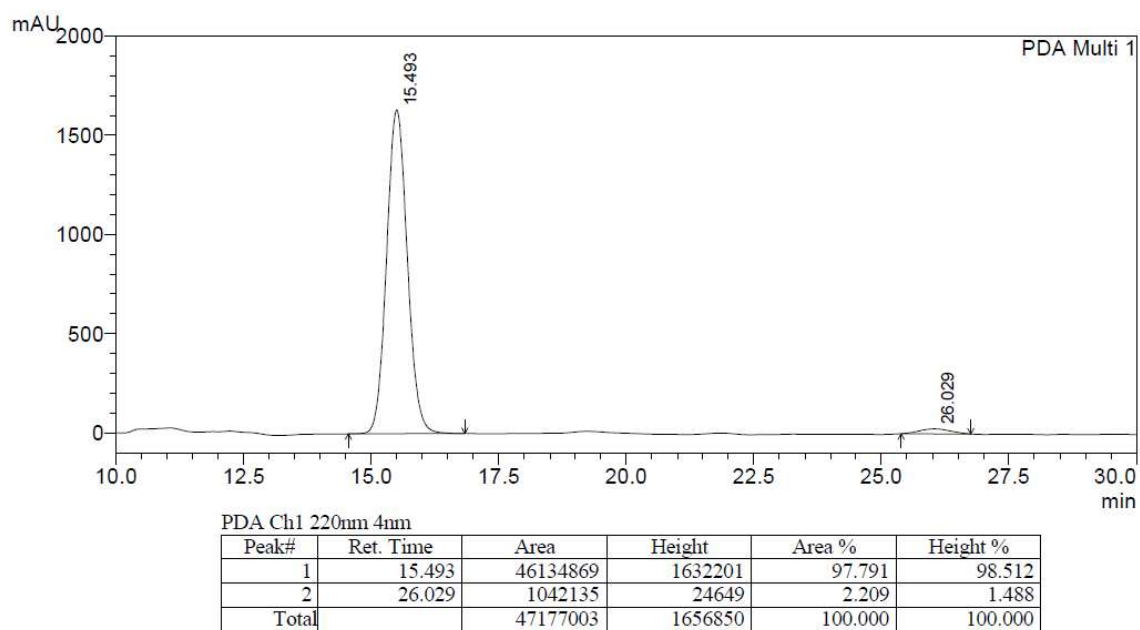
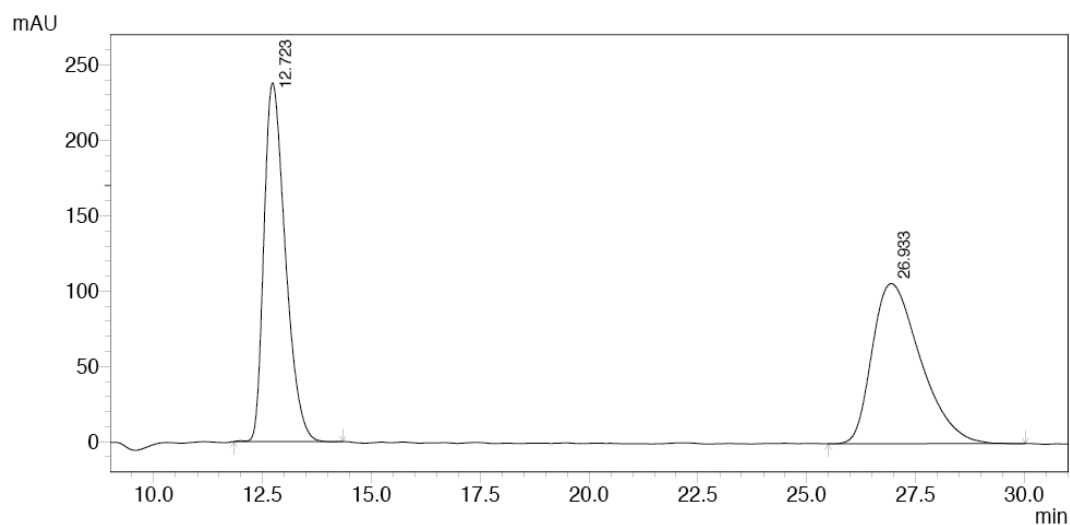


Figure s14. HPLC traces of **6ic** (Scheme 4, entry 9): (a) racemic sample; (b) catalyzed by Λ -(*S,S*)- $\mathbf{3}^{3+}$ $2\text{BF}_4^- \text{BAr}_f^-$; (c) catalyzed by Λ -(*S,S*)- $\mathbf{3}^{3+}$ 3BF_4^- .

Dimethyl 2-(2-nitro-1-(2-benzyloxyphenyl)ethyl)malonate (6jc).

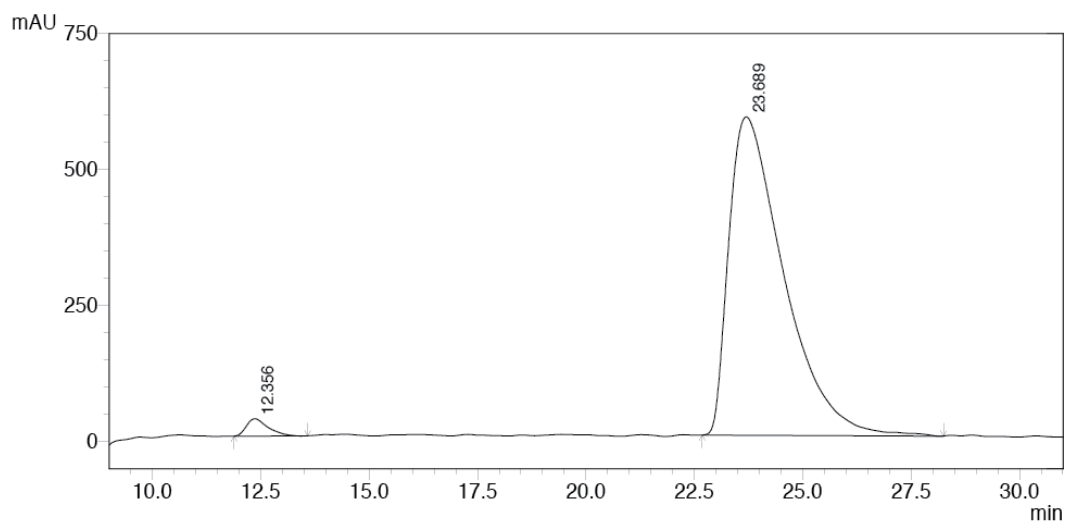
(a)



PDA 220nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 12.723 | 8141200 | 238008 | 49.819 | 69.153 |
| 2 | 26.933 | 8200292 | 106170 | 50.181 | 30.847 |
| Total | | 16341493 | 344178 | 100.000 | 100.000 |

(b)



PDA 220nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 12.356 | 1090049 | 31778 | 2.068 | 5.151 |
| 2 | 23.689 | 51627061 | 585142 | 97.932 | 94.849 |
| Total | | 52717109 | 616920 | 100.000 | 100.000 |

(c)

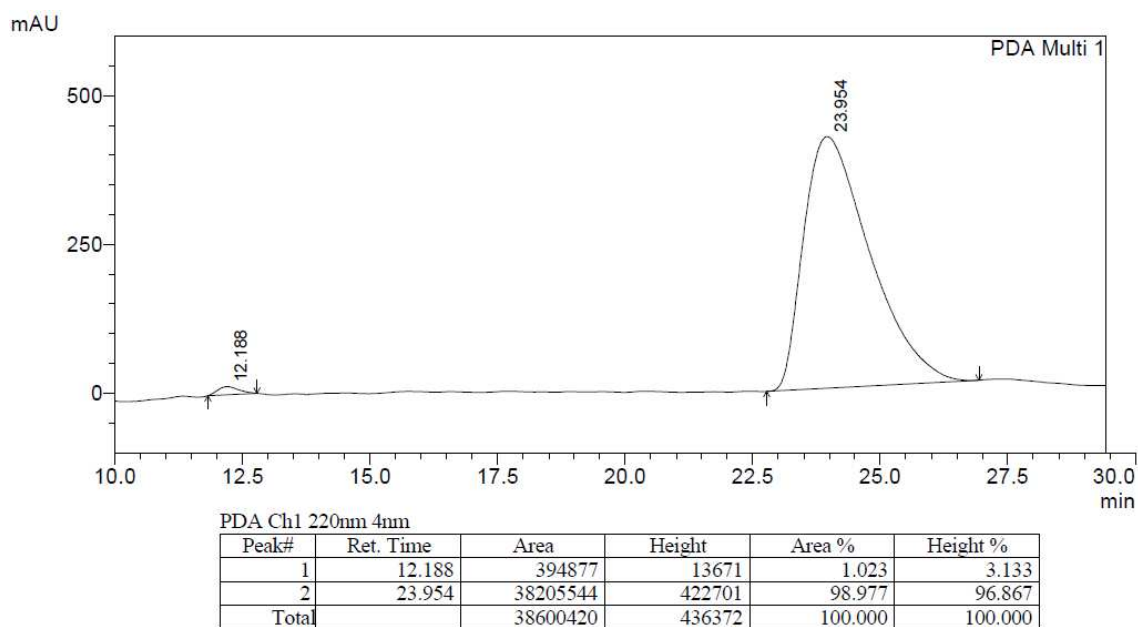
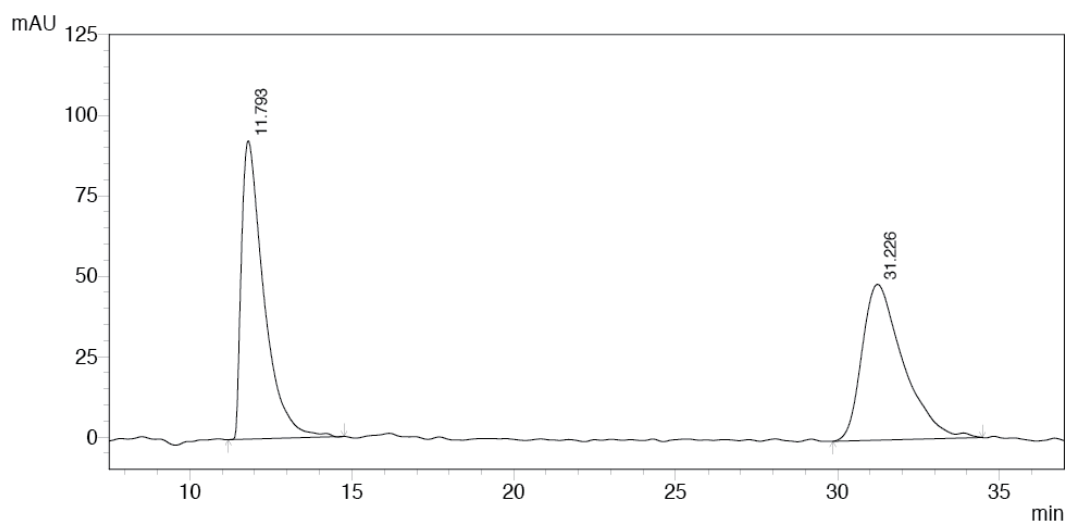


Figure s15. HPLC traces of **6jc** (Scheme 4, entry 10): (a) racemic sample; (b) catalyzed by Λ -(*S,S*)- $\mathbf{3}^{3+}$ $2\text{BF}_4^- \text{BARf}^-$; (c) catalyzed by Λ -(*S,S*)- $\mathbf{3}^{3+}$ 3BF_4^- .

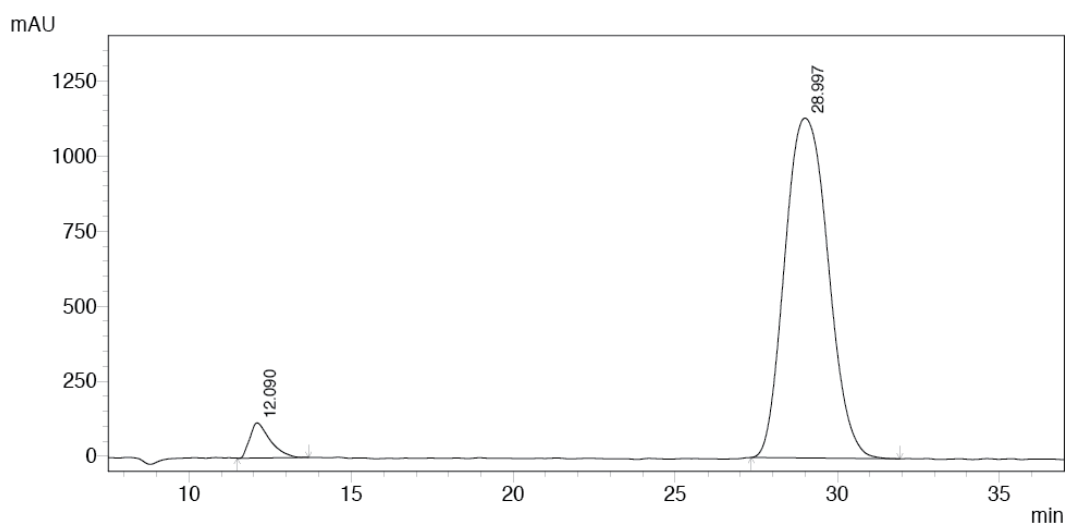
Dimethyl 2-(2-nitro-1-furyl)ethylmalonate (6kc).

(a)



| PDA 220nm | | | | | |
|-----------|-----------|---------|--------|---------|----------|
| Peak# | Ret. Time | Area | Height | Area % | Height % |
| 1 | 11.793 | 4468208 | 92606 | 51.035 | 65.690 |
| 2 | 31.226 | 4287008 | 48367 | 48.965 | 34.310 |
| Total | | 8755216 | 140974 | 100.000 | 100.000 |

(b)



| PDA 220nm | | | | | |
|-----------|-----------|-----------|---------|---------|----------|
| Peak# | Ret. Time | Area | Height | Area % | Height % |
| 1 | 12.090 | 5063370 | 116519 | 4.580 | 9.340 |
| 2 | 28.997 | 105482048 | 1131049 | 95.420 | 90.660 |
| Total | | 110545419 | 1247568 | 100.000 | 100.000 |

(c)

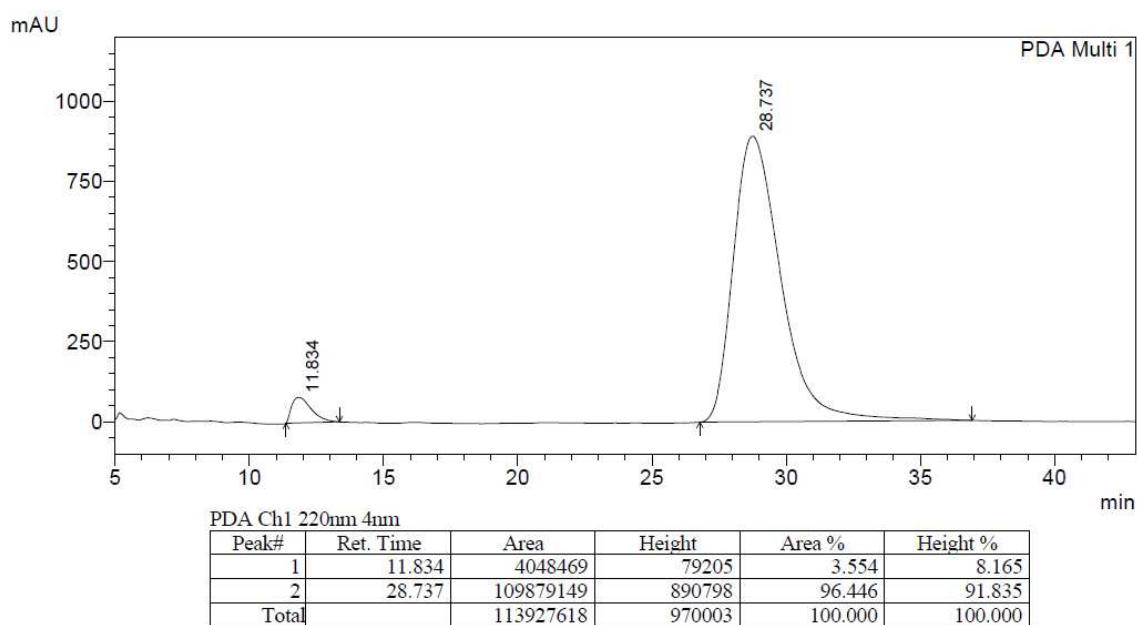
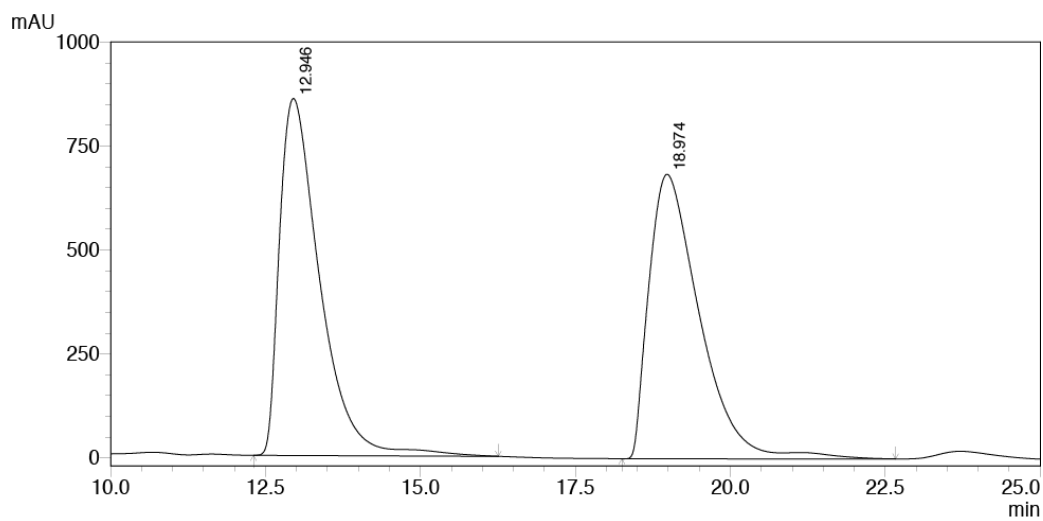


Figure s16. HPLC traces of **6k_c** (Scheme 4, entry 11): (a) racemic sample; (b) catalyzed by Λ -(*S,S*)-**3**³⁺ 2BF₄⁻BAr_f⁻; (c) catalyzed by Λ -(*S,S*)-**3**³⁺ 3BF₄⁻.

Dimethyl 2-(2-nitro-1-propylethyl)malonate (6lc).

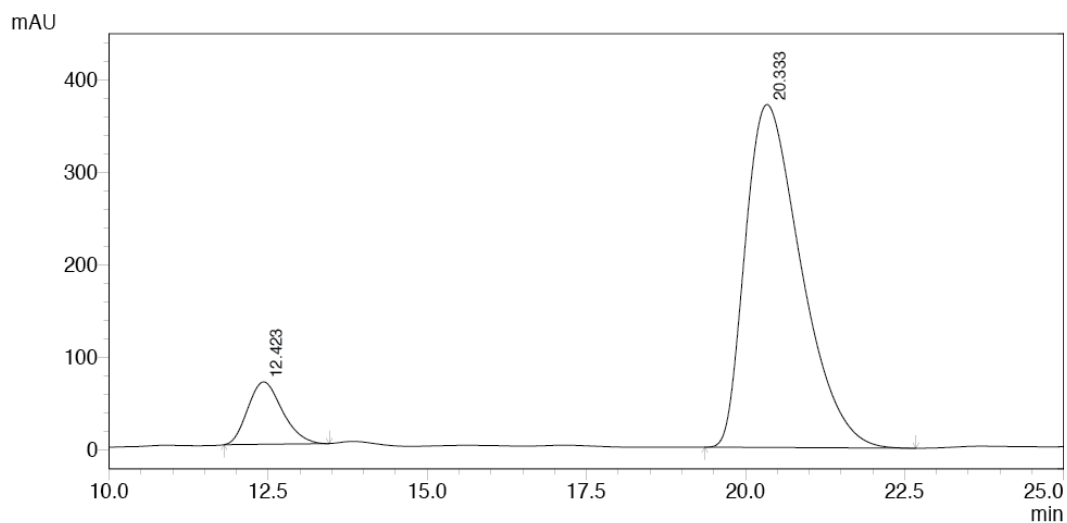
(a)



PDA 220nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|---------|---------|----------|
| 1 | 12.946 | 38973226 | 858914 | 49.978 | 55.647 |
| 2 | 18.974 | 39007602 | 684601 | 50.022 | 44.353 |
| Total | | 77980828 | 1543515 | 100.000 | 100.000 |

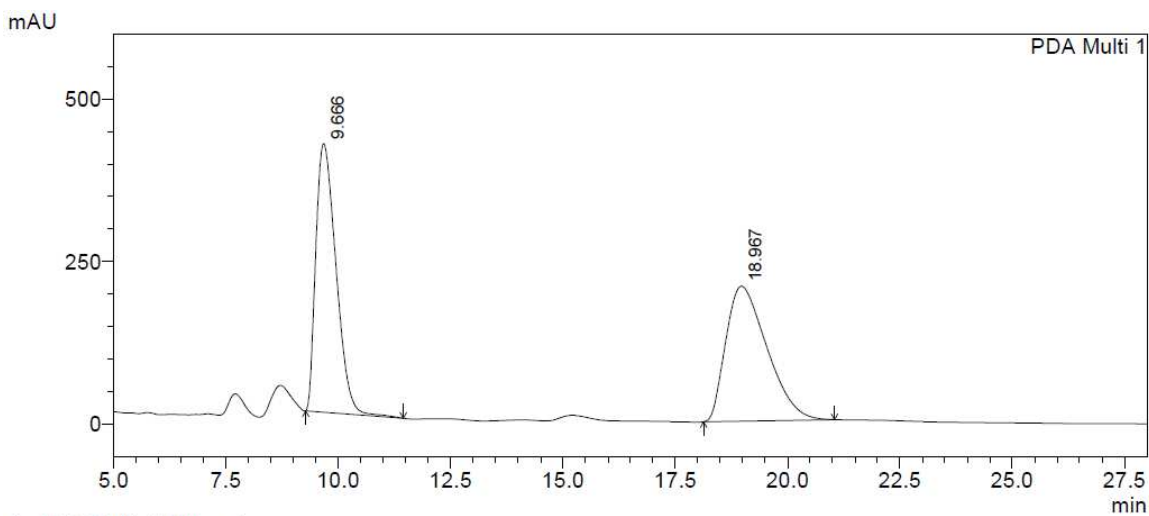
(b)



PDA 220nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 12.423 | 2538858 | 67307 | 10.100 | 15.379 |
| 2 | 20.333 | 22598513 | 370346 | 89.900 | 84.621 |
| Total | | 25137371 | 437652 | 100.000 | 100.000 |

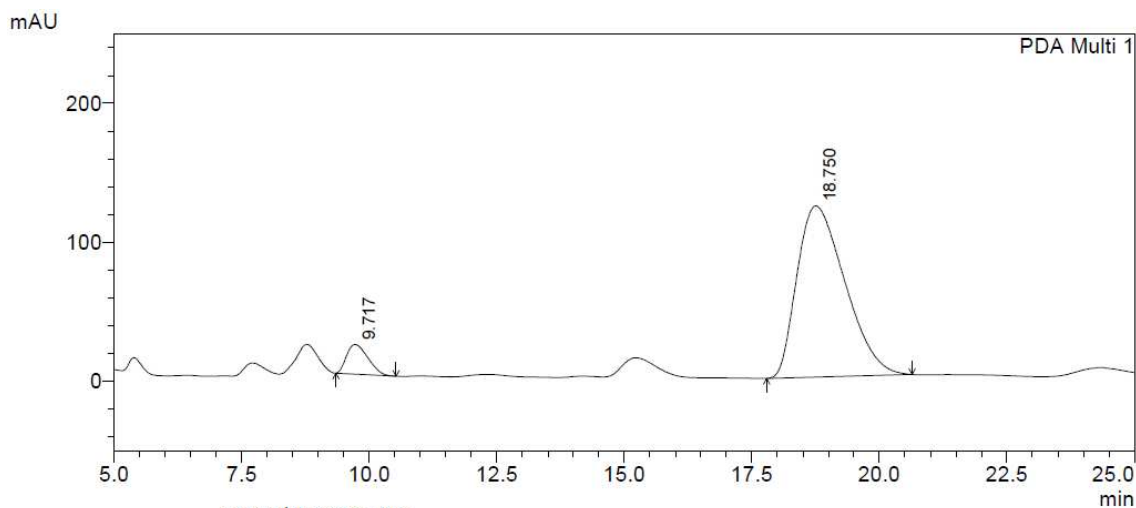
(c)



PeakTable

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 9.666 | 12941031 | 413408 | 49.424 | 66.552 |
| 2 | 18.967 | 13242511 | 207771 | 50.576 | 33.448 |
| Total | | 26183542 | 621179 | 100.000 | 100.000 |

(d)



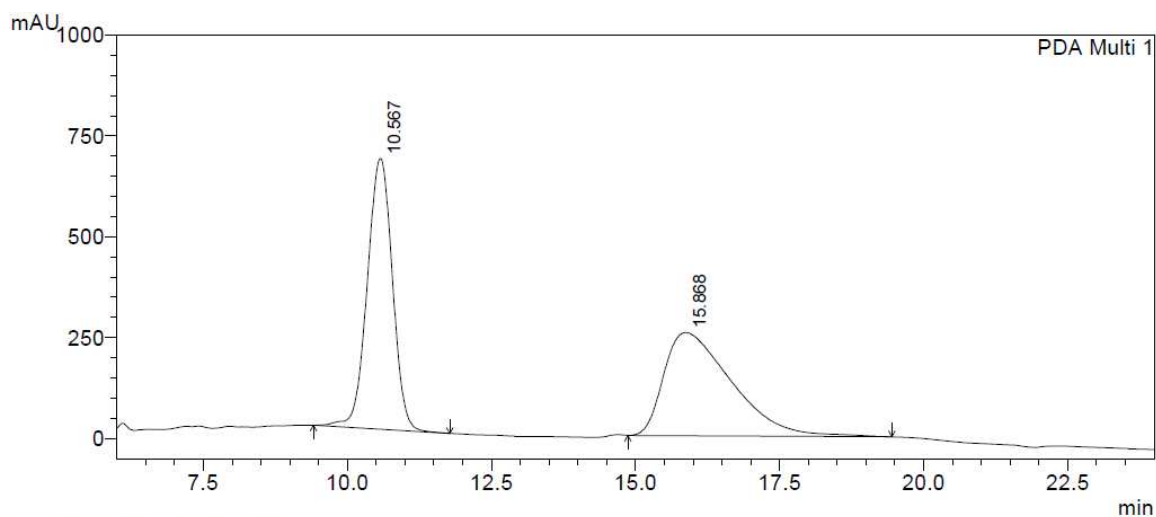
PDA Ch1 220nm 4nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|---------|--------|---------|----------|
| 1 | 9.717 | 622512 | 21401 | 7.162 | 14.797 |
| 2 | 18.750 | 8069654 | 123232 | 92.838 | 85.203 |
| Total | | 8692166 | 144634 | 100.000 | 100.000 |

Figure s17. HPLC traces of **6lc** (Scheme 4, entry 12): (a) racemic sample; (b) catalyzed by Λ -(*S,S*)-**3**³⁺ 2BF₄⁻BARf⁻ and analyzed per a; (c) racemic sample six months later (older column); (d) catalyzed by Λ -(*S,S*)-**3**³⁺ 3BF₄⁻ and analyzed per c.

***N,N'*-Bis(*t*-butoxycarbonyl)-1-hydrazino-2-oxocyclopentanecarboxylic acid ethyl ester (9a).**

(a)



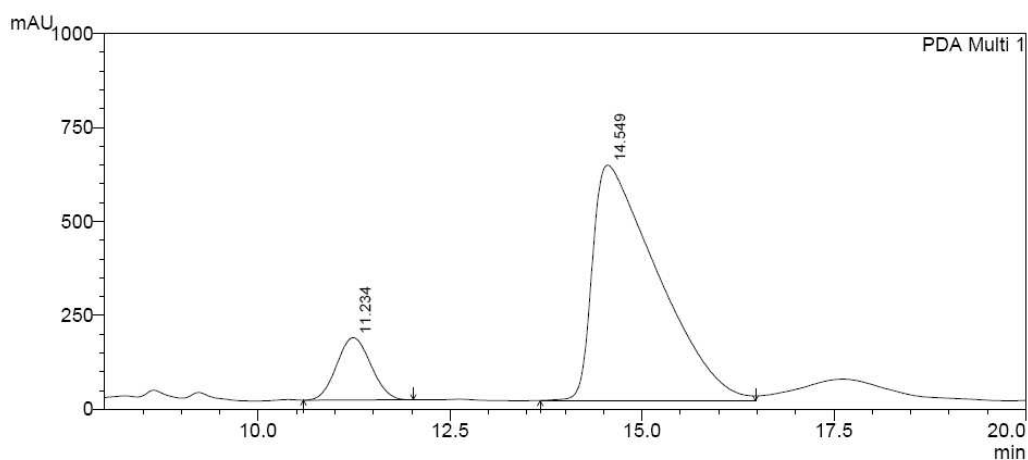
1 PDA Multi 1/210nm 4nm

PeakTable

PDA Ch1 210nm 4nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 10.567 | 20407922 | 671133 | 49.435 | 72.401 |
| 2 | 15.868 | 20874427 | 255836 | 50.565 | 27.599 |
| Total | | 41282348 | 926970 | 100.000 | 100.000 |

(b)



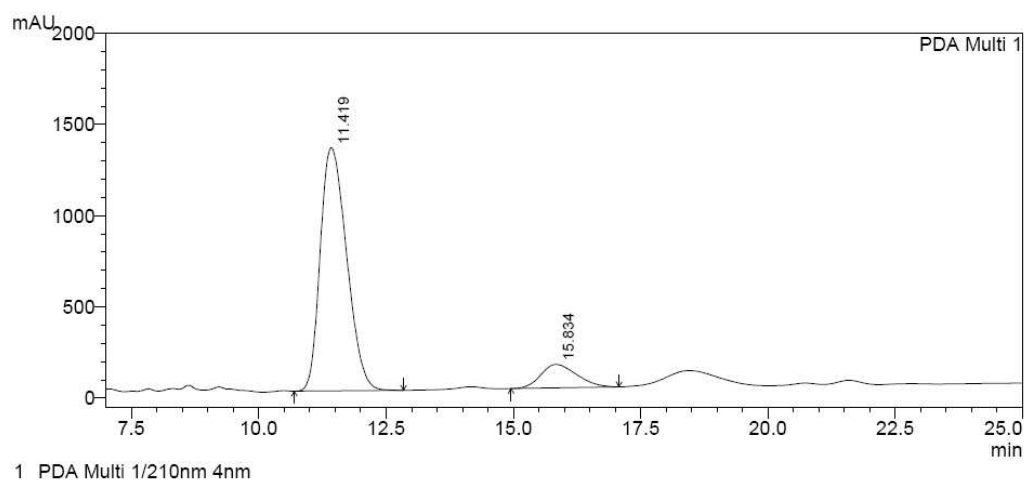
1 PDA Multi 1/210nm 4nm

PeakTable

PDA Ch1 210nm 4nm

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|--------|---------|----------|
| 1 | 11.234 | 5140005 | 165769 | 12.153 | 20.925 |
| 2 | 14.549 | 37155368 | 626417 | 87.847 | 79.075 |
| Total | | 42295373 | 792186 | 100.000 | 100.000 |

(c)



1 PDA Multi 1/210nm 4nm

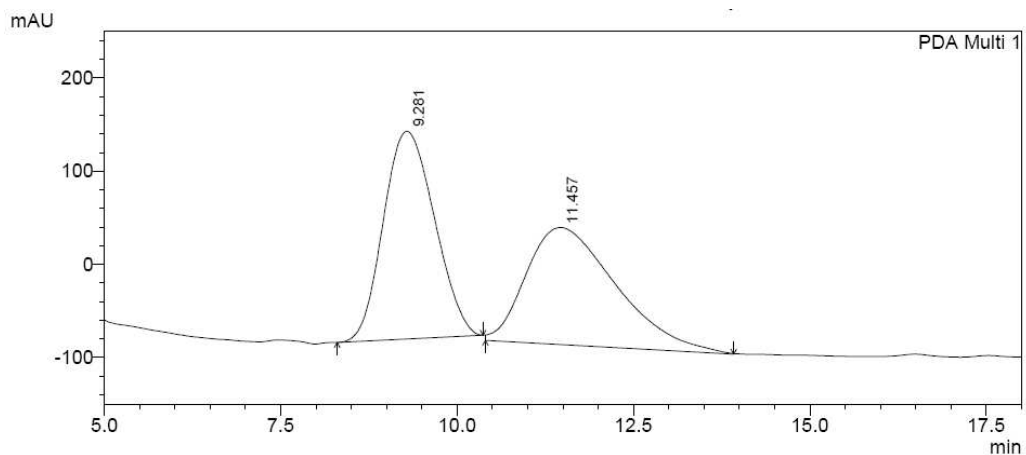
PeakTable

| Peak# | Ret. Time | Area | Height | Area % | Height % |
|-------|-----------|----------|---------|---------|----------|
| 1 | 11.419 | 48272755 | 1334808 | 88.385 | 91.220 |
| 2 | 15.834 | 6343733 | 128482 | 11.615 | 8.780 |
| Total | | 54616488 | 1463290 | 100.000 | 100.000 |

Figure s19. HPLC traces of **9a** (Scheme 5): (a) racemic sample; (b) catalyzed by Δ -(*S,S*)-**3**³⁺ 2Cl⁻BAr_f⁻; (c) catalyzed by Δ -(*S,S*)-**3**³⁺ 2Cl⁻BAr_f⁻.

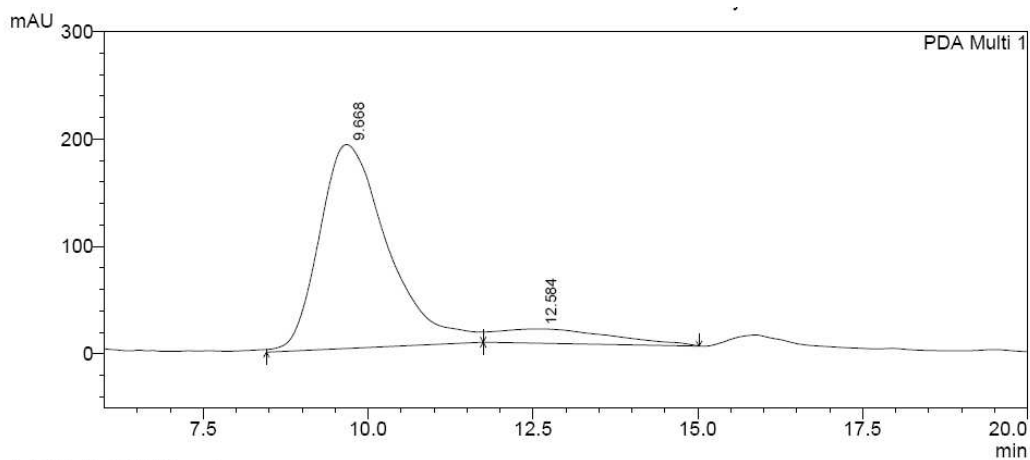
***N,N'*-Bis(*t*-butoxycarbonyl)-1-hydrazino-2-oxocyclohexanecarboxylic acid ethyl ester (9b).**

(a)



| PeakTable | | | | | |
|-------------------|-----------|----------|--------|---------|----------|
| PDA Ch1 210nm 4nm | | | | | |
| Peak# | Ret. Time | Area | Height | Area % | Height % |
| 1 | 9.281 | 11226181 | 222795 | 50.279 | 63.974 |
| 2 | 11.457 | 11101394 | 125466 | 49.721 | 36.026 |
| Total | | 22327575 | 348261 | 100.000 | 100.000 |

(b)



| PeakTable | | | | | |
|-------------------|-----------|----------|--------|---------|----------|
| PDA Ch1 210nm 4nm | | | | | |
| Peak# | Ret. Time | Area | Height | Area % | Height % |
| 1 | 9.668 | 13704382 | 189846 | 89.137 | 93.465 |
| 2 | 12.584 | 1670090 | 13274 | 10.863 | 6.535 |
| Total | | 15374472 | 203120 | 100.000 | 100.000 |

(c)

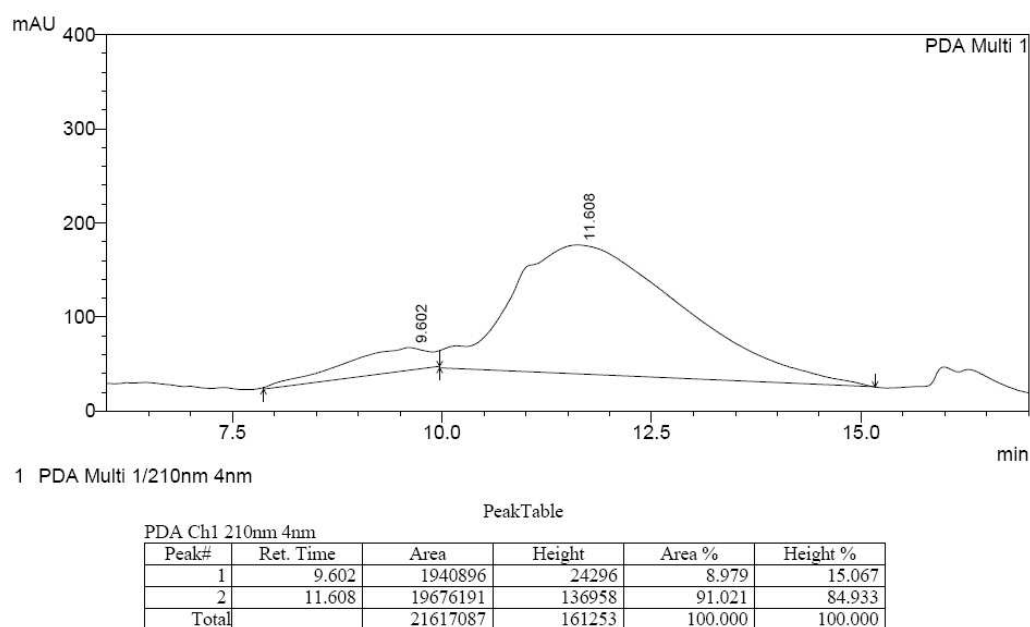


Figure s19. HPLC traces of **9b** (Scheme 5): (a) racemic sample; (b) catalyzed by Λ -(*S,S*)-**3**³⁺ 2Cl⁻BAr_f⁻; (c) catalyzed by Δ -(*S,S*)-**3**³⁺ 2Cl⁻BAr_f⁻.